

**Synbiotic (*Lactiplantibacillus pentosus* GSSK2 and isomalto-oligosaccharides)
supplementation modulates pathophysiology and gut dysbiosis in experimental metabolic
syndrome**

Sakshi Khanna¹, Mahendra Bishnoi², Kanthi Kiran Kondepudi^{2*}, Geeta Shukla^{1*}

¹Department of Microbiology, Panjab University, Chandigarh, India

²Healthy Gut Research Group, National Agri-Food Biotechnology Institute (NABI), S.A.S. Nagar, Punjab, India

*Corresponding author

Professor Geeta Shukla

Department of Microbiology, Basic Medical Sciences Block A, South Campus, Panjab University, Chandigarh, 160014, India. Email: geeta_shukla@pu.ac.in
ORCID: 0000000212122211

Dr. Kanthi Kiran Kondepudi

Healthy Gut Research Group, Food & Nutrition Biotechnology Division, National Agri-Food Biotechnology Institute (NABI), S.A.S. Nagar, Punjab 140306, India. Email: kiran@nabi.res.in

Supplementary Information

Supplementary Table S1: List of bacterial primers for q-PCR analysis of gut bacterial abundance

Bacterial primers	Forward 5'-3'	Reverse 5'- 3'
Total Bacteria	ACTCCTACGGGAGGCAGCAGT	ATTACCGCGGCTGCTGGC
<i>Bacteroidetes</i>	ACGCTAGCTACAGGCTTAACA	ACGCTACTTGGCTGGTTCA
<i>Firmicutes</i>	GCGTGAGTGAAGAAGT	CTACGCTCCCTTTACAC
<i>Lactobacillus</i>	CACCGCTACACATGGAG	AGCAGTAGGGAATCTTCCA
<i>Bifidobacterium</i>	TCGCGTCYGGTGTGAAAG	CCACATCCAGCRTCCAC
<i>Akkermensia</i>	CAGCACGTGAAGGTGGGGAC	CCTTGCGGTTGGCTTCAGAT
<i>Faecalibacterium</i>	GAGGAAGATAATGACGGTAC	ACCTCTGCACTACTCAAGA
<i>Roseburia</i>	GCGGTRCGGCAAGTCTGA	CCTCCGACACTCTAGTMCGA
<i>Ruminococcus</i>	CCCTGCGCTGTGCGAAAAAG	CCGCTGCGTACCTTTGGGAT
<i>Prevotella</i>	GATGGGGATGCGTCTGATTAG	TCCTGCACGCTACTTGGCT
<i>Enterobacteriaceae</i>	CCCTTATTGTTAGTTGCCAT	ACTCGTTGTACTTCCCATTG

Primer sequences were obtained from literature¹³ and were procured from Eurofin Genomics India Pvt. Ltd.

PCR conditions for gut bacteria abundance : q-PCR was performed in 10µl reaction volume containing 1 µl template DNA (40 ng), 1 µl forward primer, 1 µl reverse primer, 2µl RNase free water and 5 µl iTaq Universal SYBR Green Supermix (2X) and was carried out using following conditions: 95°C for 2 minutes followed by 40 cycles of 95°C for 5 s, 60°C for 30 s with melt curve at 65-95°C for 5 s.

Supplementary Table S2: List of primers for q-PCR analysis of gene expression in tissues.

Gene	Forward (5'-3')	Reverse (5'-3')	Source
GAPDH	ACGGGAAACCCATCACCATC	CTCGTGGTTCACACCCATCA	*
Adiponectin	GGAAACTTGTGCAGGTTGGATG	GGGTCACCCCTTAGGACCAAGAA	[1]
Leptin	TTCAAGCTGTGCCTATCCACAAAG	TGAAGCCCGGGAATGAAGTC	[1]
TLR-4	TGGCAGTTTCTGAGTAGCCG	GCTTTTCCATCCAACAGGGC	[2]
Claudin	TGTCCACCATTGGCATGAAG	GCCACTAATGTCGCCAGACC	[3]
CDX-2	CGCATCATCACCCCTCACCAT	CGTCCTGGTTTTCACTTGGC	*
Muc-2	GCCAGATCCCGAAACCATGT	AGGACGGACTCTATGCTGGA	[2]
GLUT-4	CAACTGGACCTGTAACCTTCATCGT	ACGGCAAATAGAAGGAAGACGTA	[4]
Glucokinase	CAGTGGAGCGTGAAGACAAA	CTTGGTCCAATTG AGGAGGA	*
CEBP-α	TTACAACAGGCCAGGTTTCC	GGCTGGCGACATACAGTACA	[5]
PPAR-γ	GACCACTCCCATTCTTTGA	CGCACTTTGGTATTCTTGGAG	[6]
IL-6	TCCTACCCCAACTTCCAATGCTC	TTGGATGGTCTTGGTCCTTAGCC	[7]
TNF-α	GTCGTAGCAAACCACCAAGC	TGTGGGTGAGGAGCACATAG	[8]
FASN	GCAGCAGCATGATGTAGCAC	AGTTGCACACCACAAGGTCA	*
HSL	CGCCTTACGGAGTCTATGC	TCTGATGGCTCTGAGTTGC	[5]

*Primer sequences were designed using Primer BLAST software and were procured from Eurofin Genomics India Pvt. Ltd.

PCR conditions for gene expression analysis in tissues: q-PCR was performed in 10 μ l reaction volume containing 1 μ l template cDNA (20 ng), 1 μ l forward primer, 1 μ l reverse primer, 1 μ l RNase free water and 5 μ l iTaq Universal SYBR Green Supermix (2X). qPCR was carried out under the following conditions: 95°C for 2 minutes followed by 40 cycles of 95°C for 5 s, 60°C for 30 s with melt curve at 65-95°C for 5 s.

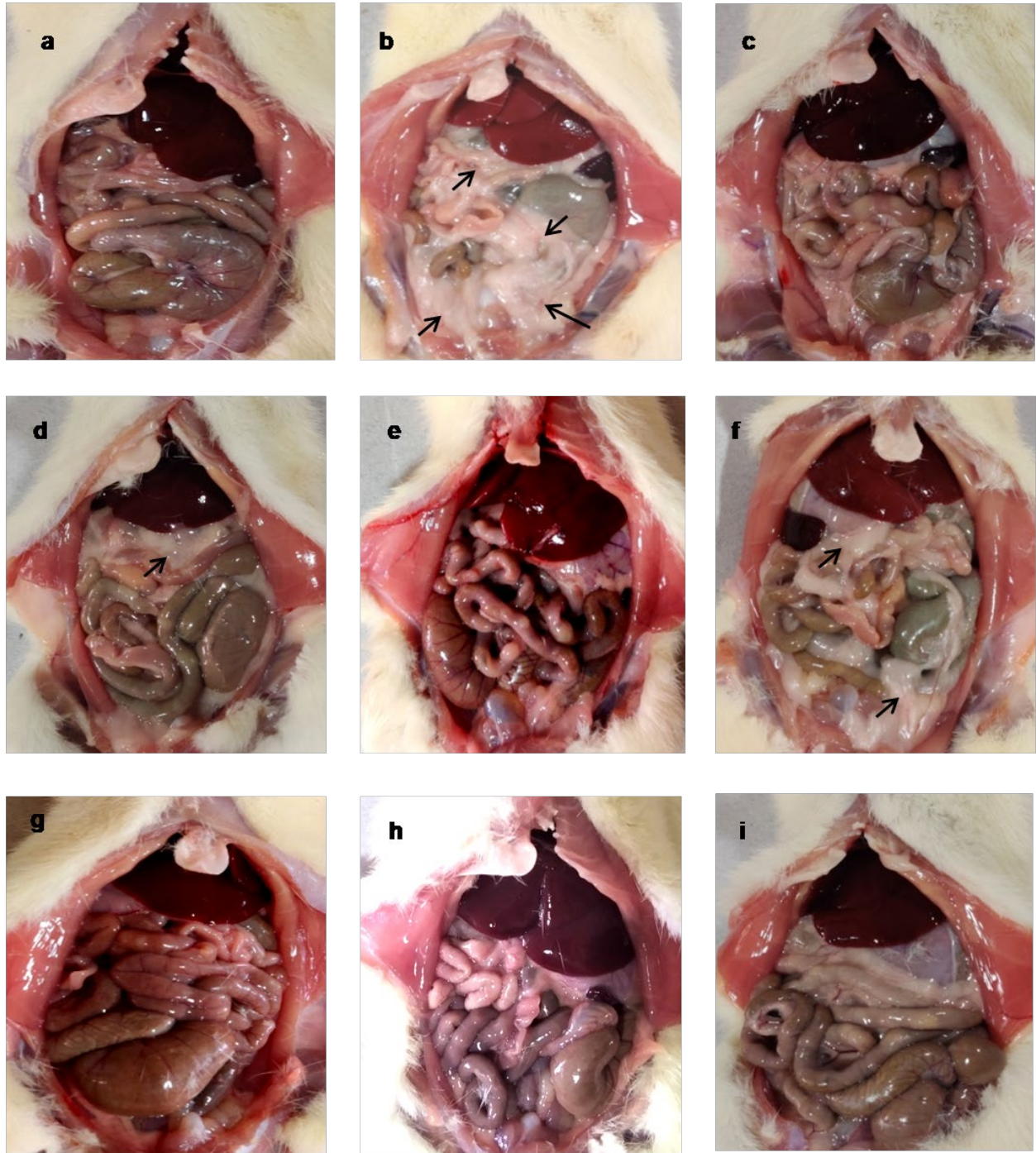


Figure S1: Gross macroscopic observation of adipose tissue of animals belonging to different groups: (a) Control, (b) HFD, (c) *L. pentosus*, (d) *L. pentosus* + HFD, (e) IMOs, (f) IMOs + HFD, (g) Synbiotic, (h) Synbiotic + HFD, (i) Orlistat + HFD. Arrows indicate adipose tissue deposits.

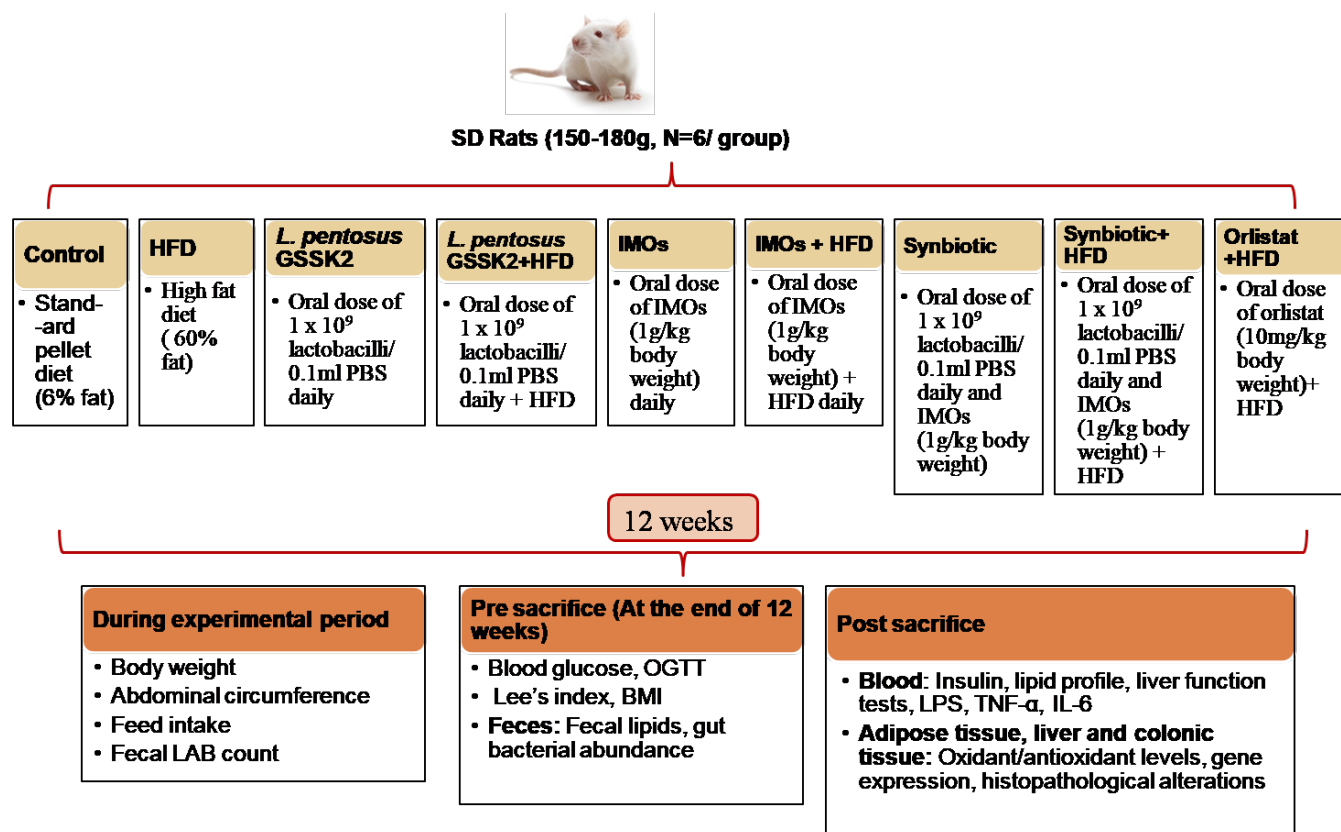


Figure S2: Flow diagram depicting different experimental groups and interventions.

References

1. Abd El-Haleim, E.A., Bahgat, A.K., Saleh, S. Resveratrol and fenofibrate ameliorate fructose-induced nonalcoholic steatohepatitis by modulation of genes expression. *World J Gastroenterol* **22**, 2931-2948 (2016)
2. Kumar, V., Mahajan, N., Khare, P., Kondepudi, K.K., Bishnoi, M. Role of TRPV1 in colonic mucin production and gut microbiota profile. *Eur J Pharmacol.* **888**, 173567; 10.1016/j.ejphar.2020.173567 (2020).
3. Takizawa, Y., Kishimoto, H., Kitazato, T., Tomita, M., Hayashi, M. Changes in protein and mRNA expression levels of claudin family after mucosal lesion by intestinal ischemia/reperfusion. *Int J Pharm* **426**, 82-89 (2012).
4. Qin, B., Polansky, M. M., Harry, D., Anderson, R. A. Green tea polyphenols improve cardiac muscle mRNA and protein levels of signal pathways related to insulin and lipid

- metabolism and inflammation in insulin-resistant rats. *Mol Nutr Food Res*. **54**, S14-23. (2010)
5. Li, X. et al. Fructus xanthii improves lipid homeostasis in the epididymal adipose tissue of rats fed a high-fat diet. *Mol Med Rep* **13**, 787-795 (2016).
 6. Alonso, M. et al. Anti-obesity efficacy of LH-21, a cannabinoid CB(1) receptor antagonist with poor brain penetration, in diet-induced obese rats. *Br J Pharmacol* **165**, 2274-2291 (2012).
 7. Parafati, M. et al. Bergamot Polyphenols Boost Therapeutic Effects of the Diet on Non-Alcoholic Steatohepatitis (NASH) Induced by "Junk Food": Evidence for Anti-Inflammatory Activity. *Nutrients* **10**, 1604; 10.3390/nu10111604 (2018).
 8. Agarwal, D., Dange, R. B., Vila, J., Otamendi, A. J., Francis, J. Detraining differentially preserved beneficial effects of exercise on hypertension: effects on blood pressure, cardiac function, brain inflammatory cytokines and oxidative stress. *PLoS One* **7**, e52569; 10.1371/journal.pone.0052569 (2012).