

Supplementary Information

AKR1C1/2 inhibition by MPA sensitizes platinum resistant ovarian cancer towards carboplatin

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Supplementary Figures

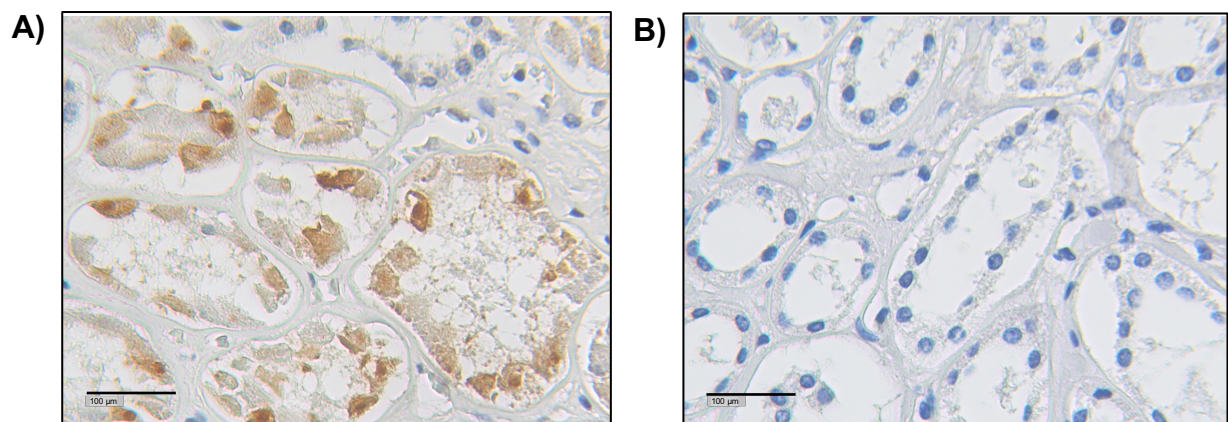


Figure S1. Positive (A) and negative (B) system control for AKR1C1/2 staining in kidney (25x magnification, scale bar=100 µm).

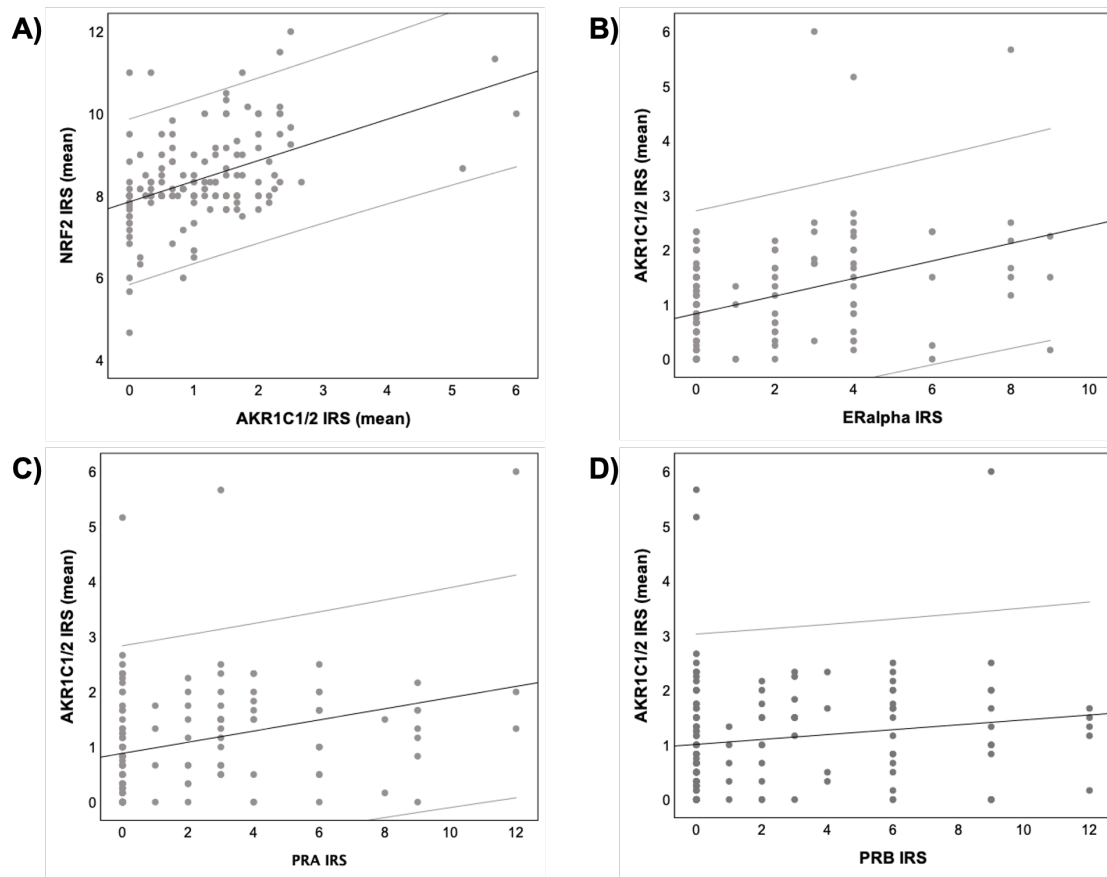


Figure S2. Graphic representations of the correlations between NRF2, AKR1C1/2 and the hormone receptors shown in Table 2 of the main manuscript. IRS of NRF2 (mean) and AKR1C1/2 (mean) were correlated to each other and to the IRS of hormone receptors using Spearman's correlation analysis. A linear fit as well as a 95% confidence interval were inserted into the scatter plots.

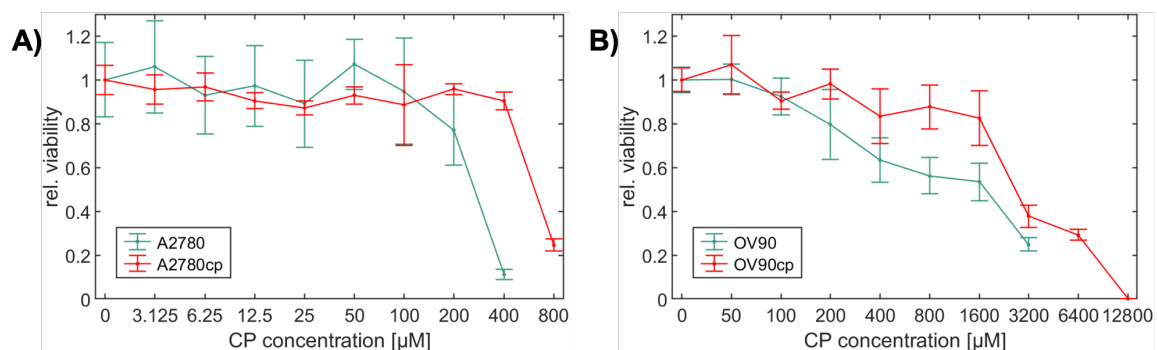
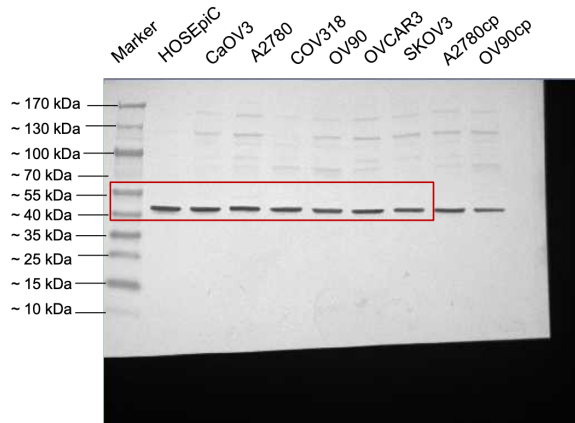
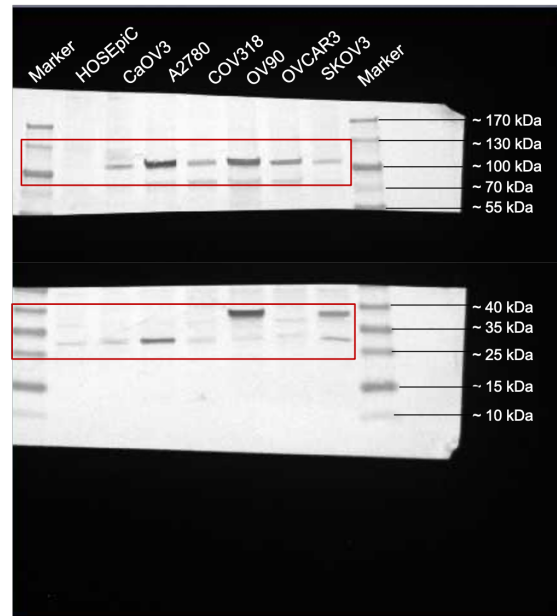


Figure S3. OV90/OV90cp (B) is more resistant towards CP than A2780/A2780cp (A). The viability was measured by MTT assay after 24h treatment with raising CP concentrations. To assess the chemoresistance of the cell lines, the IC50 value was determined (see Table 4).

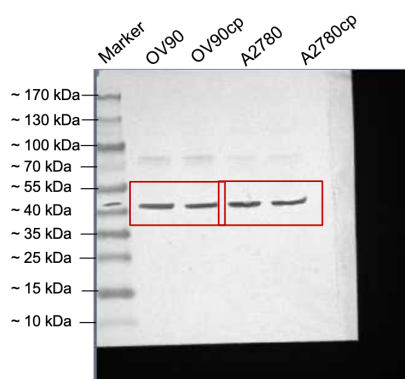
A) β -actin



NRF2 and AKR1C1/2



B) β -actin



NRF2 and AKR1C1/2

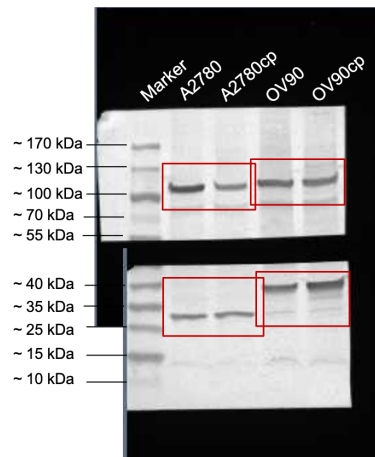


Figure S4. Full-length western blots of Figure 3 main manuscript (red boxes). A) Basal expression of NRF2 and AKR1C1/2 of ovarian cancer cell lines compared to the benign ovarian epithelial cell line HOSEpC. B) Protein expression of NRF2 and AKR1C1/2 of the endometrioid EOC cell line A2780 and the serous EOC cell line OV90 and their resistant clones. β -actin served as control. Western blot detection was performed with VECTASTAIN ABC-AmP Reagent (Vector Laboratories, Burlingame, CA, USA). Pictures were captured with Bio-Rad Universal Hood II (Bio-Rad Laboratories Inc., Hercules, CA, USA) and the corresponding Software Quantity One enabled a quantitative analysis of the blots.

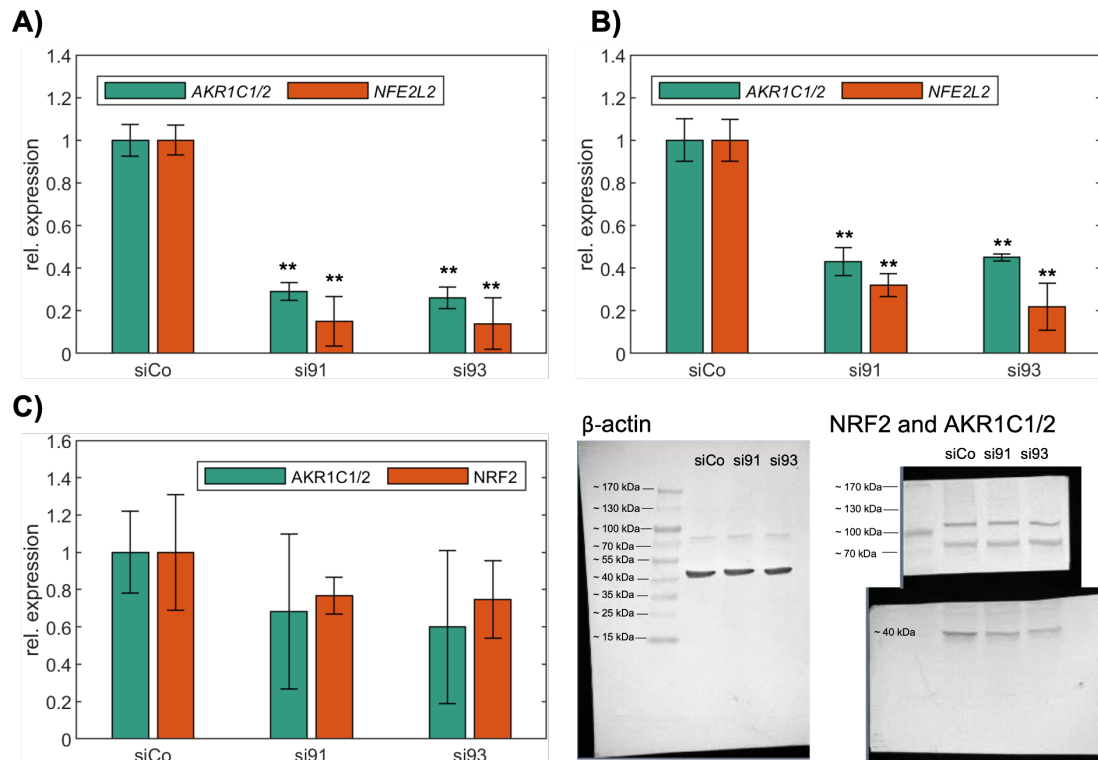


Figure S5. Successful downregulation of NRF2 and AKR1C1/2 by siRNA knockdown.

The efficiency of *NFE2L2* silencing was investigated on mRNA level by qPCR in OV90 (A) and OV90cp cells (B) and on protein level by western blot analysis (OV90, C) 24h after transfection of si*NFE2L2* (**: $p < 0.01$).

Supplementary Tables

Table S1. Descriptive statistics of expression analysis.

AKR1C1/2		NRF2	
Total	140	Total	150
IRS>0	115	IRS>0	150
Mean IRS	1.12	Mean IRS	8.36
Median IRS	1	Median IRS	8
IRS Range	0-6	IRS Range	4-12
IRS	Frequency	IRS	Frequency
0	25	4	1
0.17	5	4.67	1
0.25	3	5.67	1
0.33	8	6	2
0.5	9	6.33	2

0.67	9	6.5	3
0.75	1	6.67	2
0.83	6	6.83	2
1	10	7	2
1.17	5	7.17	2
1.25	2	7.33	3
1.33	6	7.5	3
1.5	12	7.67	6
1.67	8	7.75	1
1.75	4	7.83	6
1.83	1	8	33
2	8	8.17	10
2.17	3	8.33	12
2.25	2	8.5	10
2.33	6	8.67	3
2.5	3	8.83	5
2.67	1	9	6
5.17	1	9.17	4
5.67	1	9.25	1
6	1	9.33	1
		9.5	6
		9.67	1
		9.83	1
		10	9
		10.17	2
		10.33	1
		10.5	2
		11	3
		11.33	1
		11.5	1
		12	1

Table S2. Antibodies used for western blotting.

Antibody	Dilution	Manufacturer
NRF2, monoclonal rabbit	1:50	abcam, Cambridge, UK
AKR1C1/2, polyclonal rabbit	1:50	Sigma-Aldrich Co., St. Louis, MO, USA
β-Actin, monoclonal mouse	1:1000	Sigma-Aldrich Co., St. Louis, MO, USA

Table S3. Sequences of primers used in qPCR to determine mRNA expression levels.

<i>NFE2L2</i>	forward: CAC GGT CCA CAG CTC ATC ATG reverse: GAT CTA TAT CTT GCC TCC AAA GTA TGT C
<i>AKR1C1/2</i>	forward: CAGCAGTGCGAGGGTCAGAG reverse: GCCTTCCCATACCTGACTTCTAATC
<i>ACTB</i>	forward: TCCTCCCTGGAGAAGAGCTA reverse: CGTGGATGCCACAGGACT
<i>GAPDH</i>	forward: AGCCACATCGCTCAGACAC reverse: GCCCAATACGACCAAATCC