

Supplementary Information for:

**Production of IgG1-based bispecific antibody without extra cysteine
residue via intein-mediated protein trans-splicing**

Hiroki Akiba^{1,2*}, Tomoko Ise¹, Satoshi Nagata¹, Haruhiko Kamada¹, Hiroaki Ohno², and
Kouhei Tsumoto^{1,3,4*}

¹Center for Drug Design Research, National Institutes of Biomedical Innovation, Health and
Nutrition, Ibaraki, Osaka 567-0085, Japan

²Graduate School of Pharmaceutical Sciences, Kyoto University, Sakyo-ku, Kyoto 606-8501,
Japan

³School of Engineering, The University of Tokyo, Bunkyo-ku, Tokyo 133-8656, Japan

⁴The Institute of Medical Science, The University of Tokyo, Minato-ku, Tokyo 108-8639,
Japan

Sequences of the heavy chain constant region for intein-mediated protein trans-splicing

(His)₆-MBP-Int^C-hinge(partial)-Fc(hole)

MVRLPLQCVLWGCLLTAVHPSGHHHHHHGGSGKIEEGKLVIWINGDKGYNGLAEVGGKFEKDTGIKV
TVEHPDKLEEKFPQVAATGDGPDIIFWAHDRFGGYAQSGLLAEITPDKAFQDKLYPFTWDAVRYNGK
LIAYPIAVEALSLIYNKDLLPNPPKTWEEIPALDKELKAKGKSALMFNLQEPYFTWPLIAADGGYAF
KYENGKYDIKDVGVNAGAKAGLTFLVDLIKNKHMNADTDYSIAEAAFNKGETAMTINGPWAWSNID
TSKVNYGVTVLPTFKGQPSKPFVGVLSAGINAASPNKELAKEFLENYLLTDEGLEAVNKDKPLGAVA
LKSYYEELVKDPRIAATMENAQKGEIMPNI PQMSAFWYAVRTAVINAASGRQTVDEALKDAQGSGSS
GGVKIISRKSLGTQNVYDIGVEKDHNFLLNGLVASNCFNTHTCPPCPAPELLGGPSVFLFPKPKD
TLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNG
KEYKCKVSNKALPAPIEKTISKAKGQPREPQVCTLPPSRDELTKNQVSLSCAVKGFYPSDIAVEWES
NGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGK

CH1-hinge(partial)-Int^N-MBP-(His)₆

(VH) –ASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGL
YSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKKVEPKSCLSYDTEILTVYGFPLPIGKIVEERIE
CTVYTVDKNGFVYTQPIAQWHNRGEQEVFEYCLEDGSIIRATKDHKFMTTDQMLPIDEIFERGLDL
KQVDGLPGGSGGGGSGGSKIEEGKLVIWINGDKGYNGLAEVGGKFEKDTGIKV TVEHPDKLEEKFPQ
VAATGDGPDIIFWAHDRFGGYAQSGLLAEITPDKAFQDKLYPFTWDAVRYNGKLIAYPIAVEALSLI
YNKDLLPNPPKTWEEIPALDKELKAKGKSALMFNLQEPYFTWPLIAADGGYAFKYENGKYDIKDVGV
DNAGAKAGLTFLVDLIKNKHMNADTDYSIAEAAFNKGETAMTINGPWAWSNIDTSKVNYGVTVLPTF
KGQPSKPFVGVLSAGINAASPNKELAKEFLENYLLTDEGLEAVNKDKPLGAVALKSYYEELVKDPRI
AATMENAQKGEIMPNI PQMSAFWYAVRTAVINAASGRQTVDEALKDAQGHHHHHH

CH1-hinge-Fc(knob)

(VH) –ASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGL
YSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLGGPSVFLFPK
PKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDW
LNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPCRDELTKNQVSLWCLVKGFYPSDIAVE
WESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGK

Color represents each protein component.

Highlighted, reacting extein Cys; underlined, mutations introduced in the constant region of human IgG1.

Supplementary Figures

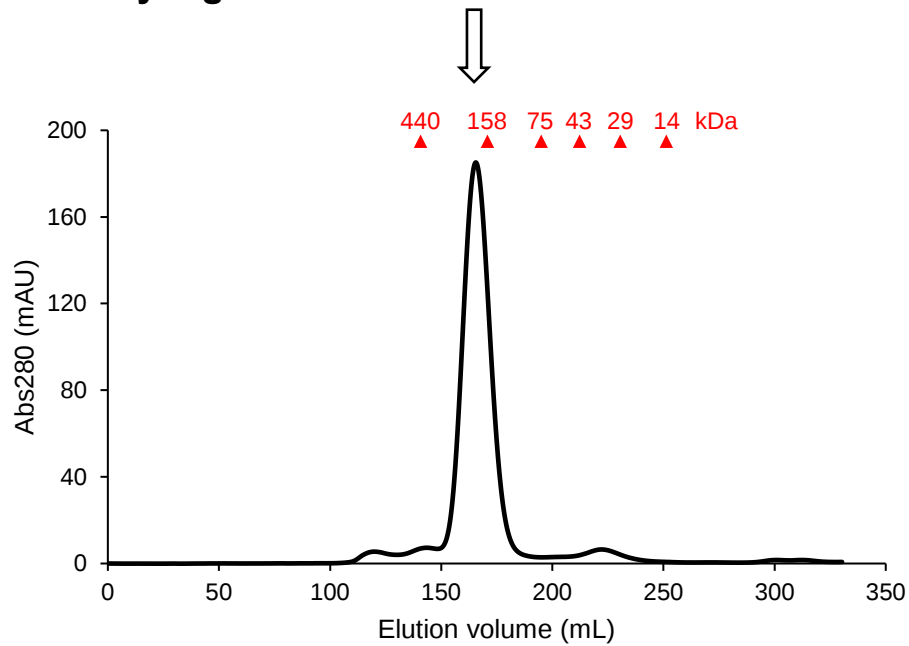


Figure S1. Size-exclusion chromatogram of Int^C/Fab^{CD30}-Fc (indicated with an arrow) obtained by transient expression in a 300-mL culture of Expi293F. Red triangles indicate the elution volume of size markers.

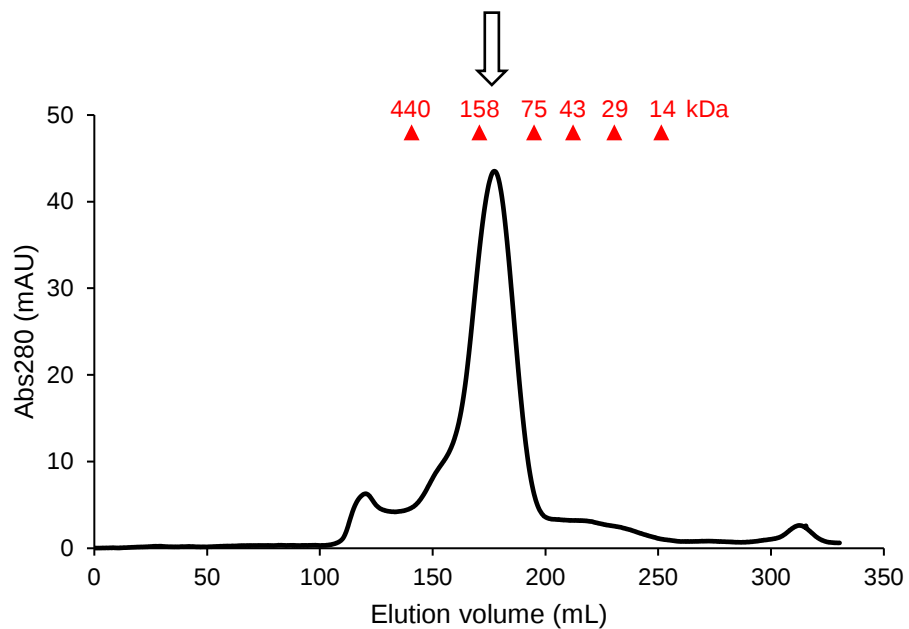


Figure S2. Size-exclusion chromatogram of Fab^{TNFR2}-Int^N (indicated with an arrow) obtained by transient expression in a 300-mL culture of Expi293F. Red triangles indicate the elution volume of size markers.

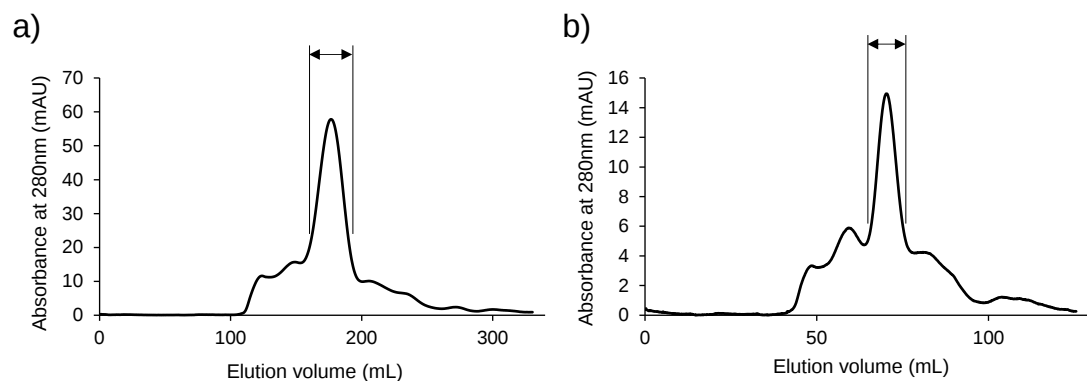


Figure S3. Size-exclusion chromatograms of Fab^{TNFR2}-Int^N using **a)** Cfa intein and **b)** Npu intein for the Int^N obtained by transient expression in 300-mL and 60-mL cultures of Expi293F, respectively, for **a** and **b**. Arrows indicate the peak area to calculate the yields.

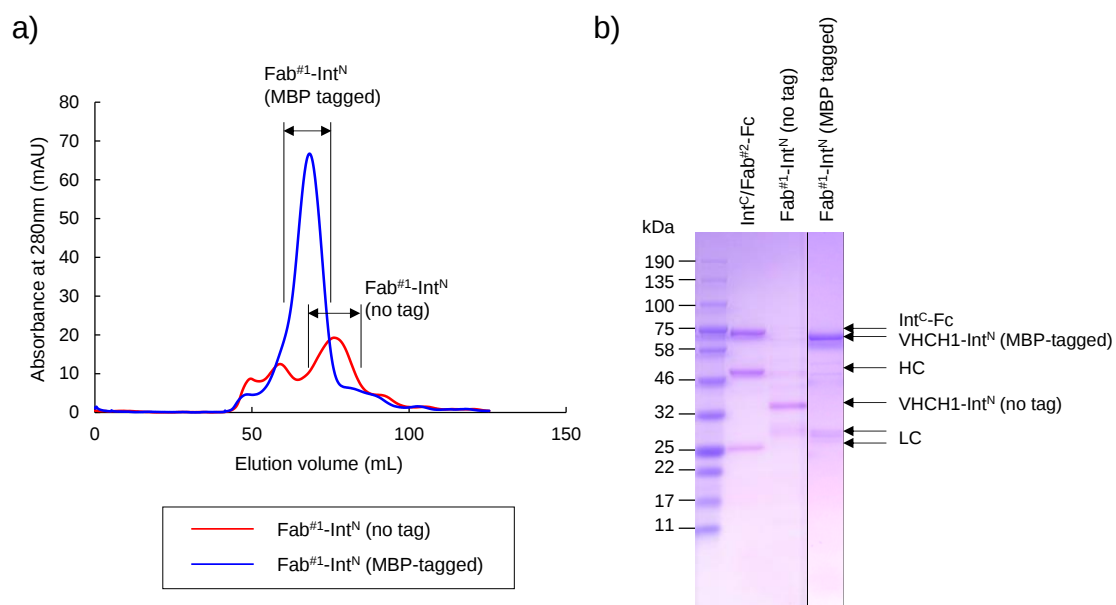


Figure S4. a) Size-exclusion chromatograms of Fab^{#1}-Int^N designed without C-terminal tag (red) or fused with maltose binding protein (MBP) tag (blue), obtained by transient expression in 30-mL cultures of Expi293F. Arrows indicate the peak area to calculate the yields. **b)** SDS-PAGE analysis of the protein in the main peaks. Black line shows the boundary of the gel images from different gels.

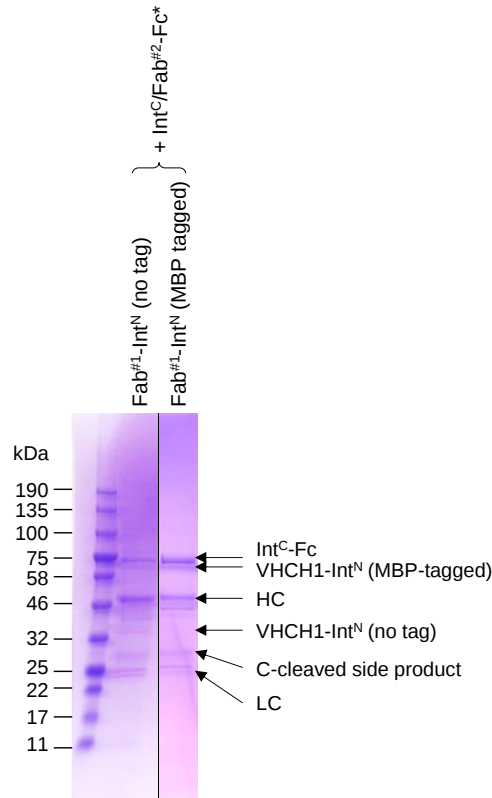


Figure S5. Comparison of the presence (MBP tagged) and absence (no tag) of C-terminal expression tag fused to IntN in an intein-mediated protein trans-splicing reaction using a Int^C-Fc construct using natural hinge sequence as the C-extein (with asterisk). Protein mixture after an overnight reaction at 37 °C in the presence of 2 mM dithiothreitol was analyzed in SDS-PAGE. Black line shows the boundary of the gel images from different gels.

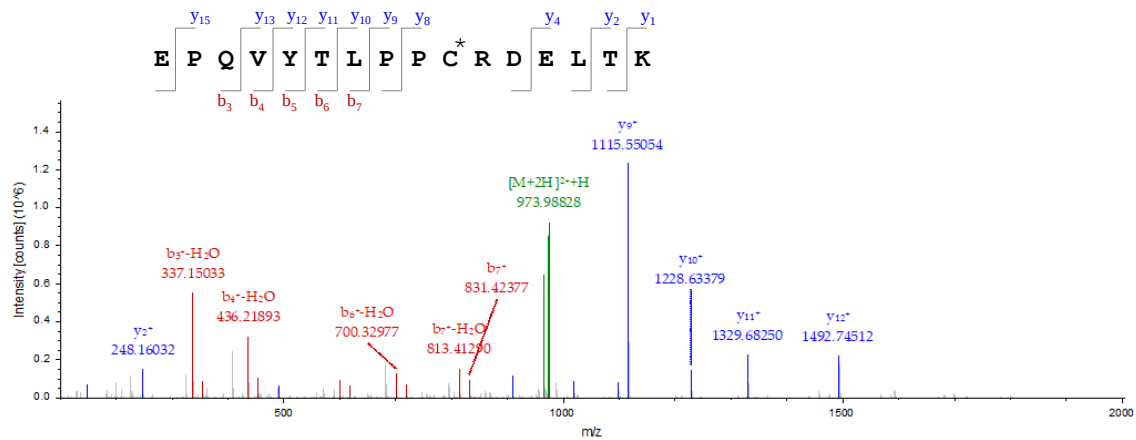


Figure S6. MS² spectrum of the selected peptide originated from the mutated Fc of T104-Fc(knob). C*, carbamidomethylated cysteine.

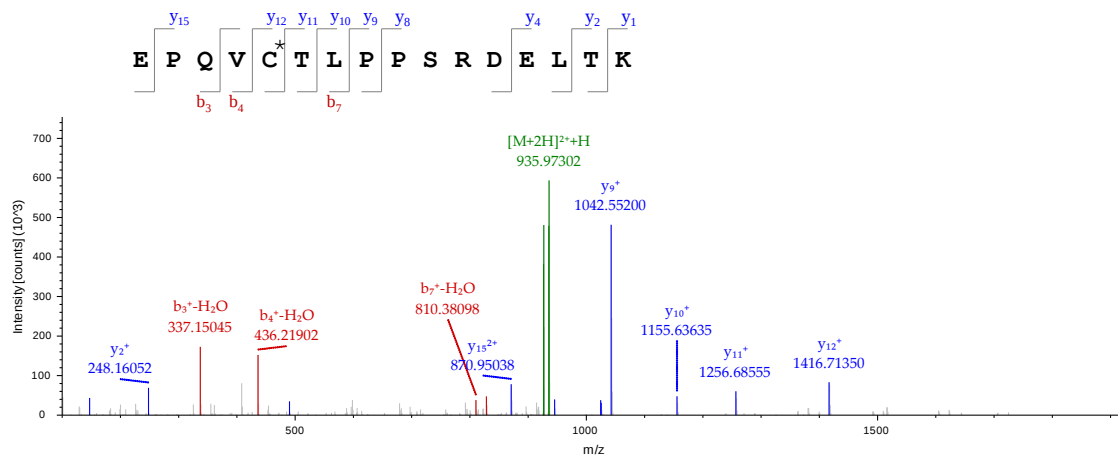


Figure S7. MS² spectrum of the selected peptide originated from the mutated Fc of TR109-Fc(hole). C*, carbamidomethylated cysteine.

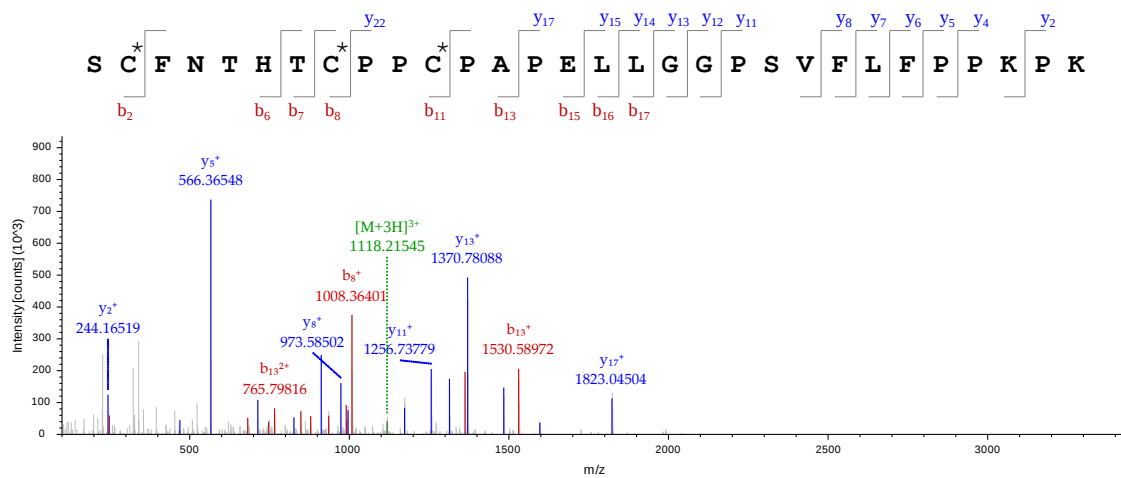


Figure S8. MS² spectrum of the peptide originated from the modified hinge region generated through the IMPTS reaction. C*, carbamidomethylated cysteine.

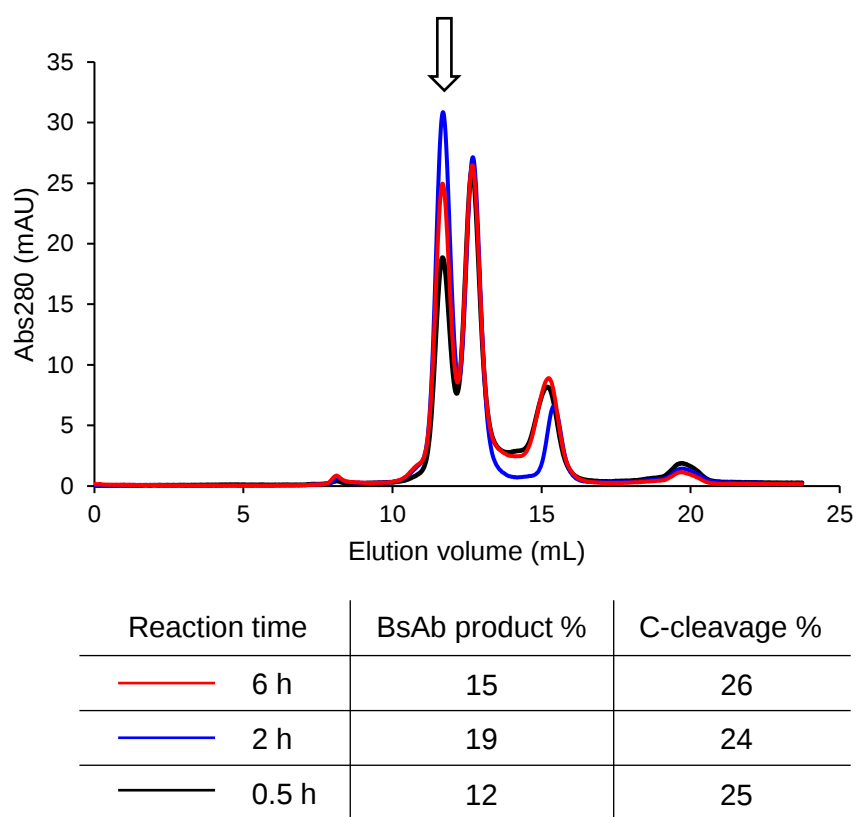
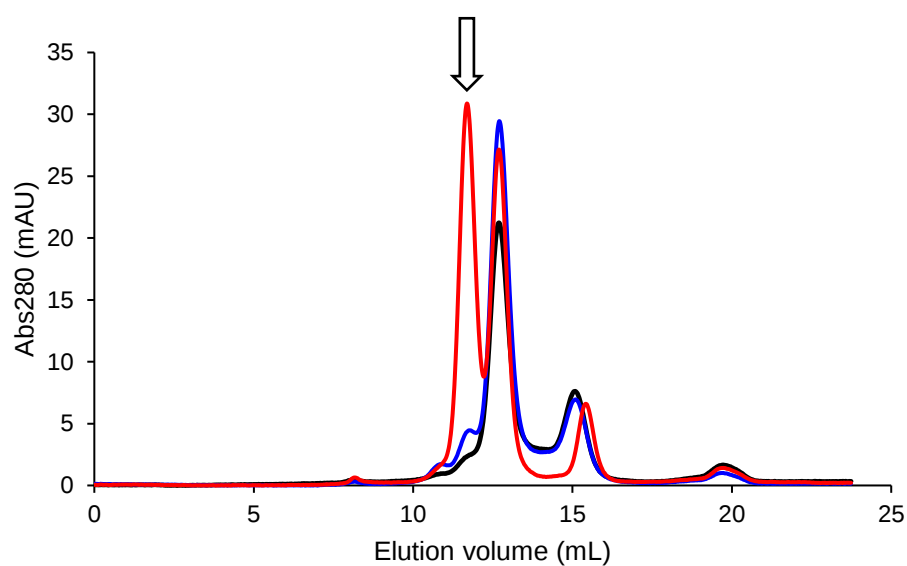


Figure S9. Time course of the IMPTS reaction in the presence of DTT in the size-exclusion chromatogram of the flow-through fraction of the amylose-affinity separation. Black, 0.5 h; blue, 2 h; red, 6 h (original condition). The arrow indicates the BsAb product. Values of % yield of the BsAb product and C-cleaved side product were determined as the ratio of the products to the precursor Int^C/Fab^{#1}-Fc in mol. The quantity of the product was calculated from the area under the chromatogram.



DTT concentration	BsAb product %	C-cleavage %
— 2 mM	19	24
— 0.5 mM	3	30
— 0.1 mM	2	23

Figure S10. Size-exclusion chromatograms of the flow-through fractions of the amylose-affinity separation in the conditions with lower concentrations of DTT. The concentration of DTT used was: black, 0.1 mM; blue, 0.5 mM; red, 2 mM (original condition). The arrow indicates the BsAb product. Values of % yield of the BsAb product and C-cleaved side product were determined as the ratio of the products to the precursor Int^C/Fab^{#1}-Fc in mol. The quantity of the product was calculated from the area under the chromatogram.

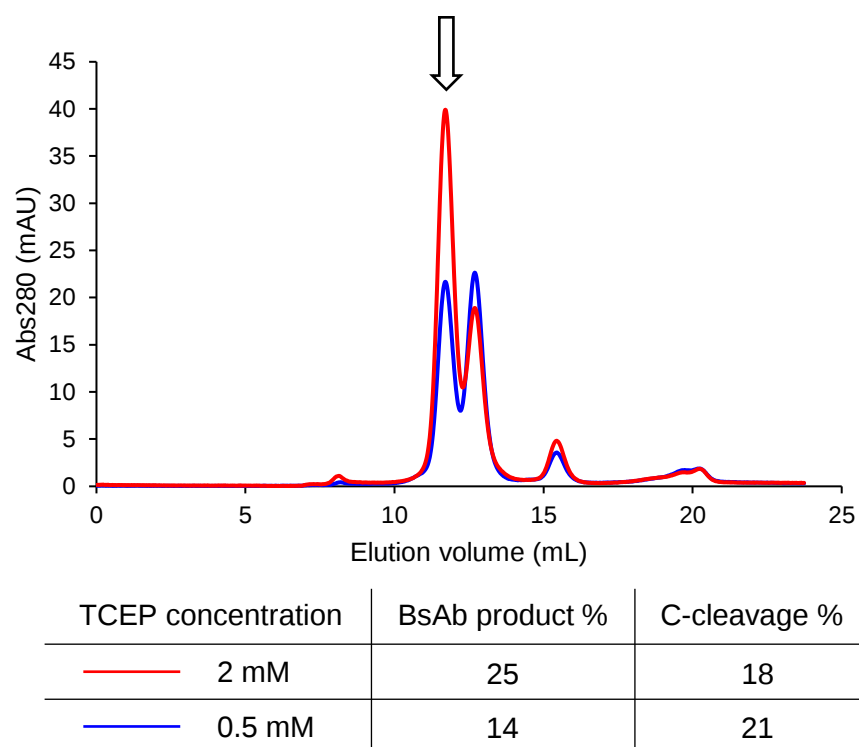
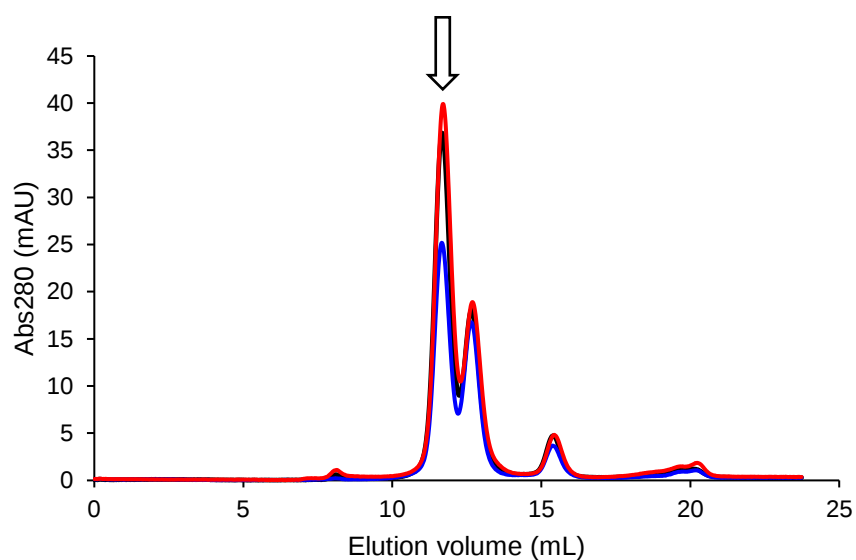


Figure S11. Size-exclusion chromatograms of the flow-through fractions of the amylose-affinity separation in the condition with lower concentration of TCEP. The concentration of TCEP used was: blue, 0.5 mM; red, 2 mM (original condition). The arrow indicates the BsAb product. Values of % yield of the BsAb product and C-cleaved side product were determined as the ratio of the products to the precursor Int^C/Fab^{#1}-Fc in mol. The quantity of the product was calculated from the area under the chromatogram.



		BsAb product %	C-cleavage %
—	Int ^C :Int ^N = 1:1.5, 37 °C	25	18
—	Int ^C :Int ^N = 1:1, 37 °C	23	17
—	Int ^C :Int ^N = 1:1, 25 °C	16	15

Figure S12. Concentration-dependency of Fab^{TNFR2}-Int^N and the effect of the temperature in the size-exclusion chromatogram of the flow-through fraction of the amylose-affinity separation. Red, original condition (Int^C : Int^N = 1 : 1.5 at 37 °C); black, decreased amount of Int^N fragment (Int^C : Int^N = 1 : 1 at 37 °C); blue, decreased temperature (Int^C : Int^N = 1 : 1 at 25 °C). The arrow indicates the BsAb product. Values of % yield of the BsAb product and C-cleaved side product were determined as the ratio of the products to the precursor Int^C/Fab^{#1}-Fc in mol. The quantity of the product was calculated from the area under the chromatogram.

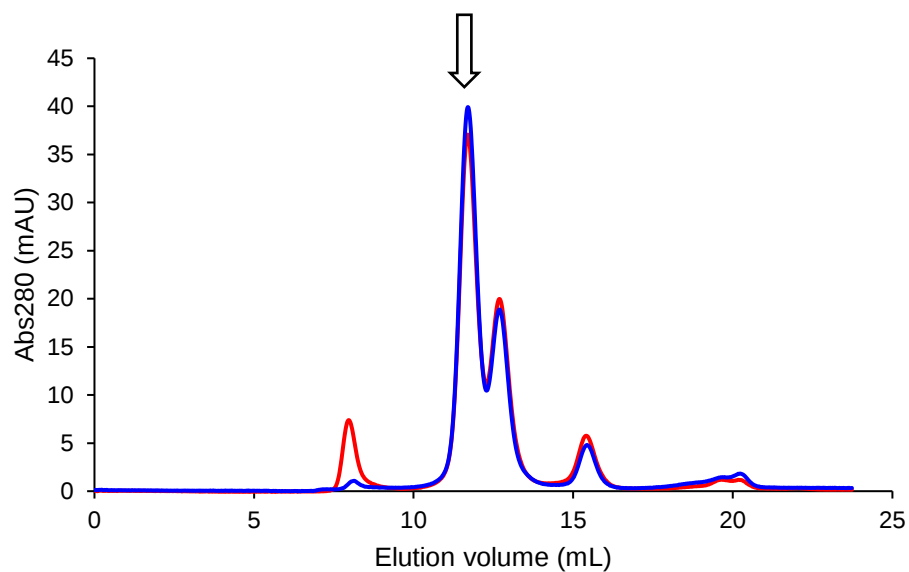


Figure S13. Time course of the IMPTS reaction in the presence of TCEP in the size-exclusion chromatogram of the flow-through fraction of the amylose-affinity separation. Blue, 2 h (original condition); red, 24 h. The arrow indicates the BsAb product.

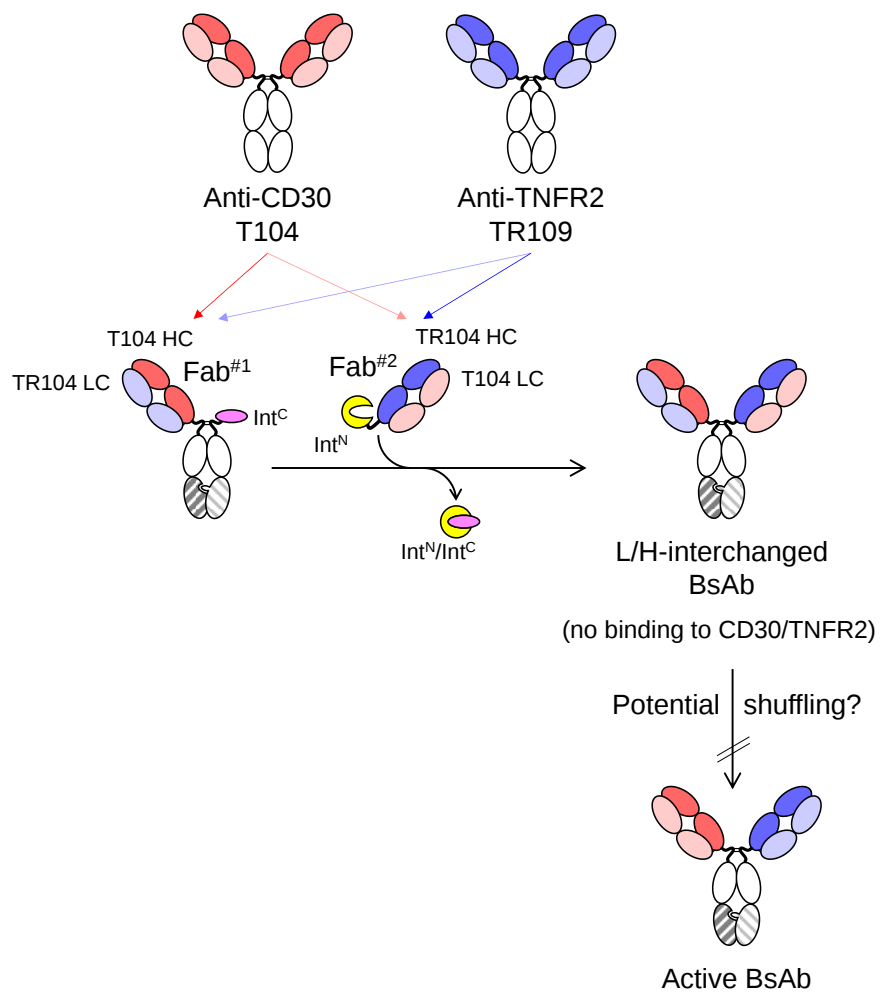


Figure S14. Design of the L/H-interchanged BsAb and the potential production of active BsAb through light- and heavy-chain shuffling (not observed in this study).

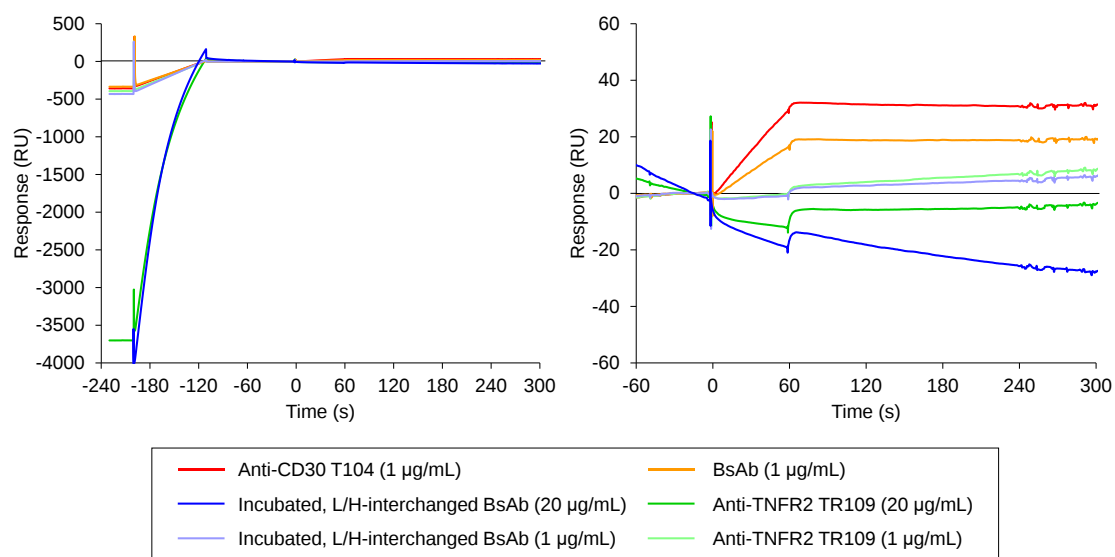


Figure S15. Interaction of the produced L/H-interchanged BsAb with recombinant CD30-rFc. Left, capture of the antibodies (–200 to –110 s), contact with CD30-rFc (0 to 60 s), and dissociation (60 to 300 s). Right, enlarged view of the contact and dissociation. Baseline did not settle during the waiting time when antibodies were captured at high concentration (20 $\mu\text{g/mL}$).

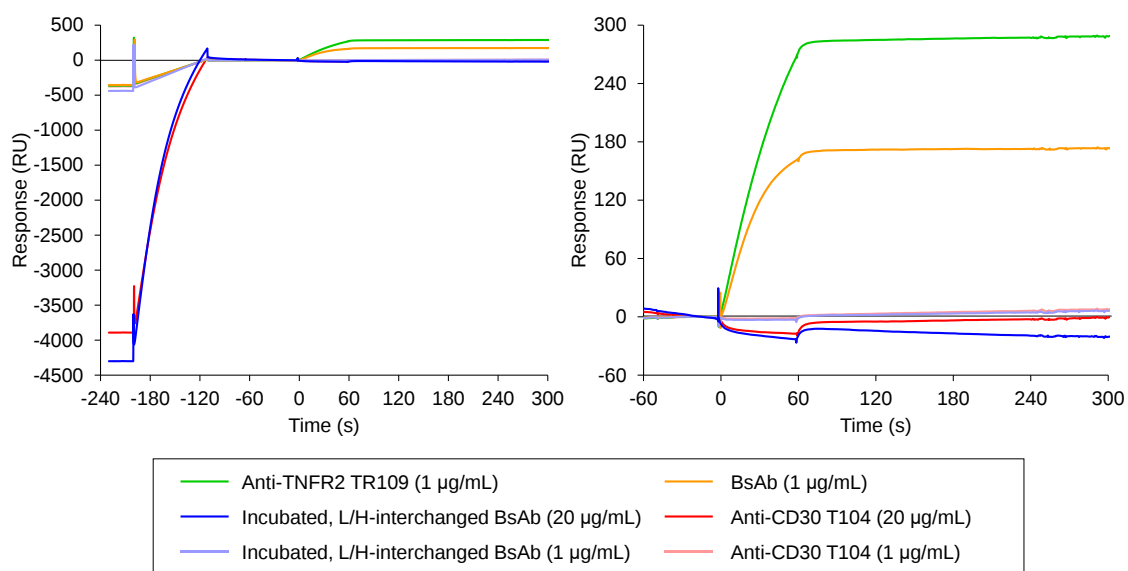
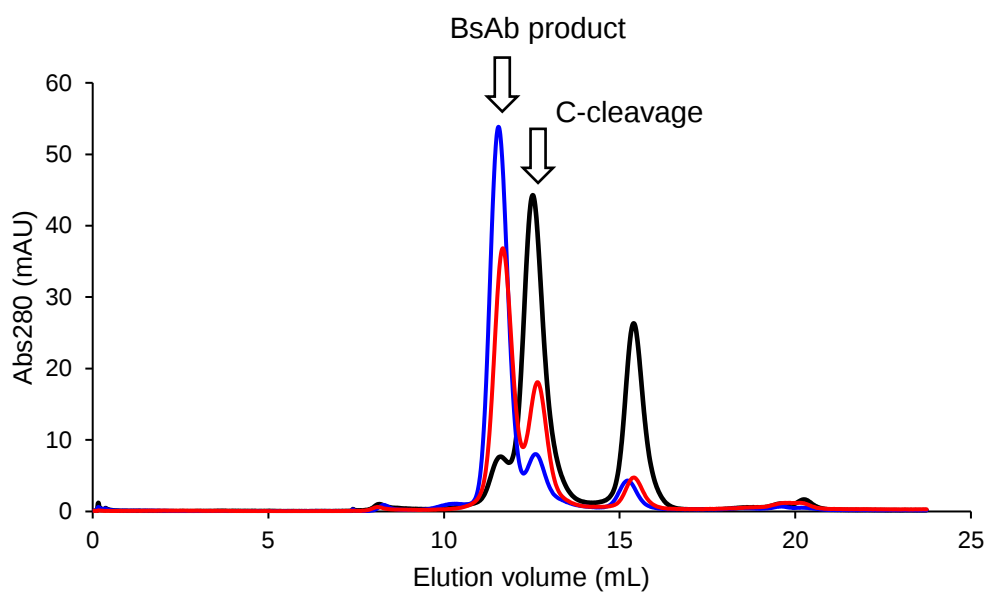


Figure S16. Interaction of the produced L/H-interchanged BsAb with recombinant TNFR2-rFc. Left, capture of the antibodies (–200 to –110 s), contact with TNFR2-rFc (0 to 60 s), and dissociation (60 to 300 s). Right, enlarged view of the contact and dissociation. Baseline did not settle during the waiting time when antibodies were captured at high concentration (20 $\mu\text{g/mL}$).



	BsAb product %	C-cleavage %
— FN mutation (this study)	23	17
— AEYCFN insertion (Hofmann et al)	32	8
— CFN insertion (Han et al)	5	47

Figure S17. Size-exclusion chromatograms of the flow-through fractions of the amylose-affinity separation after the IMPTS reaction as a comparison of three different designs of extein sequences in the hinge region. Values of % yield of the BsAb product and C-cleaved side product were determined as the ratio of the products to the precursor Int^C/Fab^{#1}-Fc in mol. The quantity of the product was calculated from the area under the chromatogram.

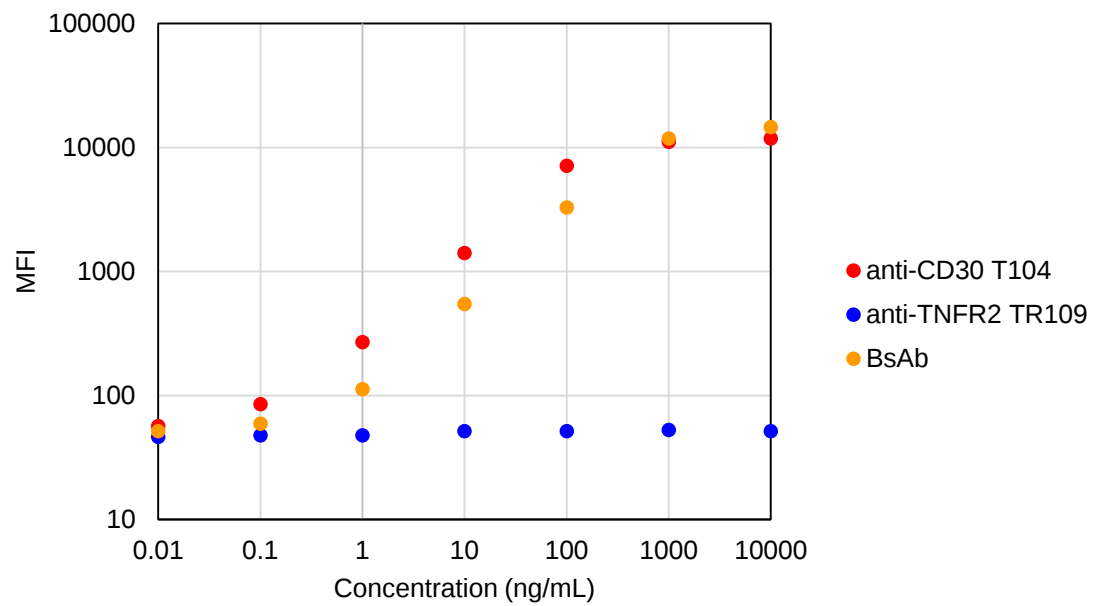


Figure S18. Concentration dependency of median fluorescence intensity of T104 and the BsAb interacting with the CD30-expressing cells.

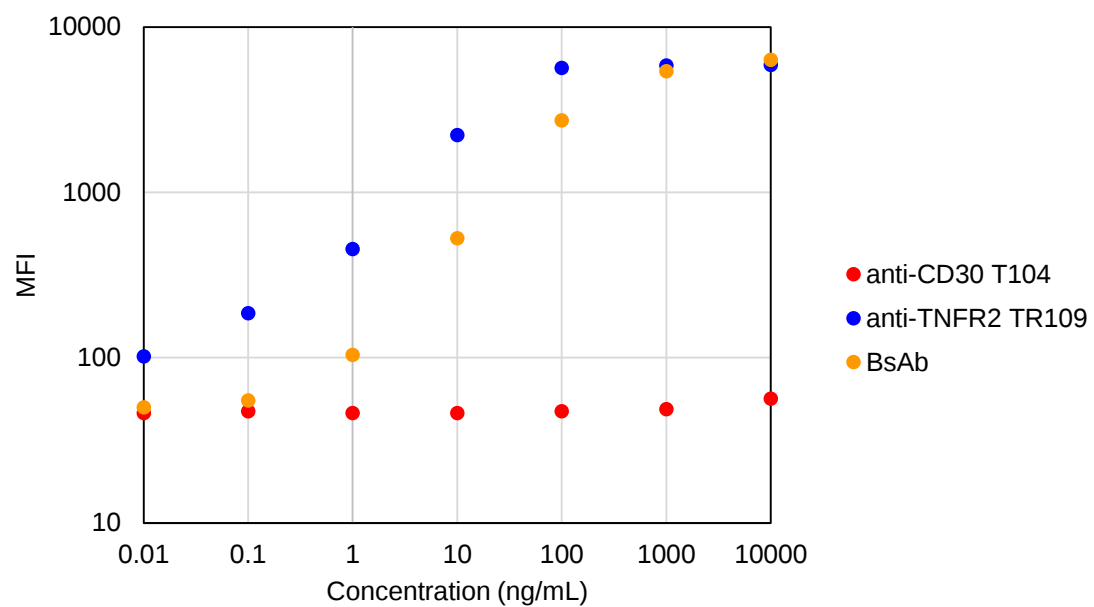


Figure S19. Concentration dependency of median fluorescence intensity of TR109 and the BsAb interacting with the TNFR2-expressing cells.