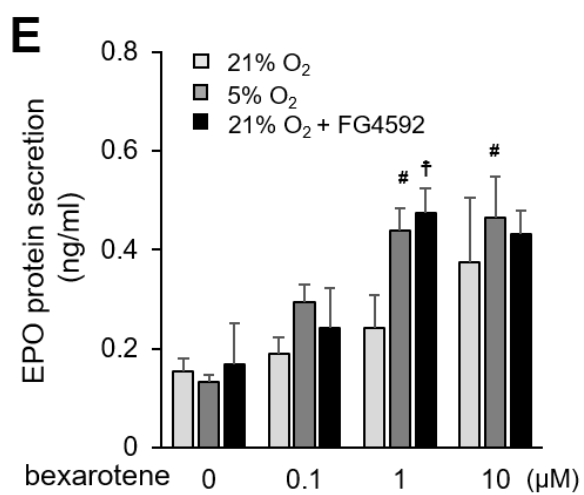
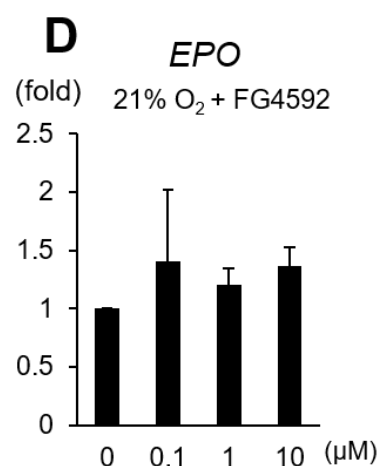
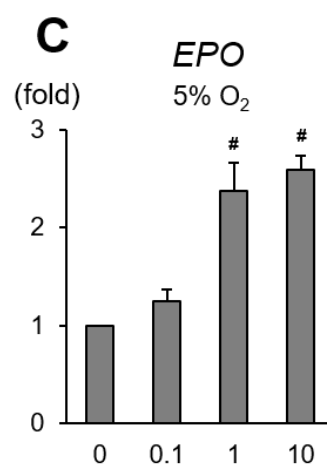
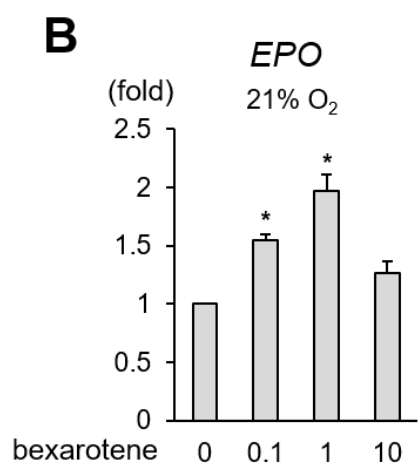
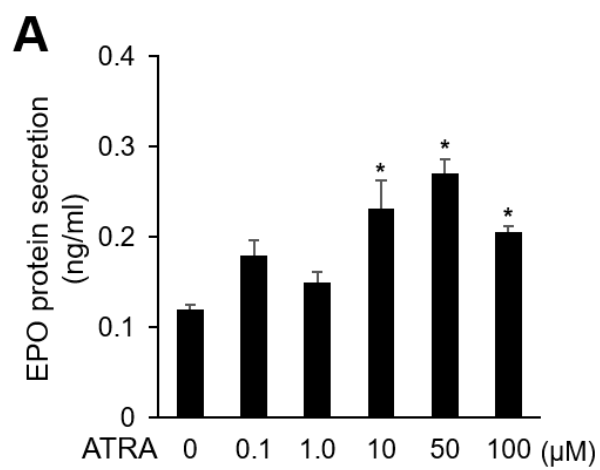


Supplementary Information

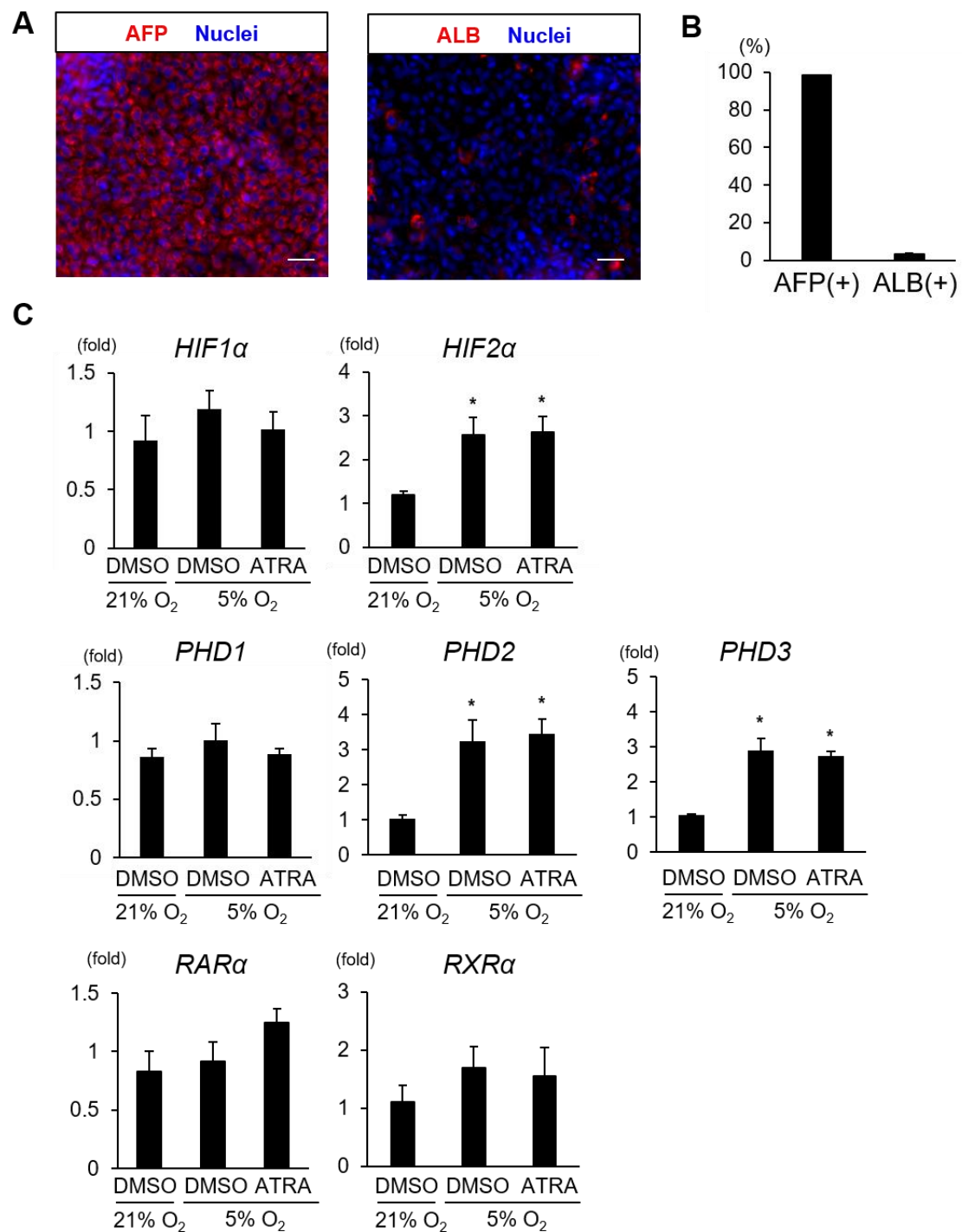
Retinoic acid regulates erythropoietin production cooperatively with hypoxia-inducible factors in human iPSC-derived erythropoietin-producing cells

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Supplementary Fig. 1. Effects of RA and hypoxic signals on EPO production by hiPSC-EPO cells.

(**A**) Effects of various concentrations of ATRA on EPO protein secretion by hiPSC-EPO cells under normoxic conditions (21% oxygen). (**B - E**) Effects of bexarotene treatment on EPO mRNA expression (**B - D**) and protein secretion (**E**) by hiPSC-EPO cells under normoxia (21% oxygen; **B, E**, light gray), hypoxia (5% oxygen; **C, E**, dark gray), and normoxia with PHD inhibitor treatment (10 μ M FG4592: **D, E**, black), as analyzed by qRT-PCR and ELISA, respectively. Note that the qRT-PCR analysis in (**B - D**) were performed independently. The data from four independent experiments (n=5 for **A** and n=4 for **E**) or three independent experiments (n=3 for **B - D**) are represented as the means $\pm$ SEM in (**A - E**). Statistical analysis was performed using one-way ANOVA with Dunnett's test. *p<0.05 versus the samples treated with DMSO under normoxic conditions in (**A, B**). #p<0.05 versus the samples treated with DMSO under hypoxic conditions in (**C, E**). †p<0.05 versus the samples treated with FG4592 but without bexarotene under normoxic conditions in (**E**).



Supplementary Fig. 2. Effects of RA and HIF signals on EPO production by hiPSC-EPO cells.

(**A**) Immunostaining analysis of hiPSC-EPO cells under normoxic conditions (21% oxygen) for AFP and ALB. Scale bars, 50 μ m. (**B**) Percentage of AFP(+) or ALB(+) cells in (**A**). (**C**) qRT-PCR analysis of the expressions of *HIF1 α* , *HIF2 α* , *PHD1*, *PHD2*, *PHD3*, *RAR α* and *RXR α* by hiPSC-EPO cells treated with DMSO under normoxic or hypoxic conditions (5% oxygen) or with ATRA under hypoxic conditions. The data from three independent experiments (n=4 for **B** and n=3 for **C**) are represented as the means $\pm$ SEM in (**B**, **C**). *p<0.05 versus the samples treated with DMSO under normoxic conditions by one-way ANOVA with Bonferroni's test.



Supplementary Figure 3. Full unedited gel for Figure 1A.

Supplementary Table 1. The chemical compounds used in this study.

Chemical compound	Function	Company
all-trans-retinoic acid (ATRA)	RAR agonist	Sigma-Aldrich
Bexarotene	RXR agonist	R&D
AGN193109	RAR antagonist	SANTA CRUZ
FG4592 (roxadustat)	PHD inhibitor	Cayman Chemical
Molidustat	PHD inhibitor	Cayman Chemical
Daprodustat	PHD inhibitor	Selleck Biotech
deferrioxamine (DFO)	PHD inhibitor	Sigma-Aldrich
dimethyloxalylglycine (DMOG)	PHD inhibitor	Sigma-Aldrich

Supplementary Table 2. The primer sequences used in this study.

Gene	Orientation	Primer sequence (5'-3')
<i>EPO</i>	Sense	TGTGGATAAAGCCGTCAGTG
	Antisense	GATTGTTCGGAGTGGAGCAG
<i>Epo</i>	Sense	TTGTGCAGAAGGTCCCAGAC
	Antisense	GCAGTGAAGTGAGGCTACGT
β - <i>ACTIN</i>	Sense	CAATGTGGCCGAGGACTTTG
	Antisense	CATTCTCCTTAGAGAGAAGTGG
<i>GAPDH</i>	Sense	GAAGGTGAAGGTCGGAGTC
	Antisense	GAAGATGGTGATGGGATTTC
<i>Gapdh</i>	Sense	AACTTTGGCATTGTGGAAGG
	Antisense	GGATGCAGGGATGAGTTCT
<i>RARα</i>	Sense	ATGTCCAAGGAGTCTGTGAG
	Antisense	TGTCCCAGAGGTCAATGTC
<i>RARβ</i>	Sense	GGTTTCACTGGCTTGACCAT
	Antisense	GGCAAAGGTGAACACAAGGT
<i>RARγ</i>	Sense	GGCTGTGGGACAAGTTCAGT
	Antisense	GTCGGAGAAGGTCATGGTGT
<i>RXRα</i>	Sense	TTCTCCACCCAGGTGAACTC
	Antisense	GAGCTGATGACCGAGAAAGG
<i>RXRβ</i>	Sense	TCCCACACTTTTCCTCCTTG
	Antisense	ATGGCTCCTACTCCTGCTGA
<i>RXRγ</i>	Sense	TCGTCTGGGCAGATTATTCC
	Antisense	AGGGCTTGATGTCCTCTGAA

<i>DLK1</i>	Sense	TGGCTTCTCAGGCAATTCT
	Antisense	CTGCAGGTCTTGTCGATGAA
<i>AFP</i>	Sense	AAATGCGTTTCTCGTTGCTT
	Antisense	GCCACAGGCCAATAGTTTGT
<i>ALB</i>	Sense	CGCTATTAGTTCGTTACACCA
	Antisense	TTTACAACATTTGCTGCCCA
<i>HNF4A</i>	Sense	TGCGACTCTCCAAAACCCTC
	Antisense	ATTGCCCATCGTCAACACCT
<i>SALL4</i>	Sense	CAGATCCACGAGCGGACTCA
	Antisense	CCCCGTGTGTCATGTAGTGA
<i>HNF1B</i>	Sense	CCTCTCCTCCAAACAAGCTG
	Antisense	TGTTGCCATGGTGACTGATT
<i>GATA4</i>	Sense	GGAAGCCCAAGAACCTGAAT
	Antisense	GTTGCTGGAGTTGCTGGAA
<i>CK19</i>	Sense	CTCCCGCGACTACAGCCACT
	Antisense	TCAGCTCATCCAGCACCTG
<i>HIF1α</i>	Sense	CCATTAGAAAGCAGTTCCGC
	Antisense	TGGGTAGGAGATGGAGATGC
<i>HIF2α</i>	Sense	GAAGTCCCGGGATGCTGCGCG
	Antisense	ACTATGTCCTGTTAGCTCCAC
<i>PHD1</i>	Sense	AGCGGGCAGCAGCCAAAGACAAG
	Antisense	TGCCATGCGGCTCTGGGACTG
<i>PHD2</i>	Sense	AGCCCGGCTGCGAAACCATTG
	Antisense	TTCGTCCGGCCATTGATTTTGT

<i>PHD3</i>	Sense	AGATGTGGAGCCCATTTTG
	Antisense	CAGATTTCAGAGCACGGTCA
<i>RFX6</i>	Sense	TCAGCAGCATTCGTTCACTG
	Antisense	TGGGGTTTGCAAACCTGGAAG
<i>ARNT</i>	Sense	CAAGAACCACGGCCTACACT
	Antisense	TAGCTGGCCAGTCCATCTCT
<i>AHR</i>	Sense	AGTCTAATGCACGCCTGCTT
	Antisense	TGGCAGGAAAAGGGTTGGTT
<i>GATA3</i>	Sense	TCCTGTGCGAACTGTCAGAC
	Antisense	CGAGCTGTTCTTGGGGAAGT
<i>RBPJ</i>	Sense	AGTTACTGGCATGGCACTCC
	Antisense	TCCAGGAAGCGCCATCATTT
<i>AP-2α</i>	Sense	AAGAGTTCACCGACCTGCTG
	Antisense	AGGGCCTCGGTGAGATAGTT
<i>ZNF416</i>	Sense	ATGTGTCTTGCAGGAGCTCC
	Antisense	ATCAGCTTCCAGCAGAACCC
<i>GLIS3</i>	Sense	CAACGCCCCTATAAACTGC
	Antisense	CGGATGCTGGCACAAATACG
<i>EBF1</i>	Sense	CGCAACTCAAGCAGCGTATC
	Antisense	CCAAATTGGACATTGCGGCA
<i>SP2</i>	Sense	GCCCAATCTCACCAACCAGA
	Antisense	CACTGGCGTTCACAAGGTTG
<i>EBF2</i>	Sense	AGGAGACGATTTGCTGGCAA
	Antisense	CCTGACCGAATCCATCTCCG

<i>MYB</i>	Sense	ACAGATGGGCAGAAATCGCA
	Antisense	GCTGGCTGGCTTTTGAAGAC
<i>AP-2γ</i>	Sense	ACCCACACACTTAGCCATTG
	Antisense	AGCGGACACAAAAACCAACC