

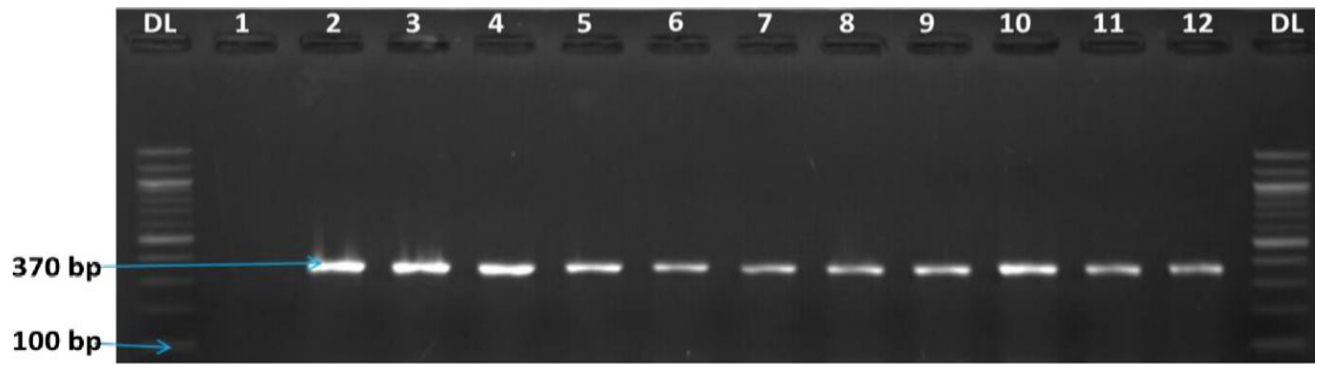


Figure S1a. Agarose gel electrophoresis of the DNA fragment generated by the simplex PCR to identify the *prs* gene (370 bp) of the *Listeria* genus. Lane DL: 100 bp DNA ladder, lane 1: -ve control, lane 2: +ve control (*L. monocytogenes* ATCC 19118), lane 3-12 positive *Listeria* genus.

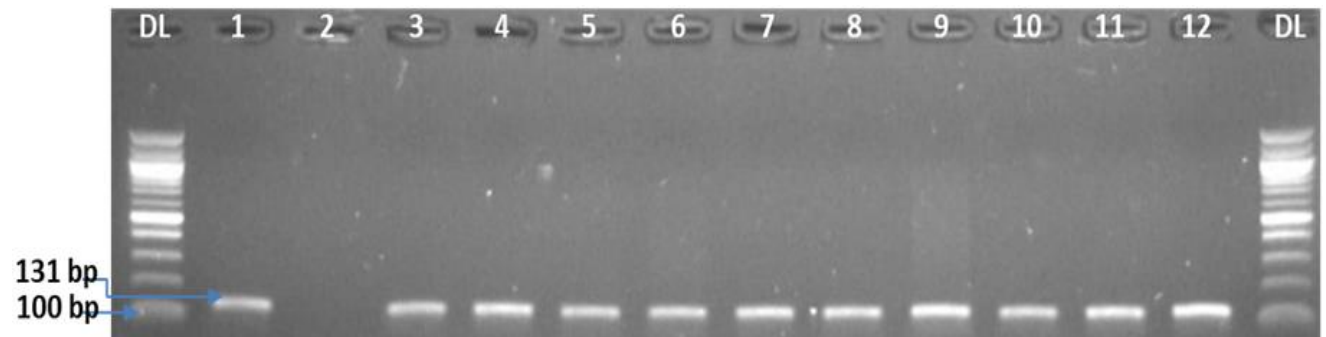


Figure S1b. Agarose gel electrophoresis of the DNA fragment generated by the simplex PCR for the amplification of the *iap* gene fragment (131 bp) for the confirmation of *L. monocytogenes*. Lane DL: 100 bp DNA ladder, lane 1: +ve control, lane 2: -ve control (*L. monocytogenes* ATCC 19118), lane 3-12 positive *L. monocytogenes* isolates.

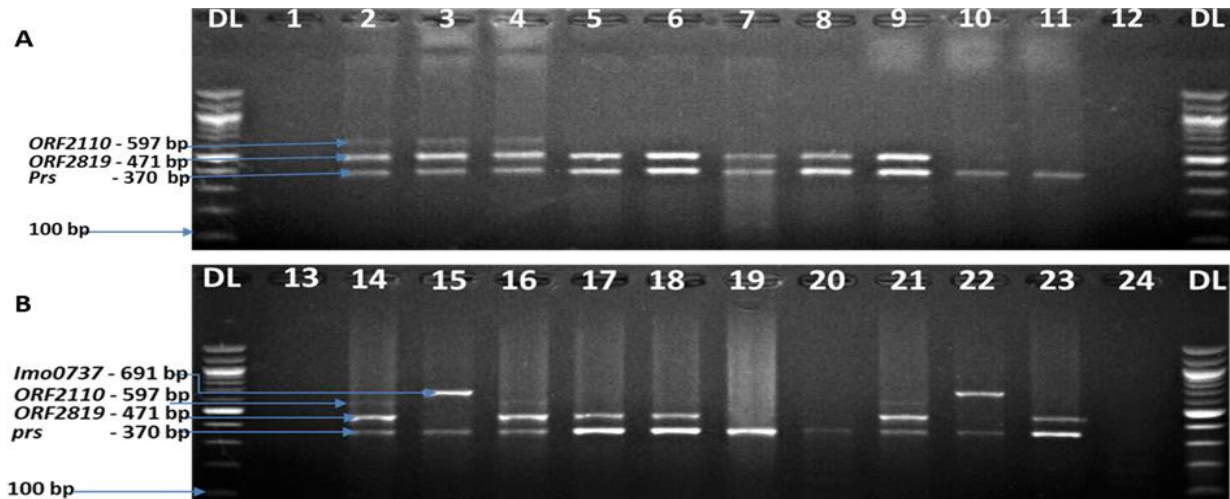


Figure S2. Agarose gel electrophoresis of the DNA fragments generated by the multiplex PCR of positive serotypes 1/2a, 1/2b and 4b strains of *L. monocytogenes*. Lane DL:100 bp DNA ladder; lane 1, 12, 13 and 24: -ve controls; lane 2 – 4, 14, 16 and 21: positive serotype 4b strains; lane 5 – 9, 17, 18 and 23: positive serotype 1/2b strains, lane 15 and 22: positive serotype 1/2a strains. All *L. monocytogenes* strains amplified the *prs* gene fragment. Genes corresponding to the amplified fragment and their molecular sizes are shown on the left.

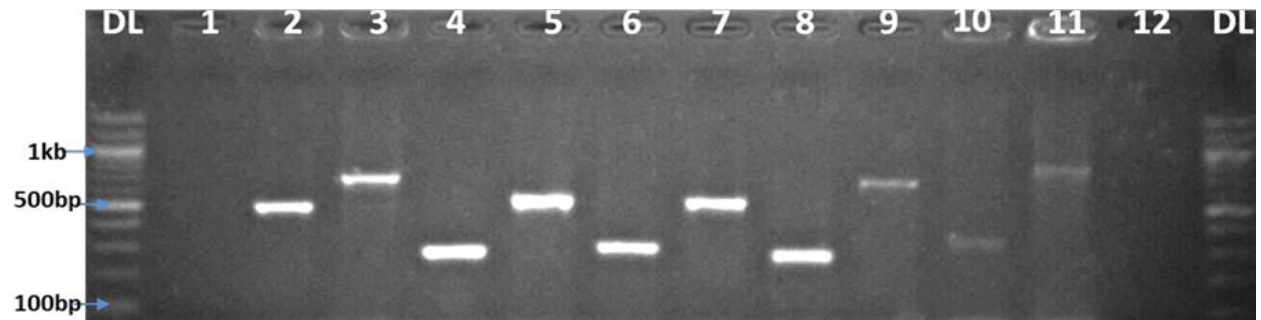


Figure S3. Agarose gel electrophoresis of the amplified DNA fragment of LIPI-1 and LIPI-2 virulence genes of *L. monocytogenes*. Lane M: 100 bp DNA ladder, lane 1 and 12: -ve control, lane 2: *hly* (496 bp), lane 3: *plcA* (674 bp), lane 4: *inlA* (256 bp), lane 5: *inlC* (517 bp), lane 6: *inlB* (272 bp) lane 7: *prfA* (479 bp), lane 8: *inlIJ* (238 bp), *actA* (650 bp), *plcB* (320 bp), and *mpl* (798 bp).

Table S1. Primer sets for the amplification of *Listeria monocytogenes* of virulence genes

Targeted genes	Primer	Sequence (5'-3')	Product size (bp)	Reference
<i>prs</i>	<i>Prs</i> -F	GCTGAAGAGATTGCGAAAGAAG	370	(Doumith et al., 2004)
	<i>Prs</i> -R	CAAAGAAACCTTGGATTTGCGG		
<i>iap</i>	<i>iap</i> -F	ACAAGCTGCACCTGTTGCAG	131	Furrer et al., 1991
	<i>iap</i> -R	TGACAGCGTGTGTAGTAGCA		
<i>inlA</i>	<i>inlA</i> -F	CCTAGCAGGTCTAACCGCAC	256	(Coroneo et al., 2016)
	<i>inlA</i> -R	TCGCTAATTTGGTTATGCCC		
<i>inlB</i>	<i>inlB</i> -F	TGATGTTGATGGAACGGTAAT	272	(Du et al., 2017)
	<i>inlB</i> -R	CTCGTGGAAGTTTGTAGATGC		
<i>inlC</i>	<i>inlC</i> -F	AATTCCCACAGGACACAACC	517	(Liu et al., 2007)
	<i>inlC</i> -F	CGGGAATGCAATTTTTCACTA		
<i>inlJ</i>	<i>inlJ</i> -F	TGTAACCCCGCTTACACAGTT	238	(Liu et al., 2007)
	<i>inlJ</i> -R	AGCGGCTTGGCAGTCTAATA		
<i>prfA</i>	<i>prfA</i> -F	AACCAATGGGATCCACAAG	479	(Jung et al., 2009)
	<i>prfA</i> -R	ATTCTGCTAACAGCTGAGC		
<i>hly</i>	<i>hly</i> -F	CAAAGTGAAGCAAAGGATGCA	496	(Jung et al., 2009)
	<i>hly</i> -R	CTAATGTATTTACTGCGTTGTTA		
<i>plcA</i>	<i>plcA</i> -F	CAGCATACTGACGAGGTGTG	674	(Jung et al., 2009)
	<i>plcA</i> -R	GATGTCCGCTCTACCTGA		
<i>plcB</i>	<i>plcB</i> -F	GCATGATATTGACAGCAAATTA	320	(Jung et al., 2009)
	<i>plcB</i> -R	TGAAATACTTTGCTCCTGTT		
<i>mpl</i>	<i>mpl</i> -F	TGTATCATCATGGTAATAGCT	798	(Jung et al., 2009)
	<i>mpl</i> -R	TGGATCCGTAAACATATTCGT		
<i>actA</i>	<i>actA</i> -F	CCAAGCGAGGTAAATACGGGA	650	(Lomonaco et al., 2012)
	<i>actA</i> -R	GTCCGAAGCATTTACCTCTTC		