

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- ☐ ☒ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- ☐ ☒ A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- ☐ ☒ The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- ☐ ☒ A description of all covariates tested
- ☐ ☒ A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- ☐ ☒ A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- ☐ ☒ For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- ☒ ☐ For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- ☒ ☐ For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- ☒ ☐ Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection -

Data analysis -

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

All figures listed have associated raw data: microscopy and immunoblotting images, and data for graphs supporting the results presented in this study are available in the San Raffaele Open Research Data Repository (ORDR, <https://ordr.hsr.it/research-data/>) with the DOI: 10.17632/wvt7kgsjvx.1

Human research participants

Policy information about [studies involving human research participants and Sex and Gender in Research](#).

Reporting on sex and gender	-
Population characteristics	-
Recruitment	-
Ethics oversight	-

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

☒ Life sciences ☐ Behavioural & social sciences ☐ Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	Sample sizes were based on previous experience with similar analysis, to achieve statistically significant differences.
Data exclusions	Generally no data were excluded. If data were excluded, this was specified in the manuscript.
Replication	Data were confirmed by replicating experiments successfully.
Randomization	Images from different experimental conditions were collected randomly, starting from one side of each coverslip (for immunofluorescence) until the established number of images was reached.
Blinding	Blinding was not relevant; images were collected randomly and quantitative analysis was based on objective parameters.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

Methods

n/a	Involved in the study	n/a	Involved in the study
☐	☒ Antibodies	☒	☐ ChIP-seq
☐	☒ Eukaryotic cell lines	☒	☐ Flow cytometry
☒	☐ Palaeontology and archaeology	☒	☐ MRI-based neuroimaging
☒	☐ Animals and other organisms		
☒	☐ Clinical data		
☒	☐ Dual use research of concern		

Antibodies

Antibodies used						
PP2A-C	Purified anti-PP2A catalytic α , clone 46	BD Transduction Laboratories™	610556	Monoclonal	Mouse	
	Recognizes methylated and and non-methylated form					
WB 1:5000						
PP2A-C	Anti-demethylated-PP2A-C,					
Clone 4B7	Santa Cruz Biotechnology	sc-13601	Monoclonal	Mouse	Specific for non-methylated form	
WB 1:1000						
PP2A-C	Anti-PP2A-C α / β , Clone 1D6	Santa Cruz Biotechnology	sc-13601	Monoclonal	Mouse	

Preferential recognition of non- methylated form									
WB 1:1000									
PP2A-C	Anti-PP2A alpha	GeneTex	GTX106334	Polyclonal	Rabbit	Specific for non-methylated form			
WB 1:5000									
B55α	Anti-B55α								
Clone 2G9	Cell Signaling	5689	Monoclonal	Mouse	WB 1:1000				
B56α	Anti-B56α								
Clone F-10	Santa Cruz sc-271151	Monoclonal		Mouse	WB 1:100				
B56γ	Anti-PP2A-B56γ, Clone E-6		Santa Cruz Biotechnology			sc-374380	Monoclonal	Mouse	WB
1:100-500									
PP2A-A	Anti-PP2A-Aα/β Clone 4G7		Santa Cruz Biotechnology			sc-13600	Monoclonal	Mouse	WB 1:250
Calnexin	Purified Mouse Anti-Calnexin		BD Transduction Laboratories™			610523	Monoclonal	Mouse	WB
1:2000									
Calnexin	Anti-Calnexin antibody produced in rabbit			Sigma					
	C4731	Polyclonal	Rabbit	WB 1:10000					
ERC1	Anti-ERC1 [ELKS-30] Against residues 21-40 of ERC1a			Abcam					
	ab50312	Monoclonal	Mouse	WB 1:1000					
ERC1	Anti-ERC1	Sigma-Aldrich	HPA019513	Polyclonal	Rabbit	IF 1:150			
FLAG	Monoclonal ANTI-FLAG® M2, clone M2			Sigma-Aldrich	F1804	Monoclonal	Mouse	WB	
1:1000									
IF 1:500									
GFP	GFP Polyclonal Antibody		Invitrogen	A11122	Polyclonal	Rabbit	WB 1:2000		
IP 2 μg									
GFP	Anti-GFP antibody	Abcam	ab13970	Polyclonal	Chicken	IF 1:1000			
Lamins	Anti-Lamin A + Lamin B1+ Lamin C			Abcam	Ab108922	Monoclonal	Rabbit	WB 1:5000	
Liprin-α1	Anti-liprin-α1 (A-5)	Santa Cruz Biotechnology			sc-376141	Monoclonal	Mouse	IP 0.5 μg	
IF 1:50									
Liprin-α1	Anti-liprin-α1	Proteintech	14175-1-AP		Polyclonal	Rabbit	WB 1:500		
IF 1:150									
Liprin-α1	Anti-liprin-α1; antigen: residues 818-1202 [6]			Polyclonal	Rabbit	WB 1:500			
Paxillin	Purified Mouse Anti-Paxillin		BD Transduction Laboratories™			610052	Monoclonal	Mouse	WB
1:2000									
IF 1:150									
IP 2 μg									
Paxillin	Paxillin antibody	GeneTex	GTX125891	Polyclonal	Rabbit	IF 1:200			
Src	Clone 327 from S. Courtneidge [7]			Monoclonal	Mouse	IF 1:50			
pSrc	Phospho-Src Family (Tyr416)		Cell Signaling Technology			#2101	Polyclonal	Rabbit	IF 1:100
Tubulin	Monoclonal anti-α-Tubulin		Sigma-Aldrich	T9026	Monoclonal		Mouse	WB 1:4000	
Vinculin	Upstate		Monoclonal	Mouse					
See complete list and details in the Methods section of the manuscript									

Validation

When necessary, validation of antibodies including proper controls is described in the manuscript, or reference to previous characterization is provided.

Eukaryotic cell lines

Policy information about [cell lines and Sex and Gender in Research](#)

Cell line source(s)

COS7, MCF-7, NIH-3T3, HeLa, BT-474, SK-BR-3 and MDA-MB-231 from ATCC.

Authentication

MDA-MB-231 were either recently obtained from ATCC, or validated by IdentiCell STR allele report.

Mycoplasma contamination

We confirm that all cell lines used were negative for mycoplasma contamination.

Commonly misidentified lines
(See [ICLAC](#) register)

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