

Supplementary data to:

**rs1944919 on chromosome 11q23.1 and its effector genes
COLCA1/*COLCA2* confer susceptibility to primary biliary cholangitis**

Yuki Hitomi^{1,*}, Yoshihiro Aiba², Yosuke Kawai³, Kaname Kojima⁴, Kazuko Ueno³,
Nao Nishida^{3,5}, Minae Kawashima⁶, Olivier Gervais⁷, Seik-Soon Khor³, Masao
Nagasaki⁷, Katsushi Tokunaga³, Minoru Nakamura^{2,8,9}, Makoto Tsuiji^{1,*}

¹ Department of Microbiology, Hoshi University School of Pharmacy and
Pharmaceutical Sciences, Tokyo, Japan

² Clinical Research Center, National Hospital Organization (NHO) Nagasaki
Medical Center, Omura, Japan

³ Genome Medical Science Project, National Center for Global Health and
Medicine, Tokyo, Japan

⁴ Tohoku Medical Megabank Organization, Tohoku University, Sendai, Japan

⁵ The Research Center for Hepatitis and Immunology, National Center for Global
Health and Medicine, Ichikawa, Japan

⁶ Japan Science and Technology Agency (JST), Tokyo, Japan

⁷ Human Biosciences Unit for the Top Global Course Center for the Promotion of
Interdisciplinary Education and Research, Kyoto University, Kyoto, Japan

⁸ Department of Hepatology, Nagasaki University Graduate School of Biomedical
Sciences, Omura, Japan

⁹ Headquarters of PBC Research in NHO Study Group for Liver Disease in Japan

(NHOSLJ), Clinical Research Center, NHO Nagasaki Medical Center, Omura,
Japan

*Correspondence should be addressed to:

- Yuki Hitomi, PhD

Hoshi University School of Pharmacy and Pharmaceutical Sciences

2-4-41 Ebara, Shinagawa-ku, Tokyo 142-8501, Japan

Tel: +81-3-5498-5755; Fax: +81-3-5498-5755

E-mail: yhitomi-ty@umin.ac.jp

- Makoto Tsuiji, PhD

Hoshi University School of Pharmacy and Pharmaceutical Sciences

2-4-41 Ebara, Shinagawa-ku, Tokyo 142-8501, Japan

Tel: +81-3-5498-5902; Fax: +81-3-5498-5755

E-mail: m-tsuiji@hoshi.ac.jp

Table of contents

Supplementary Tables	3
Supplementary Figures	7

Supplementary Tables

Table S1. Oligo-nucleotide probes for the EMSA

SNP	Allele	Sequence (5' -> 3')
rs4938534	T	TCCTCTTTCCCGACGTCGGGTAGTTTCTTAA
	C	TCCTCTTTCCCGACGCCGGGTAGTTTCTTAA
rs7952497	C	CCATCCCCCTTCCCCCGTTACAAAAAGGACA
	A	CCATCCCCCTTCCCCAGTTACAAAAAGGACA
rs6589227	T	TCCTGTGCGAAGTGGTTCGGTCTAGAGGTGA
	C	TCCTGTGCGAAGTGGCTCGGTCTAGAGGTGA
rs1944919	T	TCGAGGTGGGTACTGTCTTTCAACGGGTCCT
	G	TCGAGGTGGGTACTGGCTTTCAACGGGTCCT
rs6589226	G	TTCAAGTGTTAACAAGGTAGGTTGAGGCCCC
	A	TTCAAGTGTTAACAAGTAGGTTGAGGCCCC
rs4356268	T	CCGATACGGGTGCCTTTTTCTCCGGTTGGGA
	C	CCGATACGGGTGCCTCTTTCTCCGGTTGGGA

Table S2. Primers for production of Luciferase assay constructs.

Primer Name	Sequence (5' -> 3')
rs1944919-F	GATATCTTTGCATGCATGCCATGCTC
rs1944919-R	AGATCTGAAAAACAGCAGTGCAGCTG
rs4938534-F	GGTACCTTCATGGTGTGCTTCACCTC
rs4938534-R	GAGCTCGCCTCGGGTATGTCTTTATC
rs7952497-F	GGTACCGCTTAAAACTGTGGAGGGAC
rs7952497-R	GAGCTCACATAGGAGCCCAATGAGAC
rs6589227-F	GGTACCCCTGTTTCTAGCTGAGTGAC
rs6589227-R	GAGCTCTTTCATCCAACTCCGGGGAT
rs6589226-F	GGTACCCTTCTGTGACAAGGCTTGTC
rs6589226-R	GAGCTCGGTTCAACTCTCAGAAGCTG
rs4356268-F	GGTACCGAGGCTCTGTTCATCTTGTG
rs4356268-R	GAGCTCTTGAAACACAGTAGCCCCTC

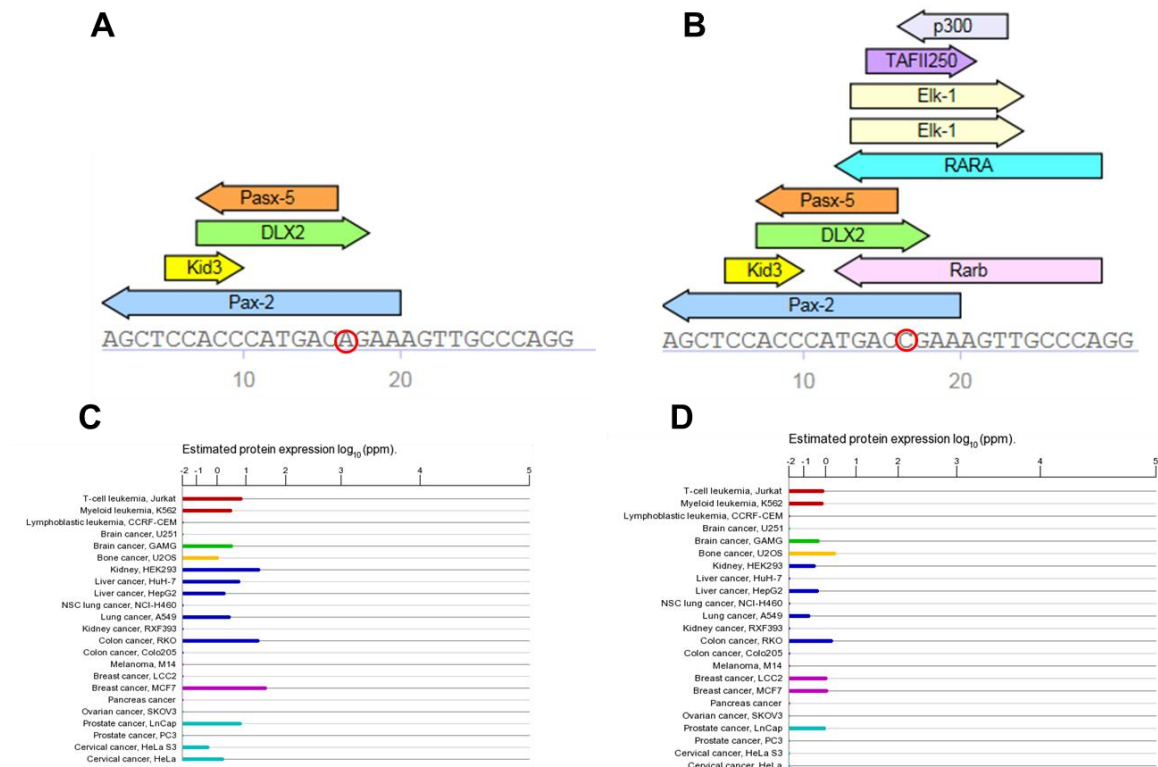
Table S3: Sequences of primers used for the synthesis of gRNAs, donor-DNA, and sequence check used for gene editing.

gRNA/donor DNA	Sequence
gRNA-1 primer-F	CCGGCCATGCTCCTGGGCAACTTT
gRNA-1 primer-R	AAACAAAGTTGCCCAGGAGCATGG
gRNA-2 primer-F	CCGGTCCTGGGCAACTTTTCGGTCA
gRNA-2 primer-R	AAACTGACCGAAAGTTGCCCAGGA
gRNA-3 primer-F	CCGGTCCTGGGCAACTTTCTGTCA
gRNA-3 primer-R	AAACTGACAGAAAGTTGCCCAGGA
donor ssDNA-G	AACAGCAGTGCAGCTGCCAGCCCTATAGCT
	CCACACATGACCGAAAGTTGCCCAGGAGCA
	TGGCATGCATGCAAATAGACT
donor ssDNA-T	AACAGCAGTGCAGCTGCCAGCCCTATAGCT
	CCACACATGACAGAAAGTTGCCCAGGAGCA
	TGGCATGCATGCAAATAGACT
Sequence check-F	CAGGAGTTTCACCAGGTGTA
Sequence check-R	GAAAAACAGCAGTGCAGCTG

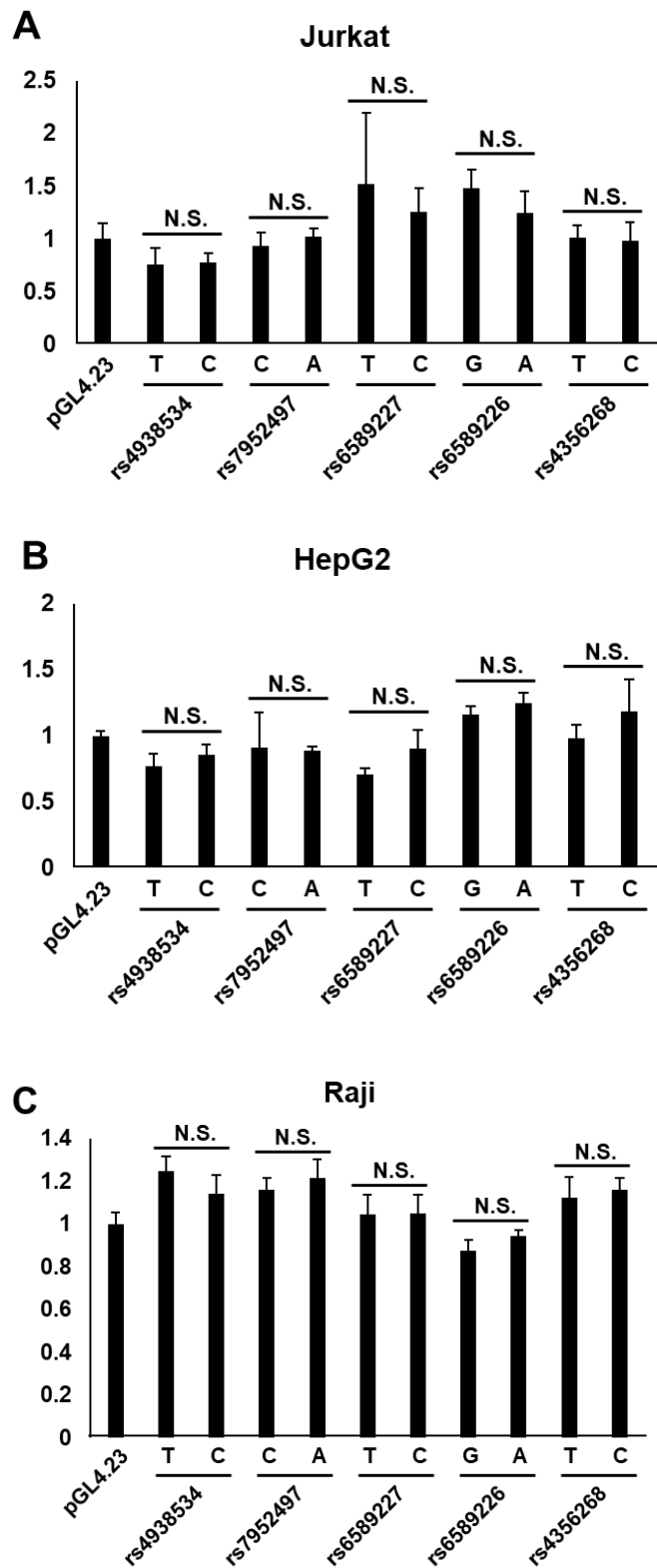
Table S4: Sequences of primers used for quantitative RT-PCR.

Gene Function	Gene name	amplicon size	Primer name	Sequence
Candidate effector gene	<i>POU2AF1</i>	71 bp	POU2AF1-F	TGGCGACCTACACCACAGT
			POU2AF1-R	CTCCTCTGTCACTGCAGAC
	<i>COLCA1</i>	155 bp	COLCA1-F	ATGTGGAGATGGACAGGGAT
			COLCA1-R	TCCTTGCTGTCCTTACAGAC
	<i>COLCA2</i>	113 bp	COLCA2-F	TCCGAGTGAAGATCACAGTG
			COLCA2-R	TGCAACTGGGTCTGAAAGGT
Housekeeping	<i>GAPDH</i>	194 bp	GAPDH-F	GCCAAGGTCATCCATGACAA
			GAPDH-R	TTCAGCTCAGGGATGACCTT

Supplementary Figures

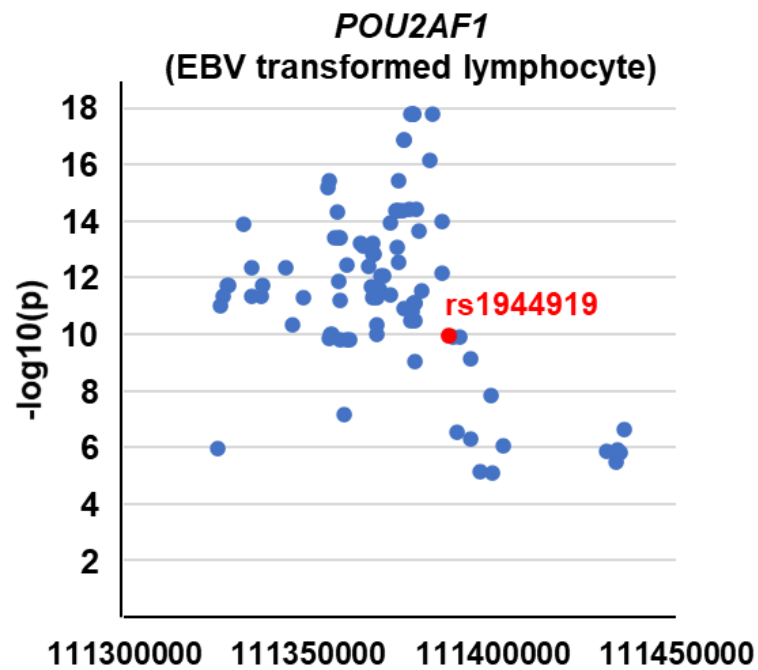


Supplementary Fig. 1. Prediction of transcription factor binding using the TRANSFAC database. (A) Predicted transcription factor binding in T allele of rs1944919. Reverse complement DNA sequences are shown. **(B)** Predicted transcription factor binding in G allele of rs1944919. Several transcription factors bind to G allele compared with T allele **(B)**. **(C and D)** EP300 **(C)** and TAF2 **(D)** expression levels in cancer cell lines. Both Jurkat and HepG2 cells showed abundant EP300 and TAF2 expression. Expression data were obtained using GeneCards from the Weizmann Institute of Science (<http://www.genecards.org/>).



Supplementary Fig. 2. Luciferase assay for other candidate SNPs.

Transcription was measured by cellular luciferase activity 24 h after transfection of Jurkat **(A)**, HepG2 **(B)**, and Raji **(C)** cells. No differences in luciferase activity were observed for the other candidate SNPs. Three independent experiments with triplicate measurements were performed for each assay. Data represent the mean $\pm$ SD; “N.S.” means non-significant (Student’s *t* test).



Supplementary Fig. 3. s-QTL mapping of *POU2AF1* in the EBV transformed lymphocyte. SNPs whose P-values less than 1.0×10^{-4} were shown. rs1944919, shown as red solid dots, did not exhibit the strongest association with *POU2AF1* splicing. rs4938534, which was the most significant SNP with PBC susceptibility on chromosome 11q23.1, did not associated with *POU2AF1* splicing ($P > 1.0 \times 10^{-4}$)