

Modulation of AggR levels reveals features of virulence regulation in enteroaggregative *E. coli*

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26 **Supplementary Table 1.** List of genes belonging to the *aggR* regulon identified in
 27 previous studies^{1,2}. The positive fold change values shown were obtained for strain 042
 28 *aggR+FRT3'UTR* compared to wt.

<i>aggR</i> regulon (according to ^{1,2})	
NCBI identifier locus tag	Fold change
EC042_pAA105 (pseudo gene)	6,3
EC042_RS26280/EC042_pAA010 (dispersin export ABC transporter ATP-binding protein AatC)	3'6
EC042_RS26265/EC042_pAA007 (dispersin export ABC transporter permease subunit AatP)	3,2
EC042_RS26275/EC042_pAA009 (dispersin export-associated protein AatB)	3,2
EC042_RS11835/EC042_2219 (inverse autotransporter)	3,2
EC042_RS26510/EC042_pAA060 (AggR-activated transcriptional regulator Aar)	2,9
EC042_RS26270/EC042_pAA008 (dispersin export ABC transporter outer membrane protein aatA)	2,8
EC042_RS05935/EC042_1127 (biofilm formation regulator BssS)	2,8
EC042_RS23675/EC042_4430 (c-di-GMP phosphodiesterase PdeC)	2,8
EC042_RS27190/EC042_pAA022 (putative hexosyltransferase CapU)	2,8
EC042_RS26460/EC042_pAA046 (aggregative adherence fimbriae II chaperone AafD)	2,4
EC042_RS26345/EC042_pAA023 (virulence factor VirK)	2,3
EC042_RS26465/EC042_pAA048 (aggregative adherence fimbriae II major subunit aafA)	2,3
EC042_RS26240/EC042_pAA003 (class 1 isoprenoid biosynthesis enzyme)	2,3
EC042_RS26245/EC042_pAA004 (isopentenyl-diphosphate delta-isomerase)	2,3
EC042_RS26325 (hypothetical pseudoprotein)	2,1
EC042_RS30530/EC042_pAA055 (dispersinaap)	2,0
EC042_RS26515 (IS630 family transposase)	2,0
EC042_pAA047 (hypothetical protein)	2,0
EC042_RS26335/EC042_pAA021 (polysaccharide deacetylase family protein)	2,0
EC042_RS26330/EC042_4581 (hypothetical protein)	1,8
EC042_RS17030/EC042_3187 (AAA family ATPase)	1,8
EC042_4581 (hypothetical protein)	1,8
EC042_RS24360/EC042_4562 (type VI secretion system contractile sheath small subunit tssB)	1,8
EC042_RS24365/EC042_4563 (type VI secretion system contractile sheath large subunit tssC)	1,8
EC042_RS24370/EC042_4564 (type VI secretion system protein AaiC/Hcp2)	1,7

EC042_RS24375/EC042_4565 (putative type VI secretion protein)	1,7
EC042_RS16990 (hypothetical protein)	1,6
EC042_RS26470/EC042_pAA050 (IS1-like element IS1A family transposase)	1,6
EC042_RS30500/EC042_pAA006 (pseudogene - transposase)	1,6
EC042_RS30525/EC042_pAA053 (IS3 family transposase)	1,6
EC042_RS26480/EC042_pAA052 (aggregative adherence transcriptional regulator AggR)	1,6
EC042_RS30410/EC042_4581A (transposase)	1,5
EC042_RS17010/EC042_3184 (hypothetical protein)	1,5
EC042_RS24435/EC042_4577 (type VI secretion system ATPase TssH)	1,5
EC042_RS16995/EC042_3181 (PerC family transcriptional regulator)	1,5
EC042_RS24385/EC042_4568 (type VI secretion system baseplate subunit TssG)	1,5
EC042_RS24390/EC042_4569 (type VI secretion system tip protein VgrGTssI)	1,5
EC042_RS17015 (pseudogene)	1,5
EC042_RS12055/EC042_2249 (type IV toxin-antitoxin system toxin CbtA)	1,5
EC042_RS24430/EC042_4576 (putative type VI secretion protein)	1,5
EC042_4581B (hypothetical protein)	1,4
EC042_pAA033 (transposase)	1,4
EC042_RS24490/EC042_4587 (IS110-like element ISEc45 family transposase)	1,4
EC042_RS24460/EC042_4582 (hypothetical protein)	1,4
EC042_RS24380/EC042_4566 (type VI secretion system baseplate subunit TssF)	1,4
EC042_RS17005/EC042_3183 (phosphoadenosine phosphosulfate reductase)	1,4
EC042_RS26390/EC042_pAA031 (aggregative adherence fimbria II usher protein aafC)	1,4
EC042_RS28315/EC042_3188 (transposase)	1,3
EC042_pAA005A (conserved hypothetical protein)	1,3
EC042_RS24475/EC042_4584 (IS110 family transposase)	1,3
EC042_RS24415/EC042_4574 (type VI secretion system baseplate subunit TssK)	1,3
EC042_RS24400/EC042_4571 (type VI secretion protein)	1,3
EC042_RS24395/EC042_4570 (putative type VI secretion protein)	1,3
EC042_RS24405/EC042_4572 (type VI secretion system protein TssA)	1,3

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30 **Supplementary Table 2.** List of the 45 genes exhibiting the highest fold change values in
31 strain 042 *aggR+FRT3'UTR* compared to their expression in the wt 042 strain.

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<i>E. coli</i> 042 <i>aggR</i> + <i>FRT3'</i> UTR vs <i>E. coli</i> 042 WT	
NCBI identifier locus tag	Foldchange
EC042_RS23795 (hypothetical protein)	11,3
EC042_RS16305 (hypothetical protein)	9,4
EC042_RS17065 (CPBP family intramembrane metalloprotease)	9,4
EC042_RS21655 (hypothetical protein)	7,5
EC042_RS16275 (hypothetical protein)	6,6
EC042_RS16360 (invasion protein)	5,7
EC042_RS16280 (hypothetical protein)	5,7
EC042_RS09160 (cell division inhibition protein DicB)	5,0
EC042_RS12795 (hypothetical protein)	5,0
EC042_RS05725 (trifunctional transcriptional regulator/proline dehydrogenase/L-glutamate gamma-semialdehyde dehydrogenase)	4,9
EC042_RS09020 (hypothetical protein)	4,7
EC042_RS29440 (hypothetical protein)	4,7
EC042_RS12450 (PTS galactitol transporter subunit IIB)	4,5
EC042_RS07860 (phage lysis protein EssD)	4,4
EC042_RS10140 (N-succinylarginine dihydrolase AstB)	4,3
EC042_RS10145 (succinylglutamate-semialdehyde dehydrogenase AstD)	4,3
EC042_RS10135 (succinylglutamate desuccinylase AstE)	4,2
EC042_RS29405 (hypothetical protein)	4,1
EC042_RS22490 (fatty acid oxidation complex subunit alpha FadB)	4,1
EC042_RS02130 (hydrogen peroxide resistance inhibitor IprA)	4,1
EC042_RS22485 (acetyl-CoA C-acyltransferase FadA)	3,9
EC042_RS05770 (phosphate starvation-inducible protein PhoH)	3,9
EC042_RS03040 (hypothetical protein)	3,8
EC042_RS15365 (glutaredoxin-like protein NrdH)	3,8
EC042_RS09860 (MFS transporter)	3,8
EC042_RS09040 (YdfR family protein)	3,8
EC042_RS03755 (K(+)-transporting ATPase subunit F)	3,8
EC042_RS05765 (hypothetical protein)	3,7
EC042_RS19645 (type 1 fimbrial protein)	3,6
EC042_RS26280 (dispersin export ABC transporter ATP-binding protein AatC)	3,6
EC042_RS03060 (hypothetical protein)	3,6
EC042_RS10155 (succinylornithine/acetylornithine transaminase AstC)	3,6
EC042_RS10150 (arginine N-succinyltransferase AstA)	3,5
EC042_RS20725 (hypothetical protein)	3,5
EC042_RS16260 (helix-turn-helix transcriptional regulator)	3,5
EC042_RS01925 (hypothetical protein)	3,5
EC042_RS17395 (malate synthase G GlcB)	3,5
EC042_RS09850 (YdiL family protein)	3,3
EC042_RS29570 (hypothetical protein)	3,3
EC042_RS23860 (phosphonate C-P lyase system protein PhnG)	3,3
EC042_RS21385 (hypothetical protein)	3,2
EC042_RS26265 (dispersin export ABC transporter permease subunit AatP)	3,2
EC042_RS26275 (dispersin export-associated protein AatB)	3,2
EC042_RS15285 (carbon starvation induced protein CsiD)	3,2

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34 **Supplementary Table 3.** List of the strains and plasmids used in this work.

Strain	Description	Reference
<i>E. coli</i> 042	EAEC clinical isolate	Prof. I. Henderson
<i>E. coli</i> 042 $\Delta lacZ$	<i>E. coli</i> 042 mutant for the <i>lacZ</i> gene	This work
<i>E. coli</i> 5K Rif	<i>F</i> , <i>hsdR</i> , <i>hsdM</i> , <i>thr</i> , <i>m thi</i> , <i>rpsL</i> , <i>leu</i> , <i>lacZ</i> , Rif ^r	3
<i>E. coli</i> 55989	EAEC clinical isolate	4
<i>E. coli</i> DH5 α	<i>fhuA2 lac(del)U169 phoA glnV44 Φ80' lacZ(del)M15 gyrA96 recA1 relA1 endA1 thi-1 hsdR17</i>	5
<i>E. coli</i> 042 pAA2-	<i>E. coli</i> 042 cured from pAA2 plasmid	6
<i>E. coli</i> 042 $\Delta lacZ$ <i>aggR::lacZ</i>	<i>E. coli</i> 042 $\Delta lacZ$ with transcriptional fusion of <i>lacZ</i> reporter gene under the control of <i>aggR</i> promoter	6
<i>E. coli</i> 042 $\Delta lacZ$ <i>aggR3'UTR::lacZ</i>	<i>E. coli</i> 042 $\Delta lacZ$ with transcriptional fusion of <i>lacZ</i> reporter gene located after the end of the coding region of the <i>aggR</i> gene	This work
<i>E. coli</i> 042 <i>aggR</i> -FLAG	<i>E. coli</i> 042 with the epitope FLAG inserted in the C-terminal of the AggR protein	6
<i>E. coli</i> 042 <i>aafA</i> -FLAG	<i>E. coli</i> 042 with the epitope FLAG inserted in the C-terminal of the AafA protein	6
<i>E. coli</i> 042 <i>aap</i> -FLAG	<i>E. coli</i> 042 with the epitope FLAG inserted in the C-terminal of the Aap protein	This work
<i>E. coli</i> 042 <i>aggR::Km</i>	<i>E. coli</i> 042 with a kanamycin resistance cassette inserted in the coding region of the <i>aggR</i> gene	This work

<i>E. coli</i> 042 $\Delta aggR$	<i>E. coli</i> 042 mutant for the <i>aggR</i> gene	6
<i>E. coli</i> 042 <i>aggR</i>+Km-r3'UTR	<i>E. coli</i> 042 with a kanamycin resistance cassette inserted after the stop codon of the <i>aggR</i> open reading frame	This work
<i>E. coli</i> 042 <i>aggR</i>+FRT3'UTR	<i>E. coli</i> 042 with a FRT scar inserted after the stop codon of the <i>aggR</i> gene	This work
<i>E. coli</i> 042 <i>aggR</i>+FRT3'UTR <i>aar</i>	<i>E. coli</i> 042 with a FRT scar inserted after the stop codon of the <i>aggR</i> gene and mutant for the <i>aar</i> gene	This work
<i>E. coli</i> 042 <i>aggR</i>+FRT3'UTR <i>ast fad</i>	<i>E. coli</i> 042 with a FRT scar inserted after the stop codon of the <i>aggR</i> gene and mutant for the <i>ast</i> and <i>fad</i> operons	This work
<i>E. coli</i> 042 Δ3'UTR<i>aggR</i>	<i>E. coli</i> 042 mutant for the 3'UTR region of the <i>aggR</i> gene	This work
<i>E. coli</i> 042 ΔIRL IS1A	<i>E. coli</i> 042 mutant for the IRL sequence of the IS1A located downstream the <i>aggR</i> gene	This work
<i>E. coli</i> 042 Δ3'UTR<i>aggR</i> ΔIRL IS1A	<i>E. coli</i> 042 mutant for the 3'UTR region of the <i>aggR</i> gene and the IRL sequence of the IS1A located downstream the <i>aggR</i> gene	This work
<i>E. coli</i> 042 ΔIS1A	<i>E. coli</i> 042 mutant for the insertion element IS1A located downstream the <i>aggR</i> gene	This work
<i>E. coli</i> 042 ΔRNase E	<i>E. coli</i> 042 mutant for the C-terminal region of the RNase E protein	This work
<i>E. coli</i> 042 ΔPnp	<i>E. coli</i> 042 mutant for the polynucleotide phosphorylase protein	This work
<i>E. coli</i> 55989 <i>aggR</i> + Km-r 3'UTR	<i>E. coli</i> 55989 with a kanamycin resistance cassette inserted after the stop codon of the <i>aggR</i> open reading frame	This work
<i>E. coli</i> 55989 <i>aggR</i>+FRT3'UTR	<i>E. coli</i> 55989 with a FRT scar inserted after the stop codon of the <i>aggR</i> gene	This work

Plasmid	Description	Reference
pKD3	<i>bla</i> (Cb ^r) FRT <i>cat</i> FRT PS1 PS2 oriR6K Cm ^r	7
pKD4	<i>bla</i> (Cb ^r) FRT <i>ahp</i> FRT PS1 PS2 oriR6K Km ^r	7
pKD46	<i>bla</i> (Cb ^r) P _{BAD} <i>gam bet exo</i> pSC101 oriTS	7
pCP20	<i>bla</i> (Cb ^r) <i>cat</i> λ <i>cl857</i> /P _R / <i>flp</i> pSC101 oriTS	8
pSUB11	R6KoriV 3XFLAG-Km ^r	9
pKG136	<i>ahp</i> FRT <i>lacZY+</i> t _{his} oriR6K	10

37 **Supplementary Table 4.** List of the oligonucleotides used in this work.

Oligonucleotide	Sequence (5'-3')
aggR 042 p1	5'ACATTTTTTTCATGTGAGAATGATATGAAATTTAAACAAAACG TGTAGGCTGGAGCTGCTTC 3'
aggR 042 p2	5'TTATTGGCTTTTAAATAAGTCAAGAATTGTTTTGGTGTTATCA TATGAATATCCTCCTTAGT 3'
aggR 042 p1 up	5' GCTGCAATTAAGATACAACCCC 3'
aggR 042 3UTR p1	5'ATAACACCAAAACAATTCTTGACTTATTTTAAAGCCAATAAG TGTAGGCTGGAGCTGCTTC 3'
aggR 042 3UTR p2	5'ATATGTTTATAGCAATCTCAAATAATGATATGAAACATGTTTCA TATGAATATCCTCCTTAGT 3'
aggR 042 3UTR p1 up	5' CGCAGATTGCCTGATAAAGAC 3'
aggR 042 3UTR p2 down	5' TTGCCGTTACGCACCACTCCG 3'
aap FLAG p1	5'AGTCCAAAAATATCGTGCTCTAACCGAATGGGTAAAGACTAC AAAGACCATGACGG 3'
aap FLAG p2	5'GTGGAGAGTTGAAATTTAGCTAGAGCTAGATATTATTTAACC GACTACAAAGACCATGACGG 3'
aap FLAG p1 up	5' GCTAGCCTTCTAAAGGAGGG 3'
aap FLAG p2 down	5' ATGCGGACAACAGTGTTTACG 3'
aggR 55989 3UTR p1	5'ATAACACCAAAACAATTCTTGACTTATTTTAAAGCCAATGAG TGTAGGCTGGAGCTGCTTC 3'
aggR 55989 3UTR p2	5'ATATGTTTATAGCAATCTCAAATAATGATATAAAACATATTTCA TATGAATATCCTCCTTAGT 3'
aggR 55989 3UTR p1 up	5' GACTGTTGCGATCGTGAAGCT 3'
aggR 55989 3UTR p2 down	5' ACGGTGGCAGTCACGGTAGCG 3'
aggR IRL p1	5'TCATATCATTATTTGAGATTGCTATAAACATATTGAGATGGCG TGTAGGCTGGAGCTGCTTC 3'
aggR IRL p2	5'AGTAGCTGAACAGGAGGGACAGCTGATAGAAACAGAAGCCA CCATATGAATATCCTCCTTAGT 3'
aggR IS1a p1	5'ATTATTTGAGATTGCTATAAACATATTGAGATGGCTGAAGTTG TGTAGGCTGGAGCTGCTTC 3'
aggR IS1a p2	5'GTTTAGCGACTCGATGGAATTCGTTGTATAGATCACTTTCGC ATATGAATATCCTCCTTAGT 3'
aggR IS1a p2 down	5' CTCAATTACACCGGCTATGCC 3'
aar p1	5'TCTTATCATGTTATAAATTCAGAAAAGAGAACATTGTATTGG TGTAGGCTGGAGCTGCTTC 3'

aar p2	5'TTAGGCCAGTCTAGACAGTTTTTGTGACGACTACACTTTCATA TGAATATCCTCCTTAGT 3'
aar p1 up	5' GCCCTTTCCCATAACTCTCA 3'
aar p2 down	5' GATCGACAATCCGGGCTGAG 3'
ast p1	5'CCCTGGGCTGGATTTTCCGATGAGGTGGTGCGATGAACGCC GTGTAGGCTGGAGCTGCTTC 3'
ast p2	5'TAATACCCGCAGAATGATTTCTGCGGGTAAGTATTAGCTTATC ATATGAATATCCTCCTTAGT 3'
ast p1up	5' CGCTTACCGGCGCTGCCAGTA 3'
ast p2 down	5' TCAATTAATCAGAGCAACGGT 3'
fad p1	5'GACCAGATCACCTTGCGGATTCAGGAGACTGACATGCTTTAC GTGTAGGCTGGAGCTGCTTC 3'
fad p2	5'CGGATAAGGCGTCACGCCGCATCCGGCAAGTGGTTAAATCCG CATATGAATATCCTCCTTAGT 3'
fad p1 up	5' CTGCCGAGCGTGATCAGATCG 3'
fad p2 down	5' GTCGCATCCGGCAATCGGTGC 3'
RNase E C/Terminal p1	5'GAAACCCCGCACTACCATGTGCTGCGCGTGCGCAAAGGGGAA GTGTAGGCTGGAGCTGCTTC 3'
RNase E C/Terminal p2	5'AAGCCCTGGCAGTTACCAGGGCTTGATTGCTTGAGCTAATTAC ATATGAATATCCTCCTTAGT 3'
RNase E C/Terminal p1 up	5' GTCAGCGCCTGAGCCCATCAT 3'
RNase E C/Terminal p2 down	5' CTTAGCTGCGGCTCTGGCGG 3'
Pnp p1	5'TACCCACATTGGGCTGGGTTAGGGTTGTCATTAGTCGCGAGG GTGTAGGCTGGAGCTGCTTC 3'
Pnp p2	5'CGGTTAAAAGCCCCCGCCGAGCGGAGGGCAAATGGCAACC CATATGAATATCCTCCTTAGT 3'
Pnp p1 up	5' GCGCCTGGGTCTGCGTCGCTA 3'
Pnp p2 down	5' TGCCAGAATCACTTCCTGCTG 3'
aggR 3'	5' GAATTGTTTTGGTGTTATGCCA 3'
aggR 3' 1.2	5' GGCTTTTAAATAAGTCAAGA 3'
aggR antisense	5' CTTATGCAATCAAGAATGAG 3'
5S	5' CTACGGCGTTTCACTTCTGAGTTC 3'
aggR walking-PCR c1	5' GTCCGAATTGGTCAAAAGGAA 3'
aggR walking-PCR c2	5' AAGCCTAATGAAATATGATGT 3'
aggR walking-PCR 1	5' CTTATGCAATCAAGAATGAG 3'
aggR walking-PCR 2	5' AAGCCTAATGAAATATGATGT 3'
aggR walking-PCR 3	5' CTAAATCAGTAAGTTGGCAGC 3'
aggR walking-PCR 4	5' GTGACGTAAATCGTGTTGAG 3'
aggR walking-PCR 5	5' CGACAGCGACTTCCGTCCCAG 3'

aggR walking-PCR 6	5' CTGCGCTGATGCTGGTATGCG 3'
gapA042 RT Fw	5' TTTCCGTGCTGCTCAGAAAC 3'
gapA042 RT Rv	5' GTCAACACCAACTTCGTCCC 3'
II1bFW	5'TGTGAAATGCCACCTTTTGA3'
II1bRV	5'GGTCAAAGGTTTGGAAGCAG3'
II6FW	5'ACCAGAGGAAATTTCAATAGGC3'
II6RV	5'TGATGCACTTGCAGAAAACA3'
Hprt1FW	5'TGGATACAGGCCAGACTTTGTT3'
Hprt1RV	5'CAGATTCAACTTGCGCTCATC3'

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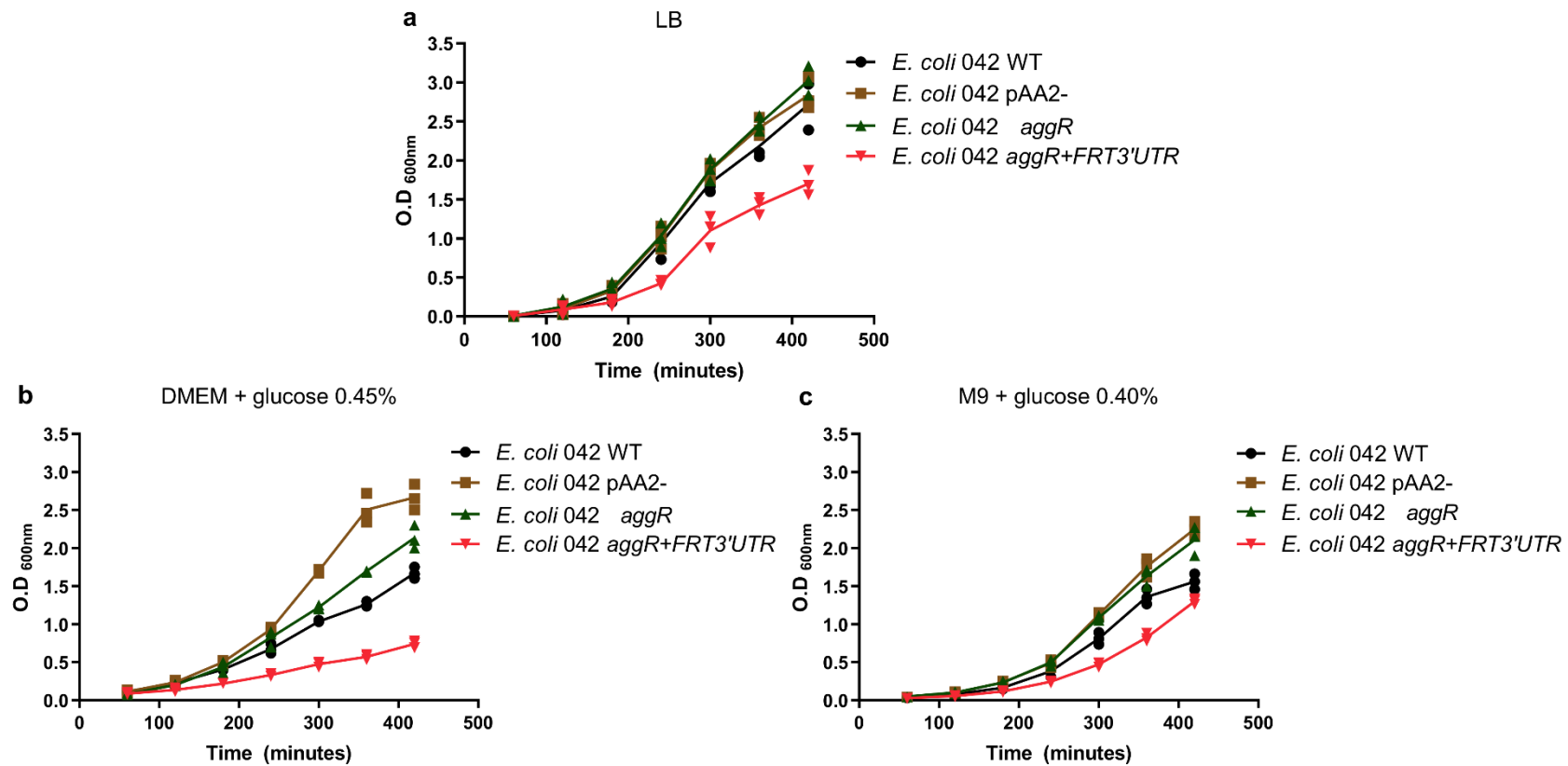
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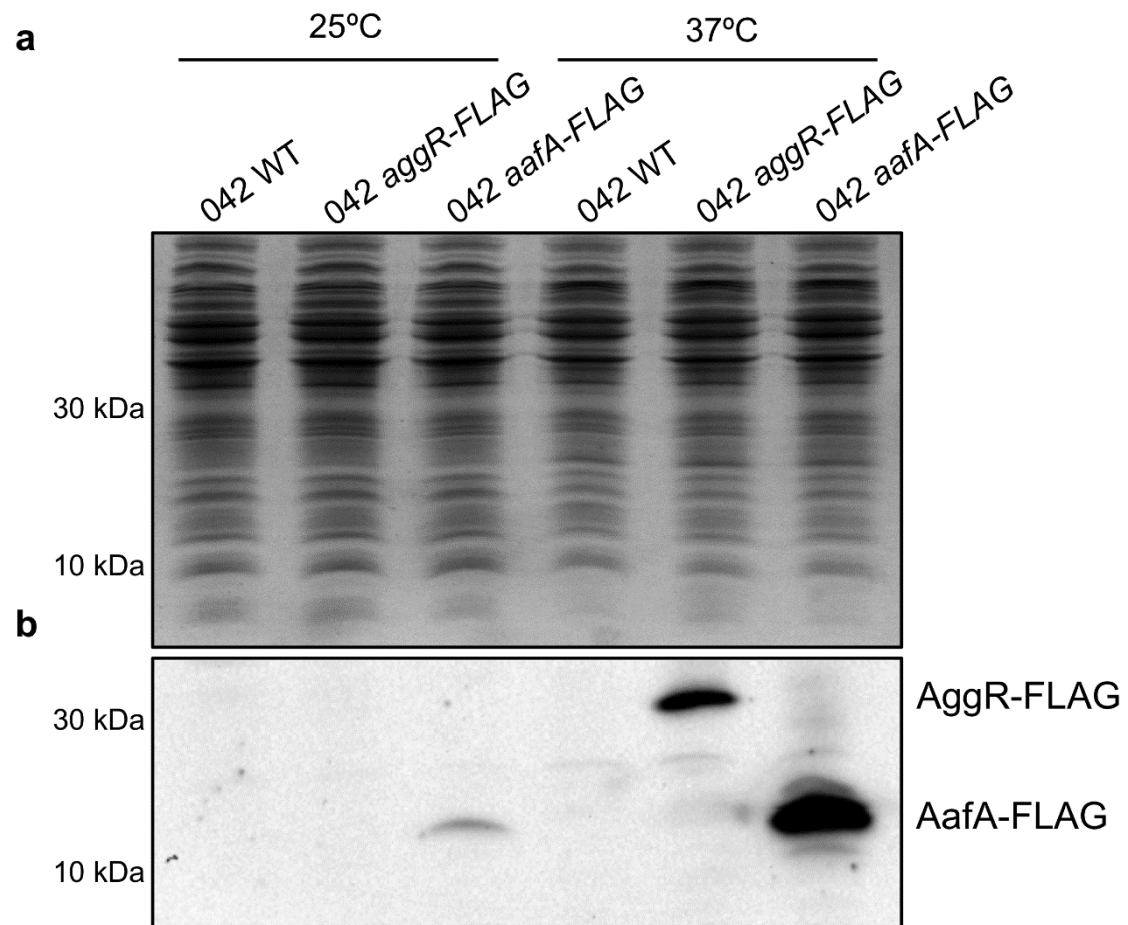
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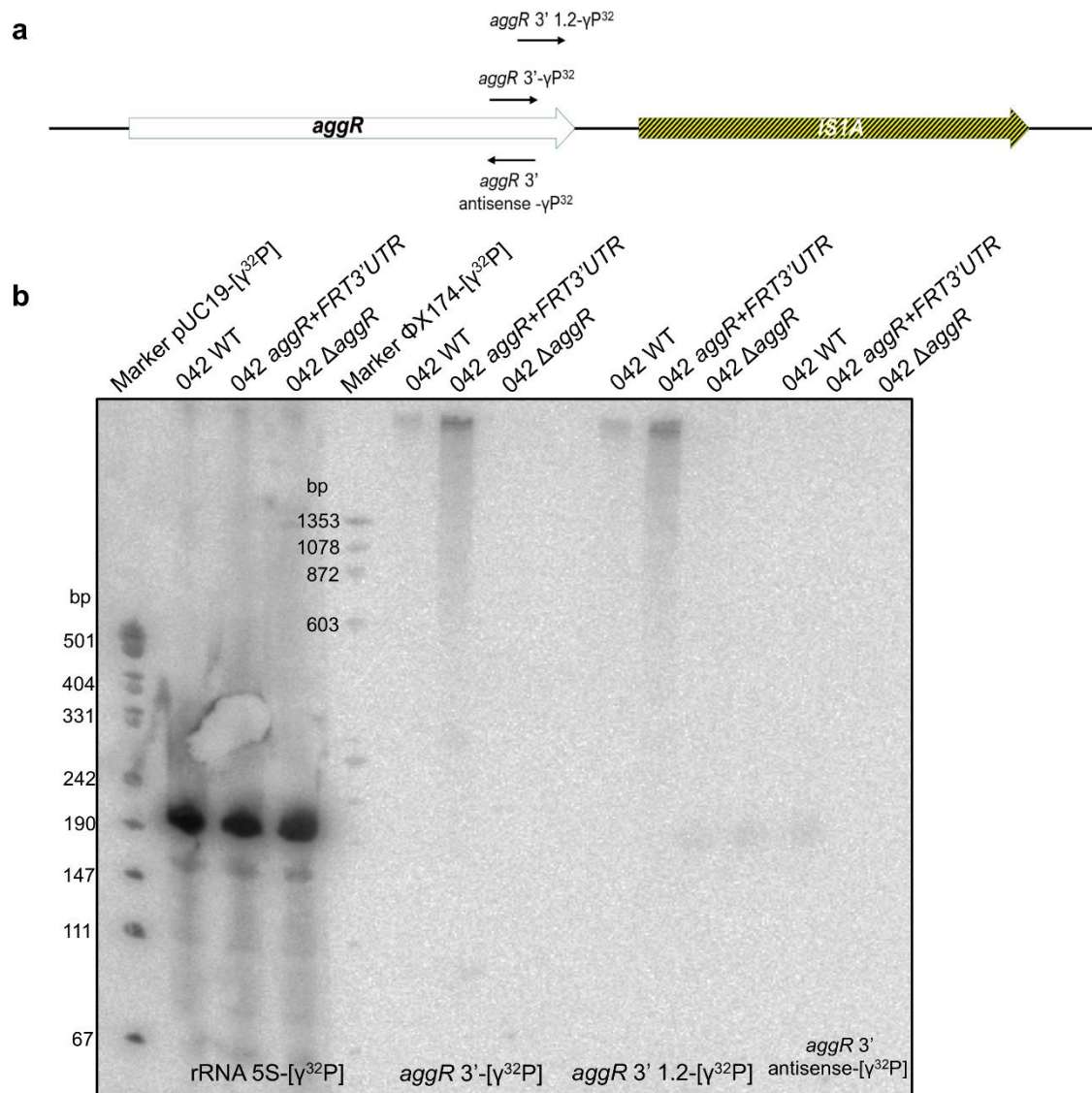
47 **Supplementary Figure 1. A FRT sequence DNA insertion in the 3'UTR of the *aggR* gene results in a reduced growth rate.** Growth curves of *E.*
 48 *coli* strains 042 wt, pAA2-, Δ *aggR* and *aggR*+FRT3'UTR in (a) LB medium, (b) DMEM + glucose 0.45% and (c) M9 minimal medium + glucose 0.4%
 49 at 37°C. Error bars correspond to the standard deviation of three independent biological replicates.

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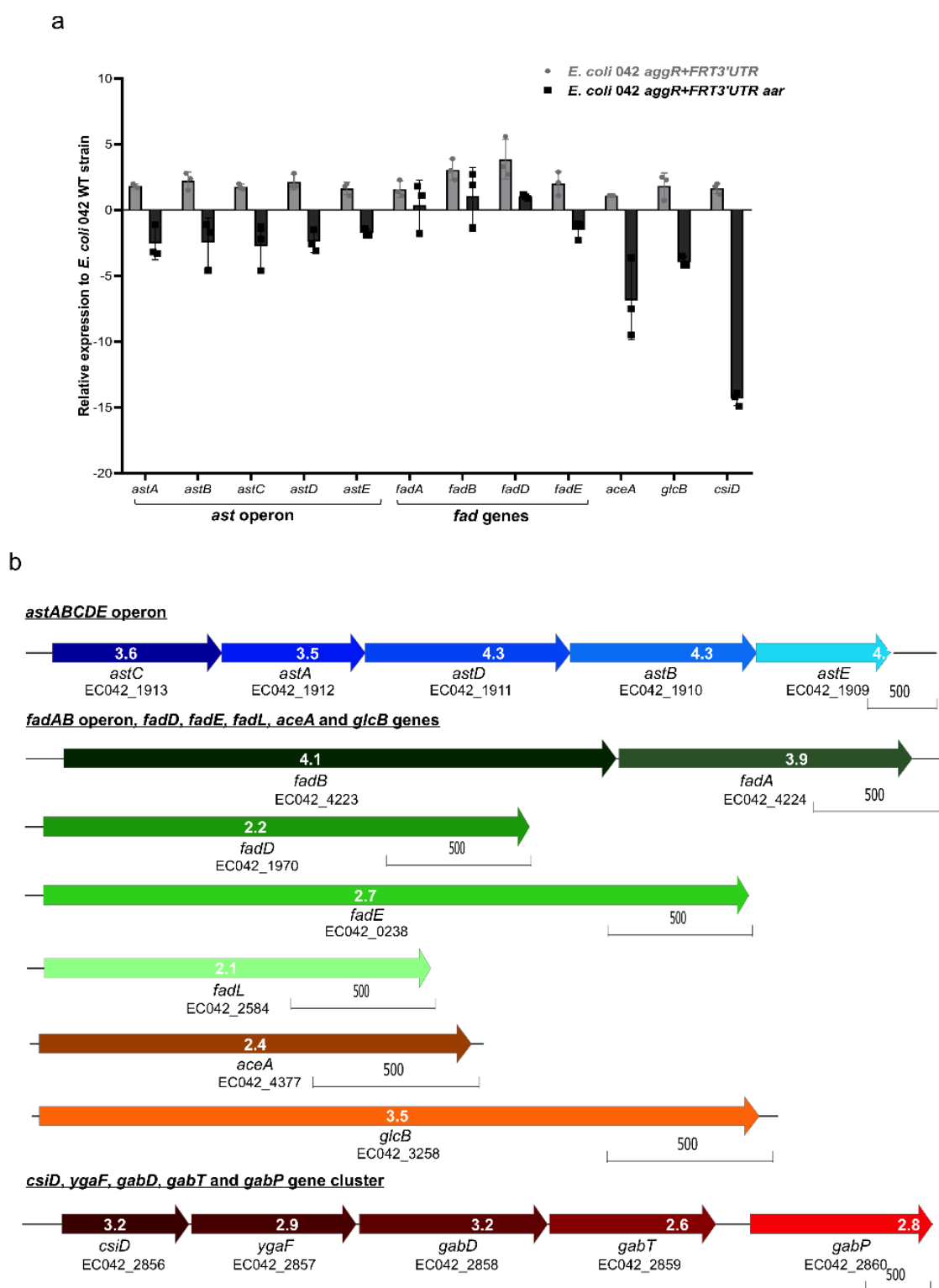


Supplementary Figure 2. AggR and AafA protein expression is temperature regulated.

(a) Corresponds to the total cell extract loaded per slot and stained with Coomassie blue (loading control). (b) Western blot analysis of the 042 *aggR*-Flag and *aafA*-Flag strains grown in LB medium at 25°C and 37°C until the OD₆₀₀ reached 2.0. Flag-tagged AggR and AafA proteins were detected using anti-Flag antibody, as indicated in the figure. The experiment was repeated three times, and a representative experiment results is presented.



Supplementary Figure 3. Northern blot analysis of the *aggR* transcript. (a) Diagram showing the localization of the probes used for the Northern blot analysis of *aggR* mRNA. (b) Full image of a representative Northern blot of the *aggR* transcript. Molecular weight markers are shown.

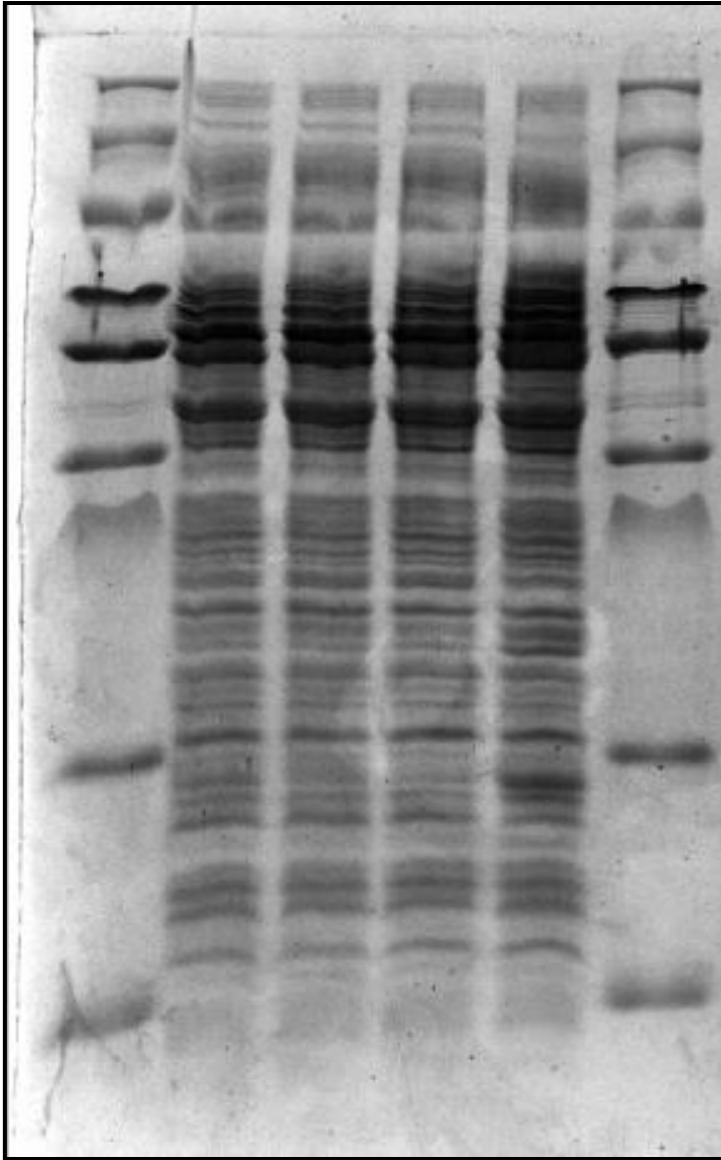


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65 **Supplementary Figure 4. Overexpression of metabolic genes in the 042 *aggR*+*FRT***

66 **3'UTR and derivatives. (a) Overexpression of metabolic genes in the 042 *aggR*+*FRT***

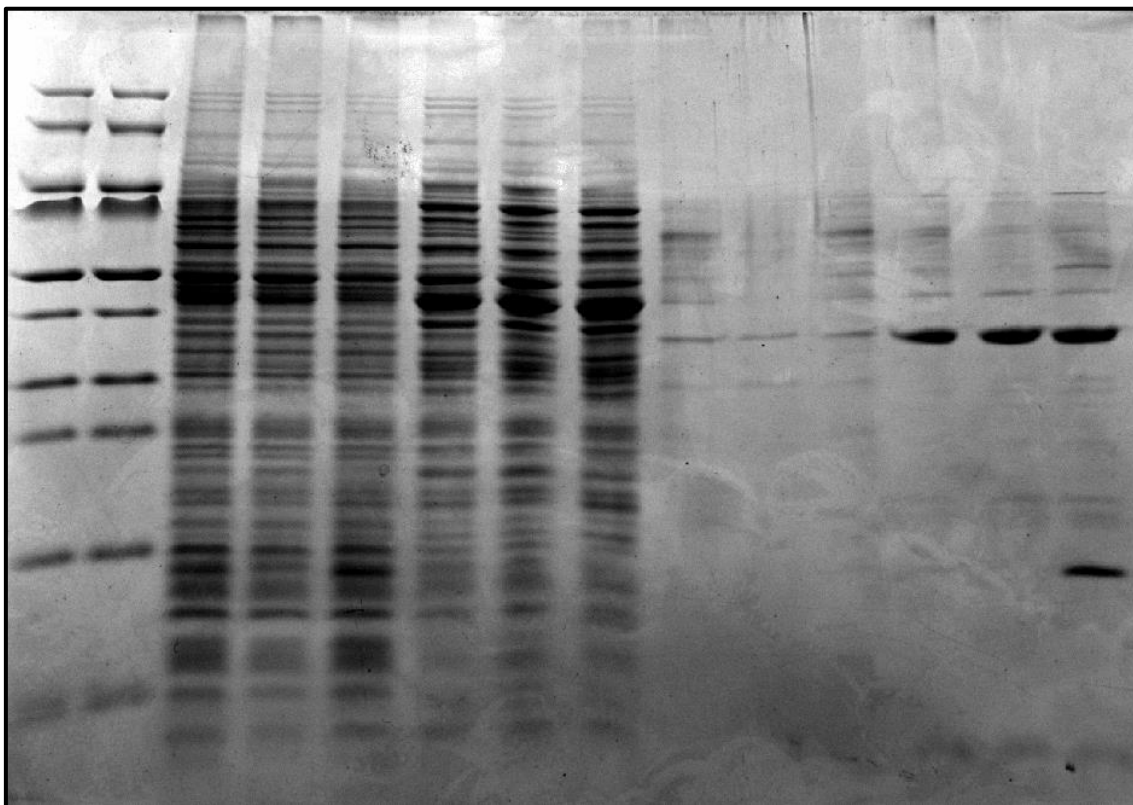
3'UTR and its *aggR+FRT 3'UTR aar* derivative compared to the wt strain. Relative expression of *ast*, *fad*, *aceA*, *glcB* and *csiD* genes quantified by RT-qPCR in *E. coli* 042 *aggR+FRT3'UTR* and its *aggR+FRT3'UTR aar* derivative. The bar shows the arithmetic mean of the results of three independent experiments and the error bar indicates the standard deviation. (b) Overexpression of metabolic genes in the 042 *aggR+FRT 3'UTR* mutant compared to the wt strain. Diagram showing the metabolic pathways leading to arginine degradation (*ast*-encoded enzymes), GABA degradation (*csiD*-, *ygaF*-, and *gabDTP*-encoded enzymes) and fatty acid degradation (*fad*-and *ace*-encoded enzymes). Foldchange values for each gene in the 042 *aggR+FRT3'UTR* mutant compared to the wt strain are shown.



Supplementary Figure 5. Uncropped SDS-PAGE image corresponding to the Figure 2a.

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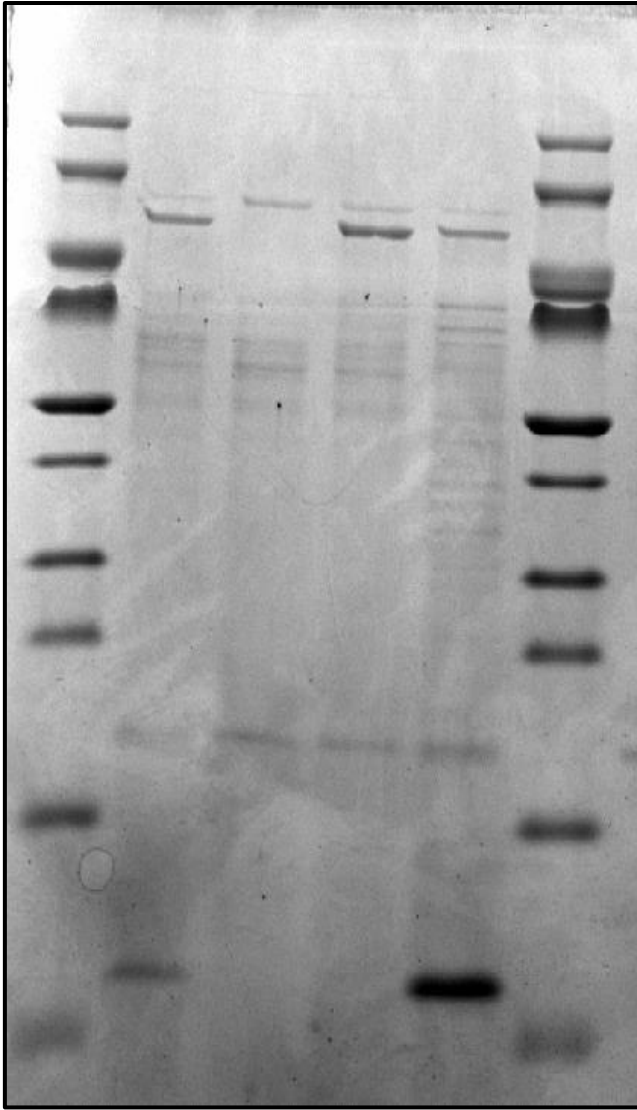
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93 **Supplementary Figure 6.** Uncropped SDS-PAGE image corresponding to the Figure 2b.

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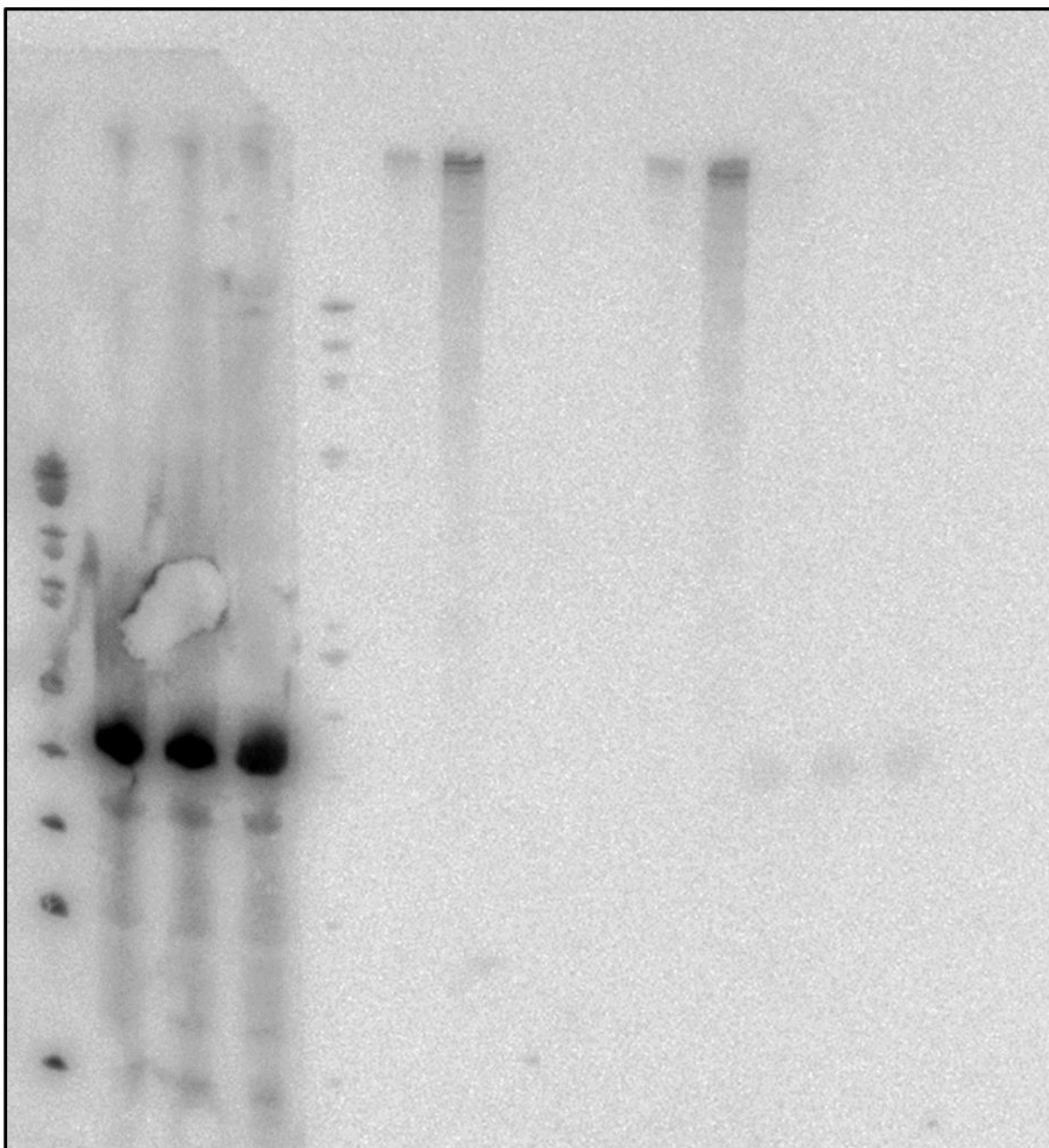
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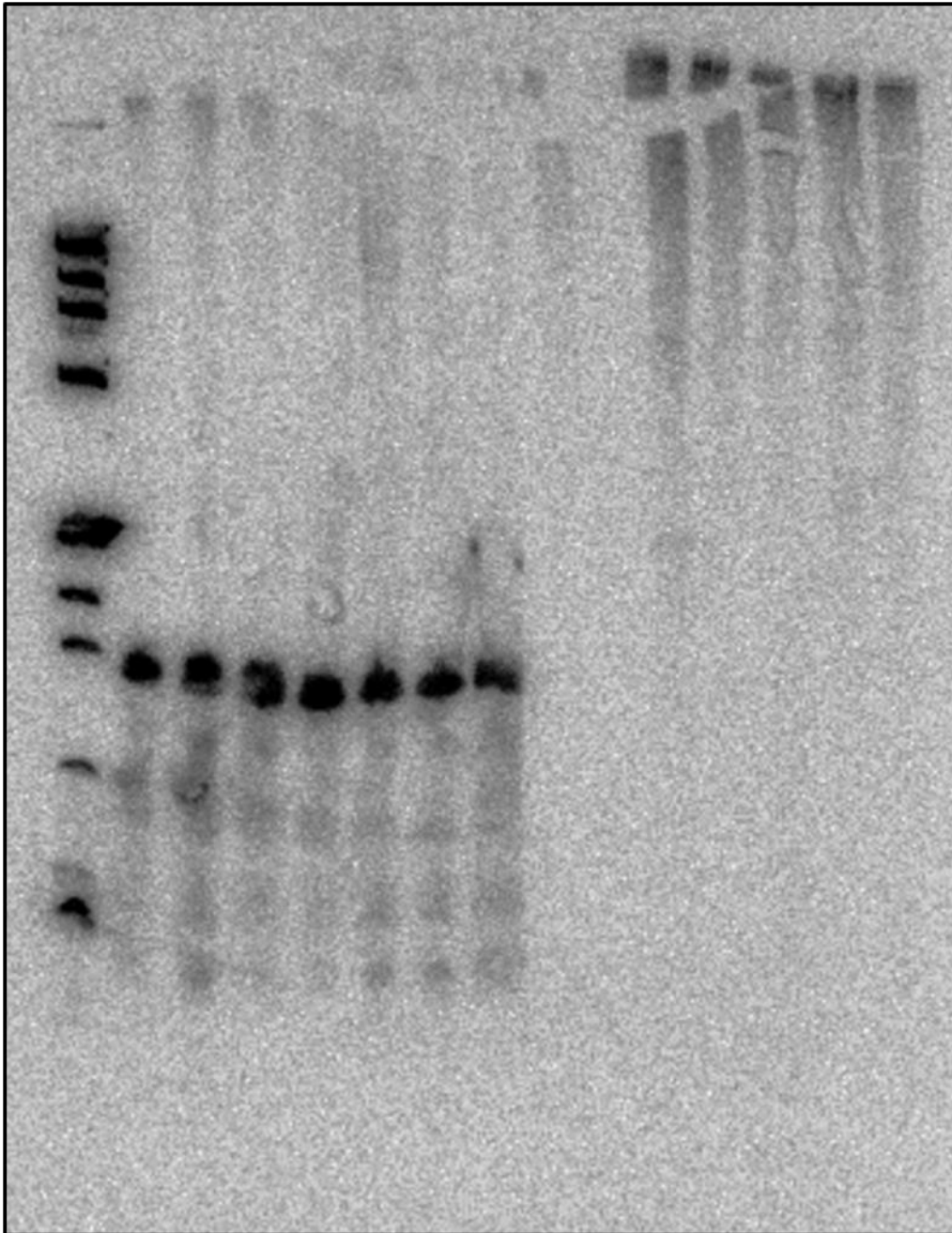
Supplementary Figure 7. Uncropped SDS-PAGE image corresponding to the Figure 2c.

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111 **Supplementary Figure 8.** Uncropped northern blot image corresponding to the Figure
112 3.

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Supplementary Figure 9. Uncropped northern blot image corresponding to the Figure 6.

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125 **Supplementary Figure 10.** Uncropped northern blot image corresponding to the Figure
126 8a.

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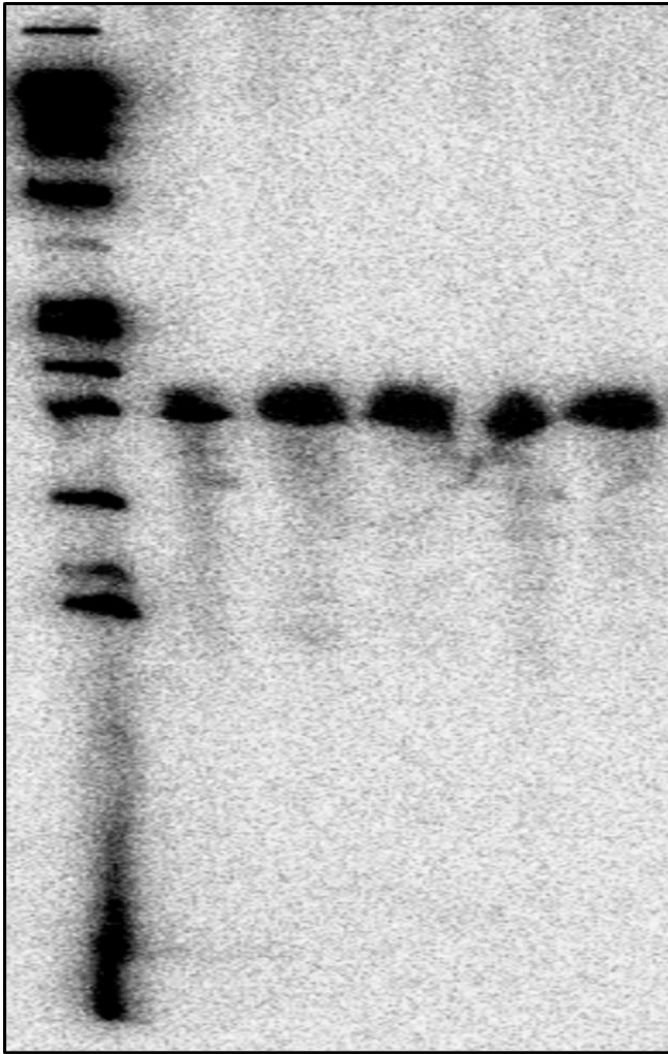
128

129

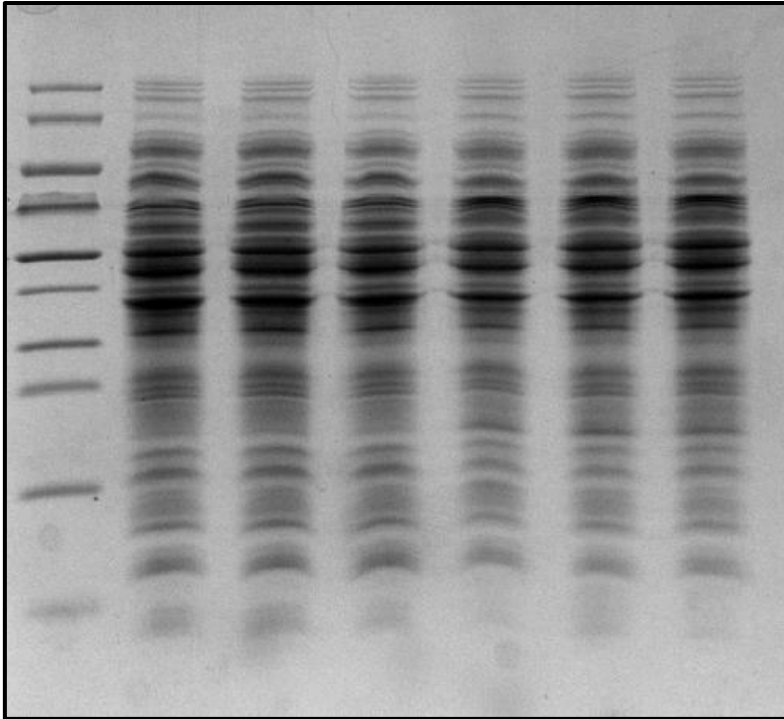
130

131

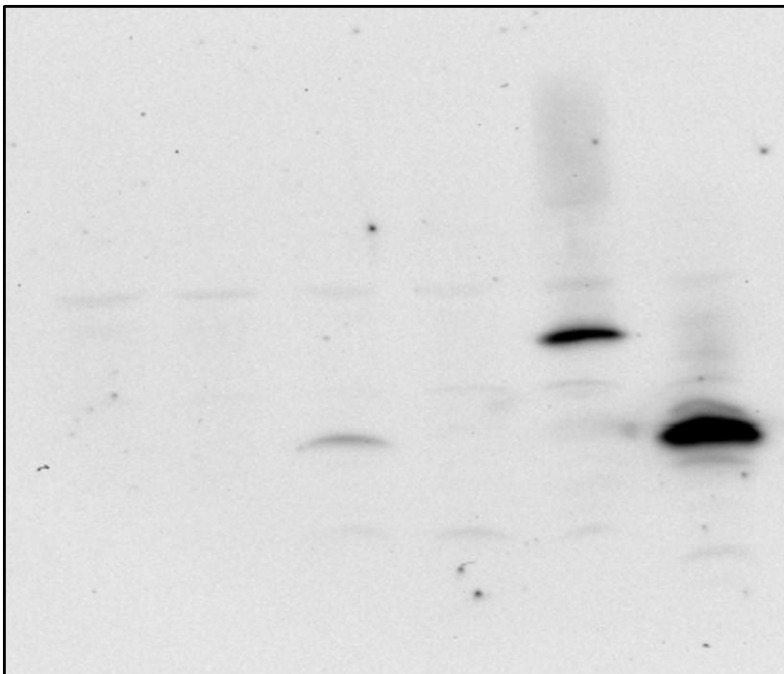
132



Supplementary Figure 11. Uncropped northern blot image corresponding to the Figure 8b.



Supplementary Figure 12. Uncropped SDS-PAGE image corresponding to the
Supplementary Figure 2a.



Supplementary Figure 13. Uncropped western blot image corresponding to the
Supplementary Figure 2b.

Supplementary references

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