

Supplementary Materials for

Sporopollenin-inspired design and synthesis of robust polymeric materials

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Supplementary Methods

All reactions were performed under nitrogen unless otherwise noted. 1,8-octanediol (TCI, >99%), 1,10-decanediol (TCI, >95%), 1,12-dodecanediol (Acrös, 99%), 1,14-tetradecanediol (Combi-Blocks, 97%), 1,16-hexadecanediol (TCI, >95%), Dess-Martin periodinane (Oakwood, 95%), polyvinyl alcohol ($M_{w,avg}$ = 31-50k, Sigma Aldrich, 98–99% hydrolyzed), polyvinyl alcohol ($M_{w,avg}$ = 85-124k, Sigma Aldrich, >99% hydrolyzed), polyvinyl alcohol ($M_{w,avg}$ = 89-98k, Sigma Aldrich, >99% hydrolyzed), polyvinyl alcohol ($M_{w,avg}$ = 146-186k, Sigma Aldrich >99% hydrolyzed), cyclohexanone (Acrös, >99%), cyclohexylamine (TCI, >99%), *n*-butyllithium (Acrös, 2.45 M), diisopropylamine (Spectrum, >99%), 11-bromo-1-undecene (Oakwood, 95%), sodium (meta)periodate (BTC, 98%), potassium permanganate (J.T. Baker, 99.4%), methyl iodide (Alfa Aesar, >98%), potassium carbonate (Sigma Aldrich, 99%), *meta*-chloroperbenzoic acid (Sigma Aldrich, <77%), sodium methoxide (Acrös, >99%), 4-dimethylaminopyridine (Acrös, 99%), *N,N'*-dicyclohexylcarbodiimide (TCI, >98%), lithium borohydride (Strem, 95%), tetrabutylammonium fluoride (Oakwood, 1 M in THF), triethylamine (Acrös, 99%), sulfur trioxide pyridine complex (TCI, >95%), coumaric acid (TCI, >98%), *tert*-butyldimethylsilyl chloride (Chem-Impex, 99%), imidazole (Sigma Aldrich, 99%), ethane thiol (Sigma Aldrich, 97%), boron trifluoride diethyl etherate (Sigma Aldrich, >46.5% BF₃ basis), acetic acid (Spectrum, >99.7%), concentrated sulfuric acid (EMD Millipore, 95%), 4 Å molecular sieves (Sigma Aldrich), ammonium chloride (Mallinckrodt, 99.5%), sodium sulfate (Sigma Aldrich, >99%), Celite (Fisher), sodium bicarbonate (Mallinckrodt, >99.7%), conc. hydrochloric acid (Macron, 36.5–38.0%), sodium hydroxide (AmericanBio, 97%), sodium chloride (EMD Millipore, 99.5%), dichloromethane (Fisher, >99.5%), dimethylsulfoxide (Fisher, >99.9%), diethyl ether (Fisher, >99.9%), tetrahydrofuran (Acrös, >99.5%, extra dry, stabilized with BHT), acetone (Fisher, >99.5%), water (distilled), petroleum ether (Fisher, 36–60 °C Fraction), hexanes (Fisher, 98.5%), ethyl acetate (Fisher, 99.5%), dimethylformamide (Acrös, 99.8%), chloroform (EMD Millipore, >99.8%), methanol (EMD Millipore, 99.9%), and dioxane (Acrös, 99.8%) were used as received without further purification unless otherwise noted. Column chromatography was conducted using Silicycle SiliaFlash P60 SiO₂ (40–63 µm). Preparative TLC was conducted using Millipore SiO₂ 60 F₂₅₄ PTLC (0.5 mm). Analytical TLC was conducting using Millipore SiO₂ 60 F₂₅₄ TLC (0.250 mm) plates. Silicone molds for polymer curing were obtained from MiniatureSweet (“Small Geometry Silicone Mold”).

Melting points were obtained using a Mel-Temp II apparatus in open capillaries and are uncorrected. Solution state ^1H and ^{13}C NMR spectra were obtained using either a Bruker Avance Neo 400 MHz spectrometer equipped with a 5 mm BBFO SmartProbe or a Bruker Avance Neo 500 MHz spectrometer equipped with a 5 mm liquid nitrogen cooled Prodigy BBO cryoprobe using either CDCl_3 (Cambridge Isotope Laboratories, 99.8%D) or $\text{DMSO}-d_6$ (Cambridge Isotope Laboratories, 99.9%D) as solvents. ^{13}C MAS ssNMR spectra were obtained on a Bruker Avance Neo 500 MHz spectrometer equipped with a 3.2 mm HX solids probe set to a MAS speed of 20 kHz. IR spectra were obtained using a Bruker Alpha 2 with a Platinum ATR accessory. High resolution mass spectrometric analysis was performed on either a JEOL AccuTOF-DART or a Thermo Fisher Scientific Q-Exactive benchtop Orbitrap. Durometer hardness testing was performed by Element Materials Technology with a Durometer Type D from Pacific Transducer Corporation. Thermogravimetric analysis was performed by Element Materials Technology with a TGA Q500 from TA Instruments. Differential scanning calorimetry was performed by Element Materials Technology with a DSC Q2000 from TA Instruments. Mechanical characterization was performed with a mechanical testing apparatus from Zwick/Roell company.

Supplementary Note 1: Sample Degree of Crosslinking (DOC) Calculation

Formula:

$$\frac{\text{mass PVA}}{1} \times \frac{\text{mol PVA}}{44 \text{ g PVA}} \times \frac{\text{mol - OH}}{\text{mol PVA}} \times \frac{\text{mol crosslinker}}{4 \text{ mol - OH}} \times \frac{\text{g}}{\text{mo}} \frac{\text{g crosslinker to achieve}}{100\% \text{ theoretical DOC}}$$

Sample Calculation:

If 1 mL of 5 wt% PVA in DMSO is to be 20% crosslinked by dialdehyde **15**:

$$0.05 \text{ g PVA} \times \frac{\text{mol PVA}}{44 \text{ g}} \times \frac{\text{mol crosslinker}}{4 \text{ mol - OH}} \times \frac{254.41 \text{ g crosslinker}}{\text{mol crosslinker}} \times 0.2 = 14.5 \text{ mg crosslinker}$$

Supplementary Tables

Table S1. DOC results for 50 mg PVA crosslinked by dialdehyde **15**

Theoretical DOC (%)	Mass of Linker (mg)
0	0.000
1	0.723
2	1.45
5	3.63
10	7.25
20	14.5
30	21.7
40	28.9
50	36.1

Table S2. Synthetic Sporopollenin Analogue Numbering Key

Sporopollenin Analogue #	6	7	8	9	10	11	12
Linker Carbon Count (#C)	16	16	16	8	10	12	14
Linker Density (%)	20	20	20	20	20	20	20
PVA MW _{avg} (g/mol)	166000	104500	40500	93500	93500	93500	93500
Sporopollenin Analogue #	13	14	15	16	17	18	26
Linker Carbon Count (#C)	16	16	16	16	16	16	16*
Linker Density (%)	5	10	20	30	40	50	20
PVA MW _{avg} (g/mol)	93500	93500	93500	93500	93500	93500	93500

* Indicates synthetic substituted crosslinker

Table S3. Chemical stability of synthetic sporopollenin polymer **15**

Solvent	Initial Mass (mg)	Final Mass (mg)	Mass Loss (%)	Notes:
CH ₂ Cl ₂	29.28	26.84	8.334	N/A
Hexanes	28.75	26.86	6.574	N/A
Ethyl Acetate	27.34	25.63	6.255	N/A
Acetone	27.51	26.08	5.198	N/A
DMF	27.61	30.00	-8.656	48 h dry time
DMSO	27.10	32.02	-18.155	48 h dry time
H ₂ O	24.81	22.57	9.029	N/A
THF	26.46	25.13	5.026	N/A
CHCl ₃	27.27	25.84	5.140	N/A
conc. H ₂ SO ₄	27.11	N/A	N/A	Blackened and crumbling
5 N NaOH	27.39	29.03	-5.988	48 h dry time, add'l mass likely due to retained NaOH in sample

Table S4. Durometer hardness (Shore D) of synthetic sporopollenin analogues

Sporopollenin Analogue #	6	7	8	9	10	11	12
Sample 1 (Shore D)	67	76	67	70	69	85	77
Sample 2 (Shore D)	69	77	69	68	69	84	75
Sample 3 (Shore D)	69	78	68	66	69	83	75
Sample 4 (Shore D)	70	77	---	62	71	84	---
Average (Shore D)	69	77	68	67	70	84	76
SD (Shore D)	1	1	1	3	1	1	1
Sporopollenin Analogue #	13	14	15	16	17	18	26
Sample 1 (Shore D)	77	68	67	71	65	66	70
Sample 2 (Shore D)	77	68	69	73	70	66	69
Sample 3 (Shore D)	73	67	70	72	65	65	75
Sample 4 (Shore D)	---	---	68	73	70	63	70
Average (Shore D)	76	68	69	72	68	65	71
SD (Shore D)	2	1	1	1	3	1	3

Overall:**Average (Shore D): 71****SD (Shore D): 5**

Table S5. Moduli (MPa) of synthetic sporopollenin analogues

Sporopollenin Analogue #	6	7	8	9	10	11	12
Sample 1 (MPa)	120	143	112	95	115	200	210
Sample 2 (MPa)	100	184	103	90	136	270	188
Sample 3 (MPa)	134	170	92	105	148	220	192
Average	118	166	102	97	133	230	197
SD	17	21	10	8	17	36	12
Sporopollenin Analogue #	13	14	15	16	17	18	26
Sample 1 (MPa)	158	105	156	157	120	86	90
Sample 2 (MPa)	111	107	160	135	106	100	86
Sample 3 (MPa)	131	87	175	116	118	95	116
Average	133	100	164	136	115	94	97
SD	24	11	10	21	8	7	16

Overall:**Average (MPa): 134****SD (MPa): 41**

Table S6. Hysteresis ratio (dimensionless) of synthetic sporopollenin analogues

Sporopollenin Analogue #	6	7	8	9	10	11	12
Sample 1	0.54	0.62	0.53	0.48	0.53	0.62	0.58
Sample 2	0.54	0.62	0.55	0.49	0.53	0.60	0.60
Sample 3	0.59	0.63	0.54	0.47	0.52	0.59	0.59
Average	0.56	0.63	0.54	0.48	0.52	0.60	0.59
SD	0.03	0.01	0.01	0.01	0.01	0.01	0.01
Sporopollenin Analogue #	13	14	15	16	17	18	26
Sample 1	0.58	0.56	0.63	0.55	0.55	0.55	0.57
Sample 2	0.58	0.57	0.63	0.55	0.54	0.56	0.56
Sample 3	0.60	0.55	0.64	0.57	0.55	0.56	0.58
Average	0.59	0.56	0.63	0.56	0.55	0.56	0.57
SD	0.01	0.01	0.01	0.01	0.01	0.01	0.01

Overall:**Average (dimensionless): 0.53****SD (dimensionless): 0.15**

Table S7. Volume swelling ratios (dimensionless) of synthetic sporopollenin analogues

Sporopollenin Analogue #	6	7	8	9	10	11	12
Sample 1	1.24	1.47	1.38	1.51	1.44	1.62	1.53
Sample 2	1.28	1.41	1.35	1.56	1.40	1.57	1.50
Sample 3	1.27	1.39	1.39	1.54	1.41	1.70	1.51
Average	1.26	1.43	1.37	1.54	1.42	1.63	1.51
SD	0.02	0.04	0.02	0.02	0.02	0.07	0.02
Sporopollenin Analogue #	13	14	15	16	17	18	26
Sample 1	2.11	1.78	1.69	1.18	1.15	1.21	1.42
Sample 2	1.97	1.66	1.73	1.17	1.10	1.16	1.28
Sample 3	1.93	1.76	1.66	1.12	1.28	1.18	1.48
Average	2.00	1.74	1.69	1.16	1.18	1.19	1.39
SD	0.10	0.06	0.03	0.03	0.09	0.03	0.10

Table S8. Weight swelling ratios (dimensionless) of synthetic sporopollenin analogues

Sporopollenin Analogue #	6	7	8	9	10	11	12
Sample 1	1.24	1.43	1.24	1.61	1.33	1.55	1.41
Sample 2	1.21	1.34	1.27	1.43	1.30	1.49	1.33
Sample 3	1.26	1.32	1.25	1.47	1.26	1.56	1.35
Average	1.23	1.37	1.25	1.50	1.30	1.54	1.37
SD	0.03	0.06	0.02	0.09	0.03	0.04	0.04
Sporopollenin Analogue #	13	14	15	16	17	18	26
Sample 1	1.74	1.57	1.59	1.09	1.06	1.09	1.34
Sample 2	1.76	1.57	1.58	1.11	0.95	1.08	1.26
Sample 3	1.65	1.57	1.53	1.09	1.04	1.13	1.28
Average	1.71	1.57	1.57	1.10	1.02	1.10	1.30
SD	0.06	0.00	0.03	0.01	0.06	0.02	0.04

Table S9. Thermogravimetric analysis of synthetic sporopollenin analogues

Sporopollenin Analogue #	6	7	8	9	10	11	12
Loss 1 (% , °C)	33.4, 208	33.3, 217	34.4, 210	38.1, 211	37.2, 215	35.0, 218	34.9, 226
Loss 2 (% , °C)	55.6, 454	55.7, 455	53.8, 455	49.3, 446	50.1, 451	50.8, 452	51.8, 451
Loss 3 (% , °C)	9.7, 540	9.6, 535	10.4, 542	11.0, 532	11.1, 530	11.4, 534	10.8, 519
Residue (%)	1.3	1.5	1.3	1.6	1.6	2.7	2.5
Sporopollenin Analogue #	13	14	15	16	17	18	26
Loss 1 (% , °C)	37.9, 205	37.0, 210	35.6, 203	32.5, 220	30.6, 226	31.6, 218	32.3, 189
Loss 2 (% , °C)	50.1, 446	50.6, 450	53.2, 444	56.8, 457	60.8, 464	61.7, 463	56.5, 450
Loss 3 (% , °C)	10.6, 535	10.7, 535	9.0, 522	9.6, 540	7.9, 544	6.0, 548	9.8, 530
Residue (%)	1.4	1.6	2.2	1.1	0.7	0.7	1.3

Overall:**Average Loss 1 (% , °C): 34.6, 213****SD (% , °C): 2.4, 9.8****Average Loss 2 (% , °C): 54.1, 453****SD (% , °C): 4.0, 5.9****Average Loss 3 (% , °C): 9.8, 535****SD (% , °C): 1.4, 8.0****Average Residue (%): 1.5****SD (%): 0.6**

Supplementary Figures

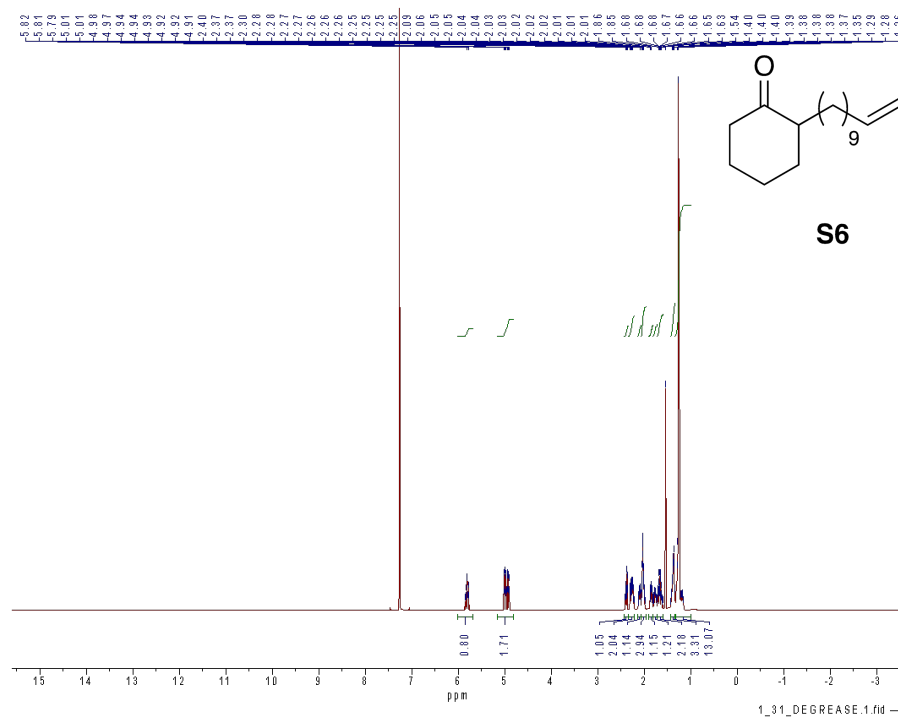


Figure S1. The ^1H NMR spectrum of compound S6

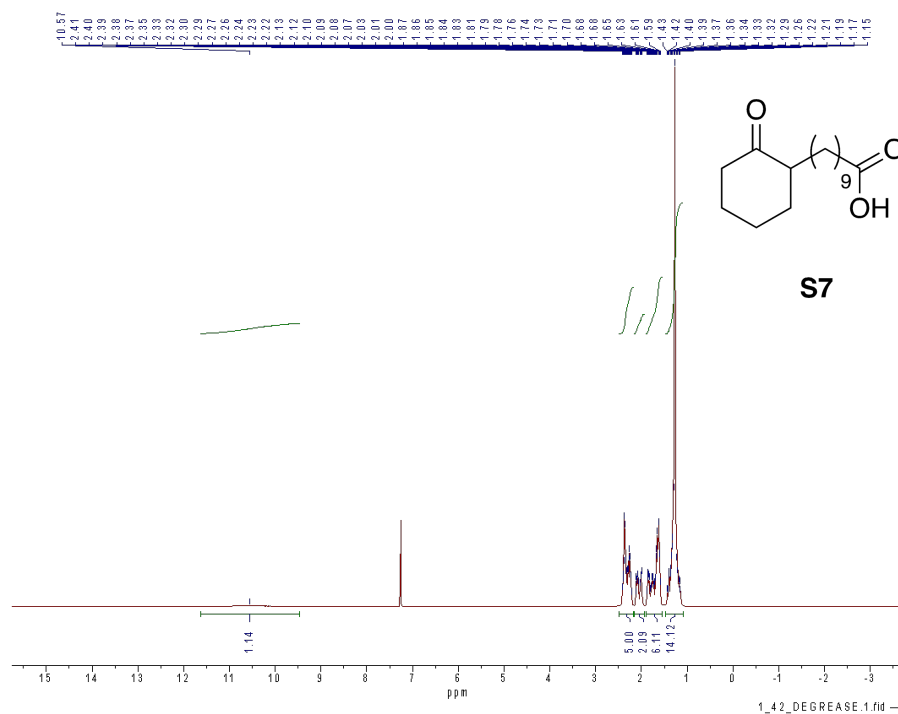


Figure S2. The ^1H NMR spectrum of compound S7

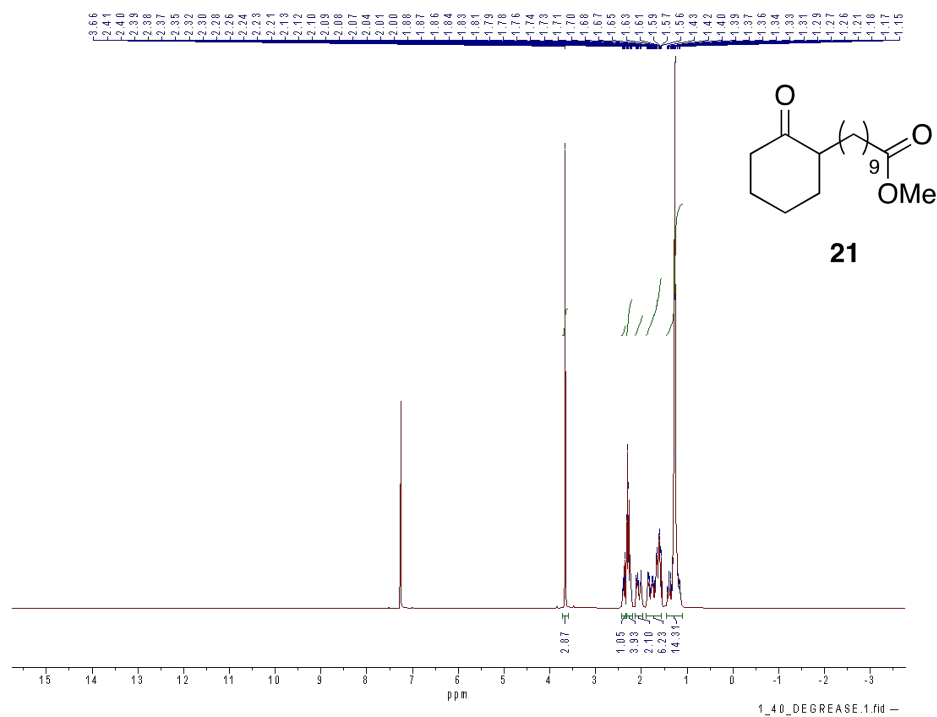


Figure S3. The ¹H NMR spectrum of compound **21**

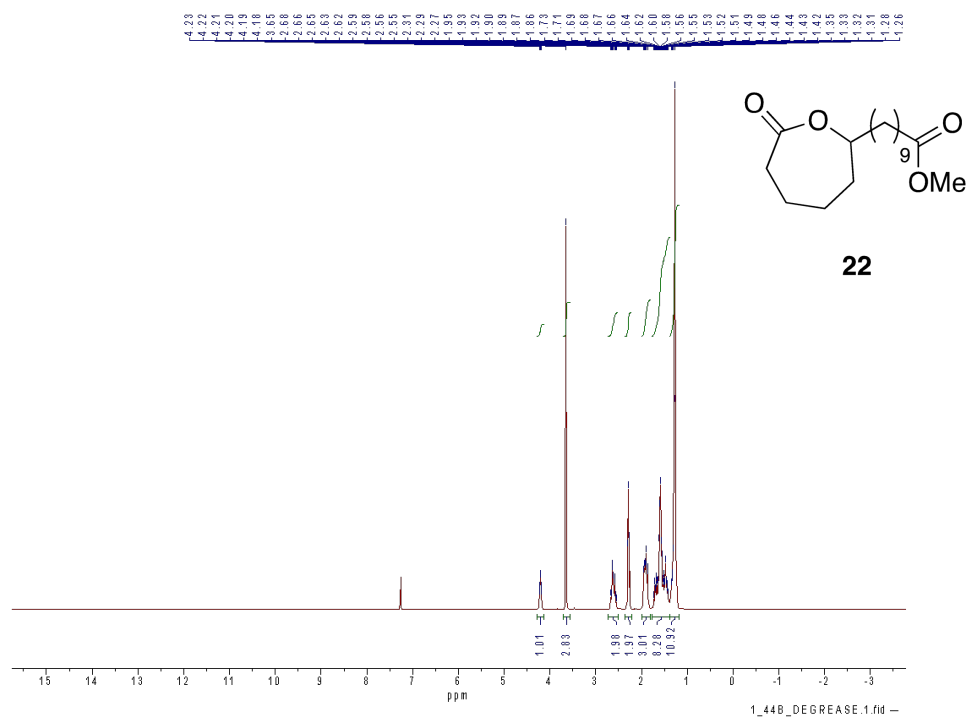


Figure S4. The ¹H NMR spectrum of compound **22**

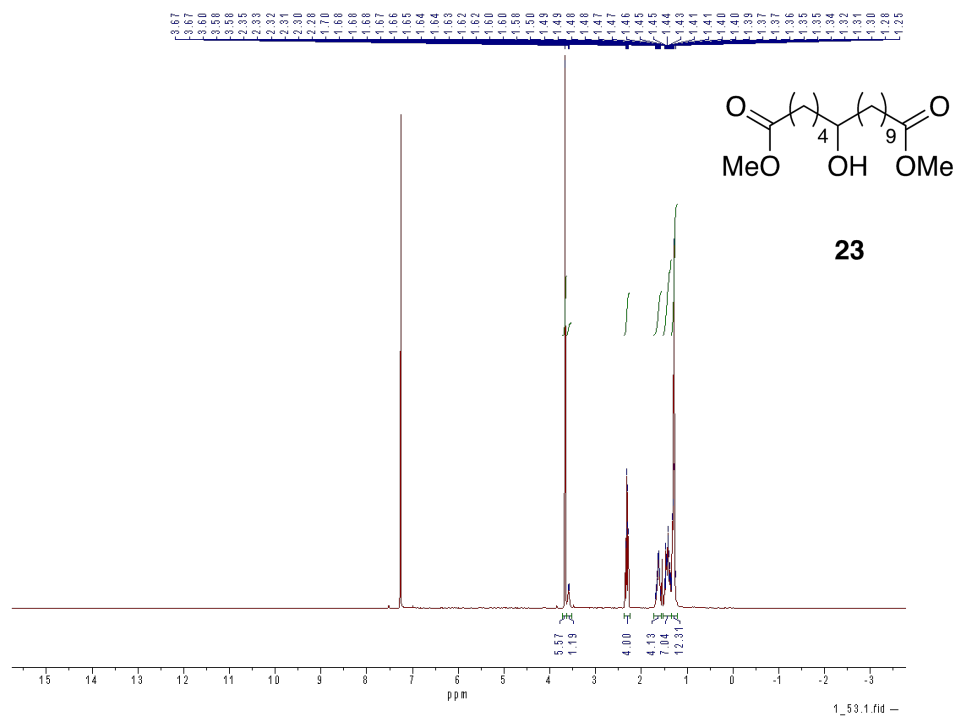


Figure S5. The ^1H NMR spectrum of compound **23**

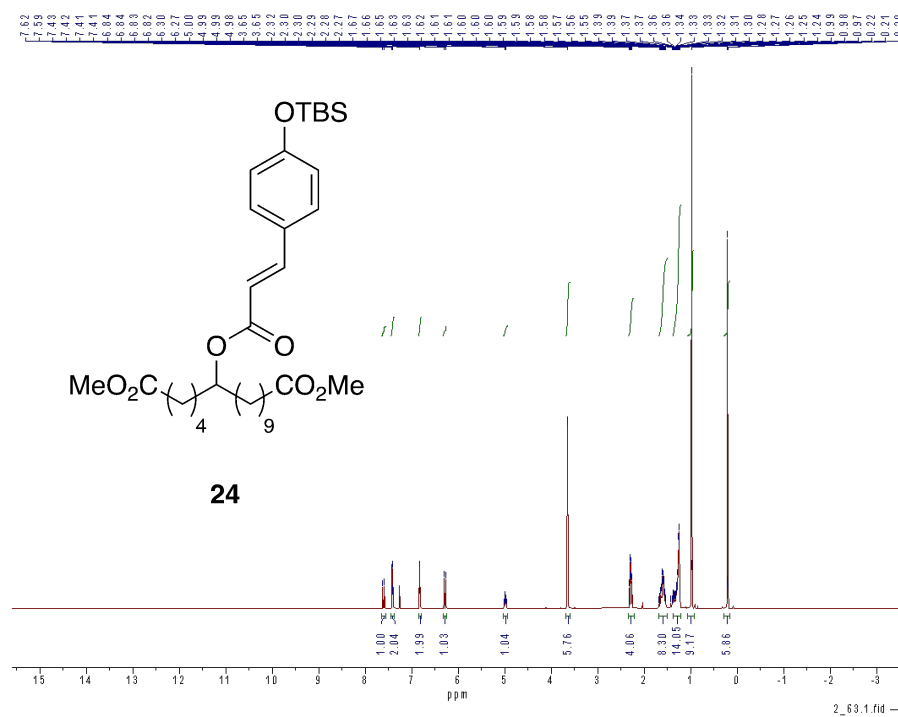


Figure S6. The ^1H NMR spectrum of compound **24**

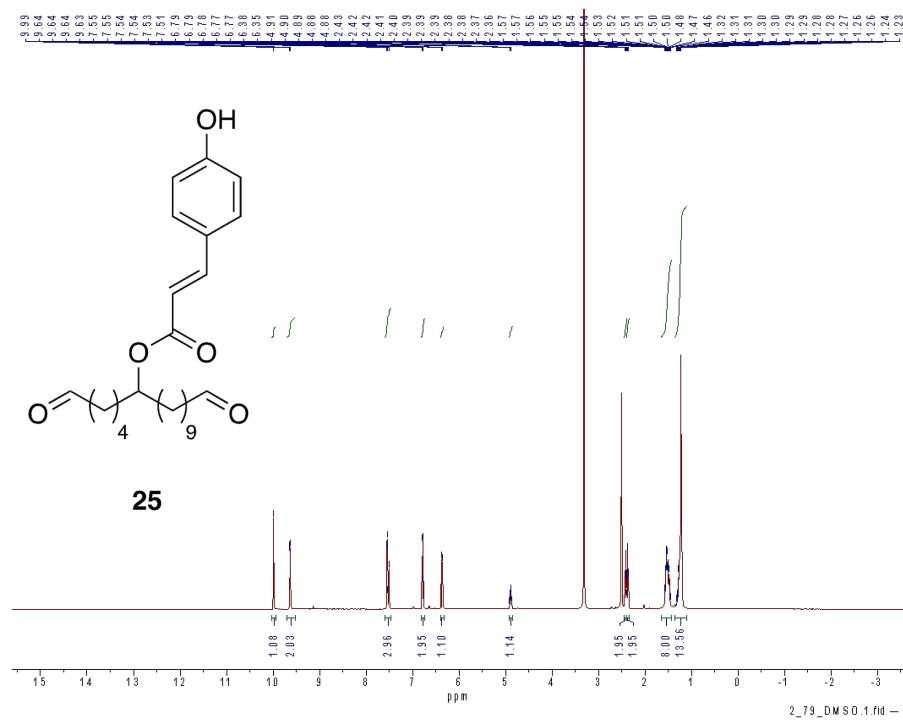


Figure S9. The ¹H NMR spectrum of compound **25**

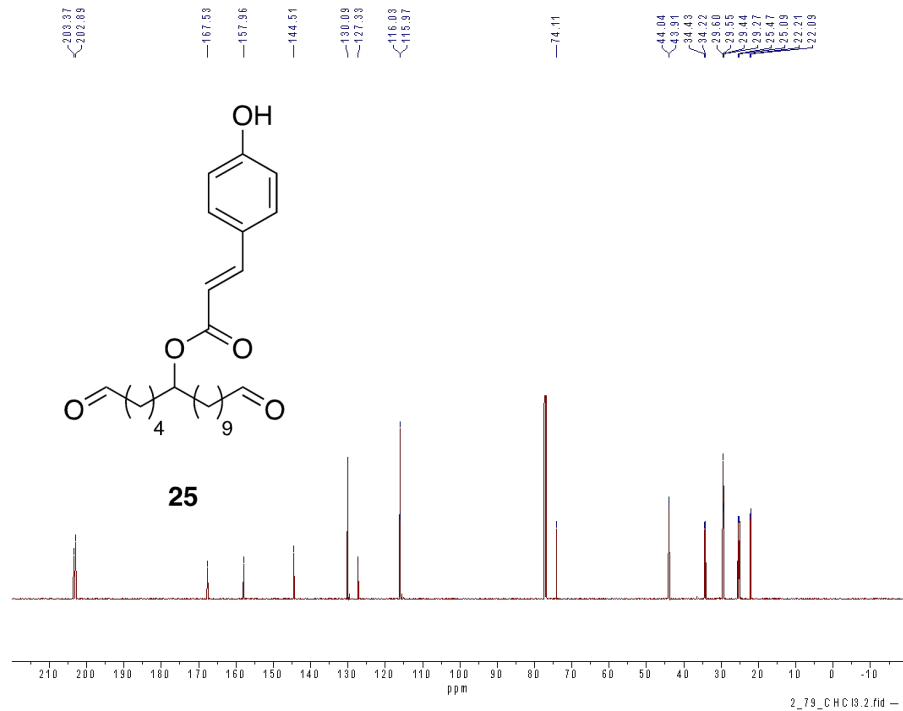


Figure S10. The ¹³C NMR spectrum of compound **25**

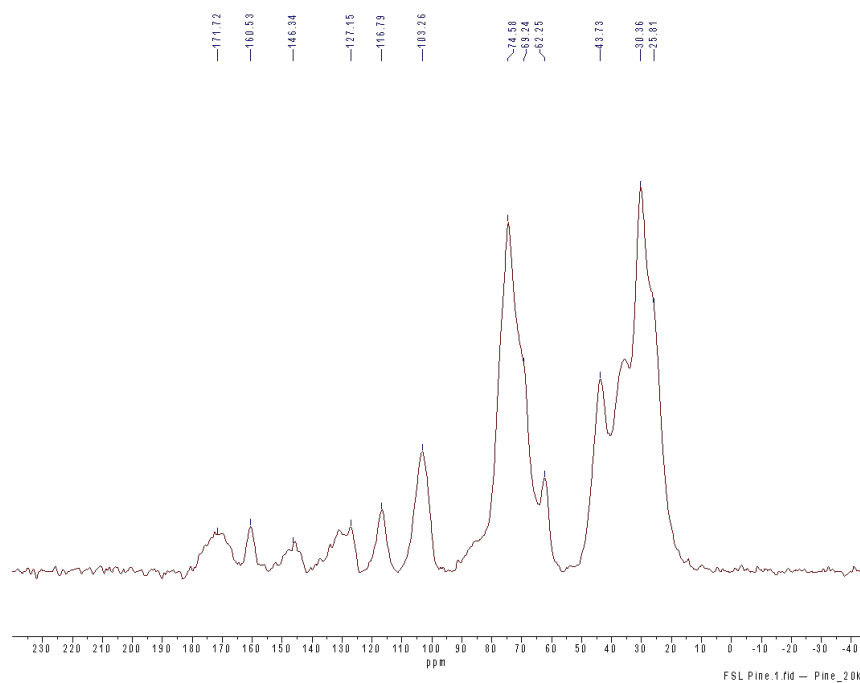


Figure S11. The ^{13}C MAS ssNMR spectrum of authentic *P. rigida* sporopollenin (**9**)

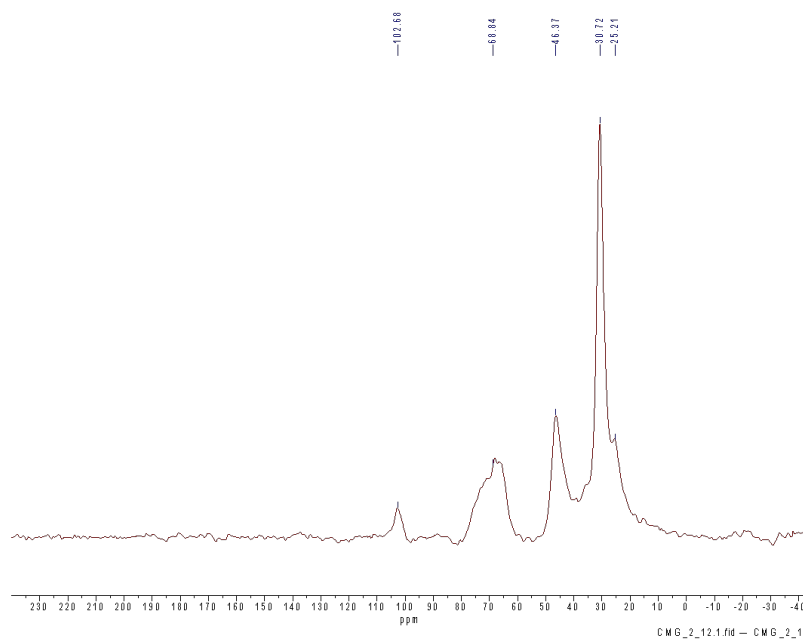


Figure S12. The ^{13}C MAS ssNMR spectrum of simplified sporopollenin analogue **15**

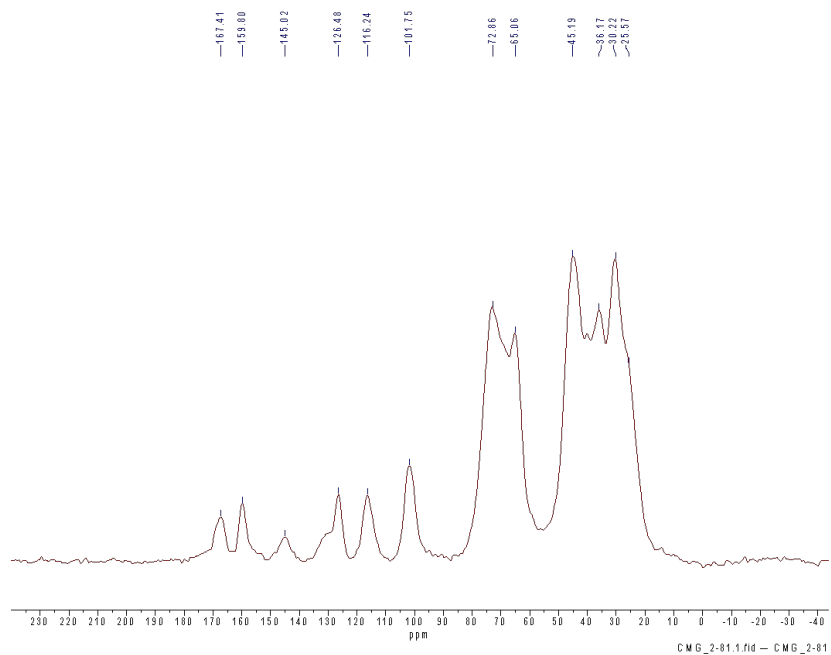


Figure S13. The ^{13}C MAS ssNMR spectrum of synthetic linker sporopollenin analogue **26**

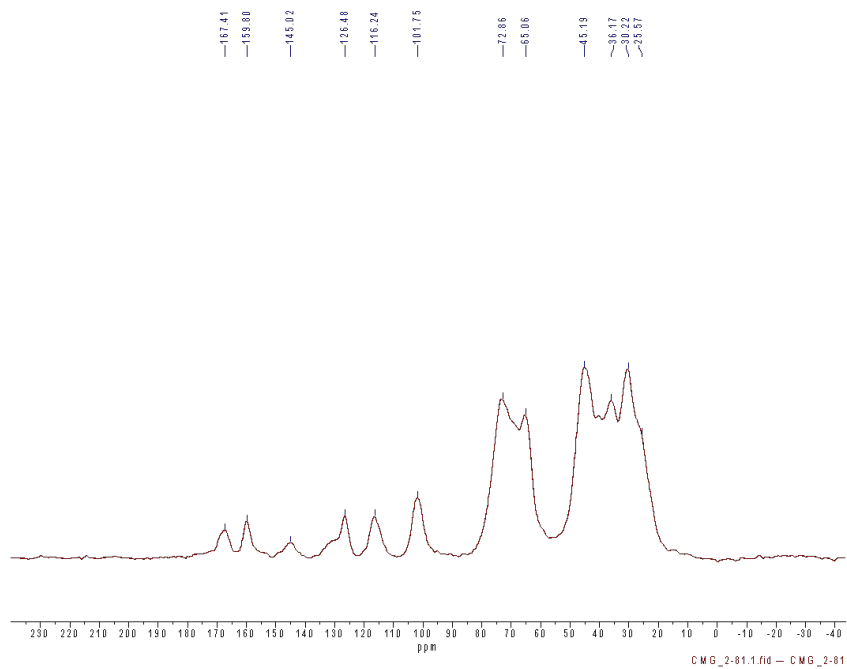


Figure S14. Overlaid ^{13}C MAS ssNMR spectra of authentic *P. rigida* sporopollenin (blue) (**9**), simplified sporopollenin analogue **15** (green), and synthetic linker sporopollenin analogue **26** (red)

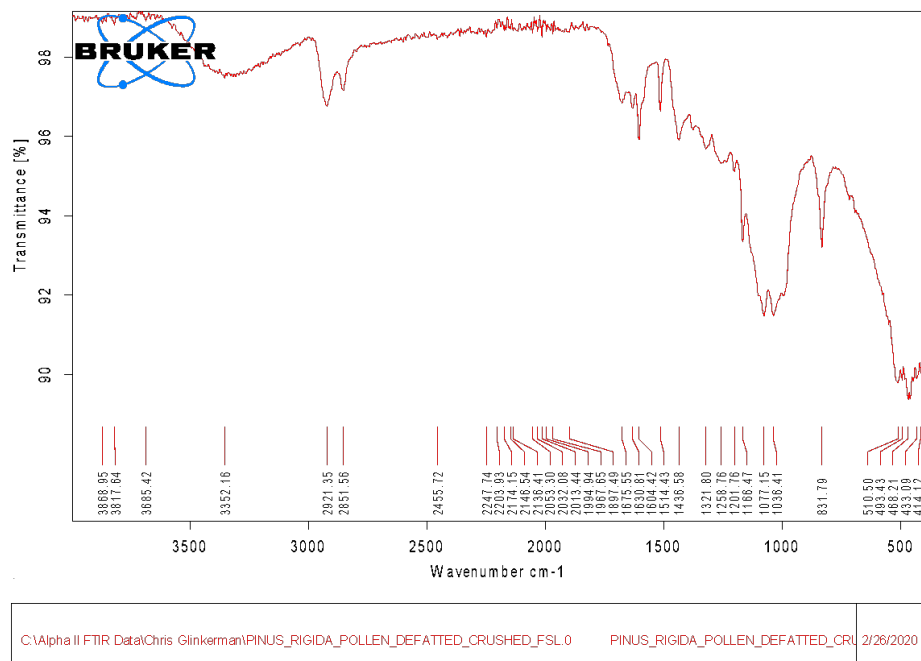


Figure S15. The IR spectrum of authentic *P. rigida* sporopollenin

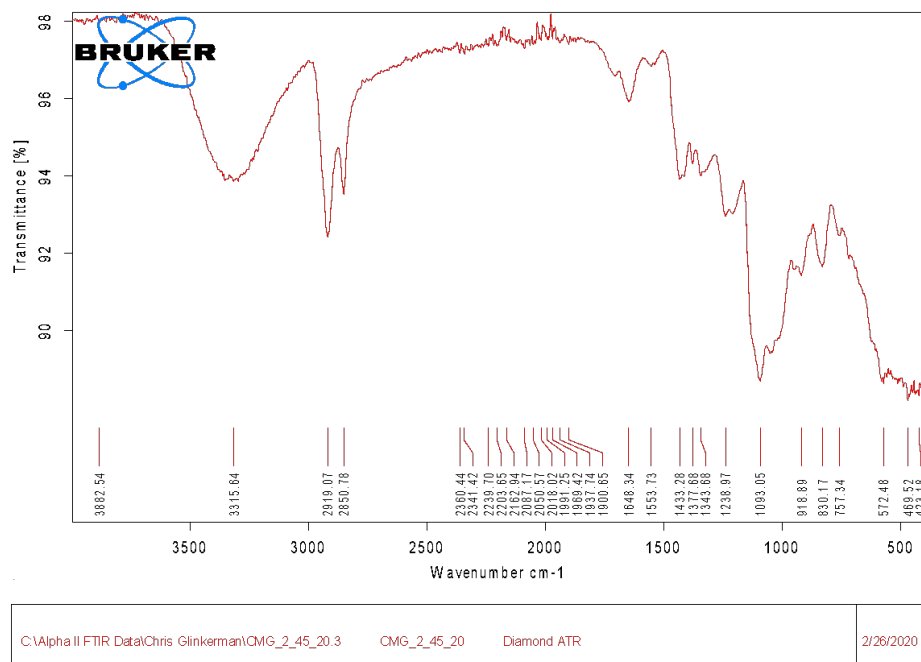


Figure S16. The IR spectrum of simplified sporopollenin analogue 15

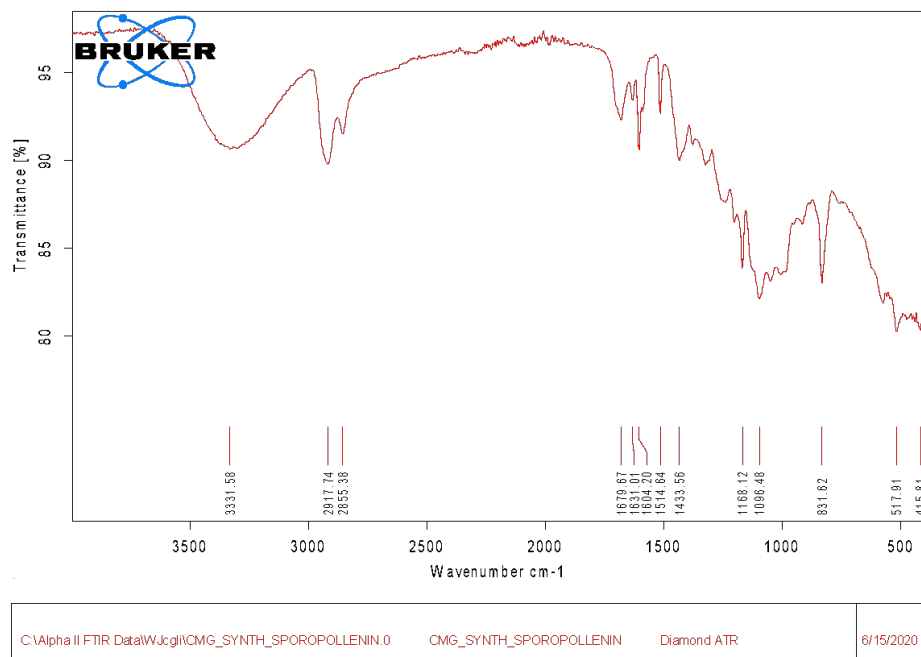


Figure S17. The IR spectrum of synthetic linker sporopollenin analogue **26**

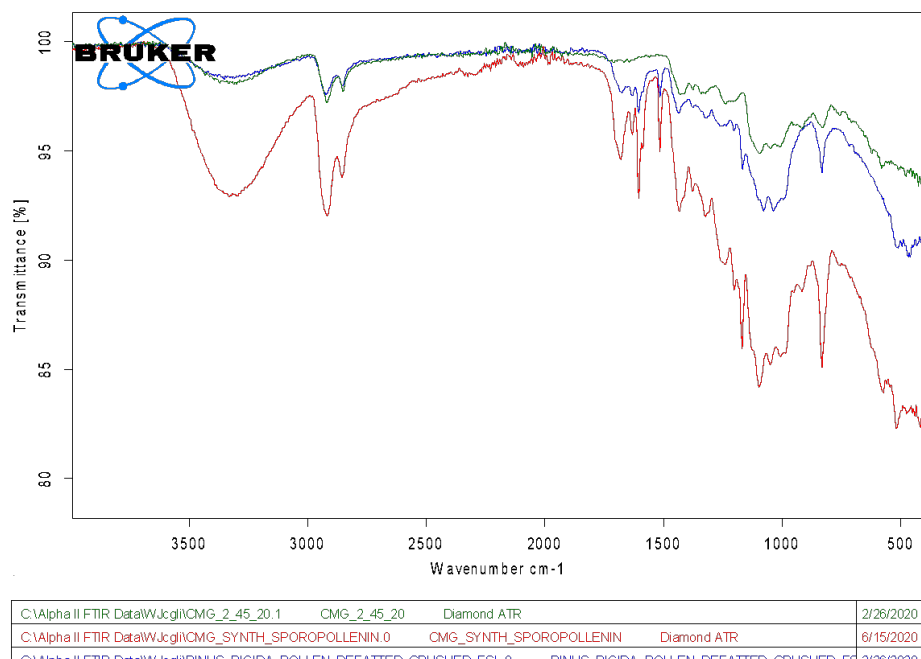


Figure S18. Overlaid IR spectra of authentic *P. rigida* sporopollenin (blue), simplified sporopollenin analogue **15** (green), and synthetic linker sporopollenin analogue **26** (red)

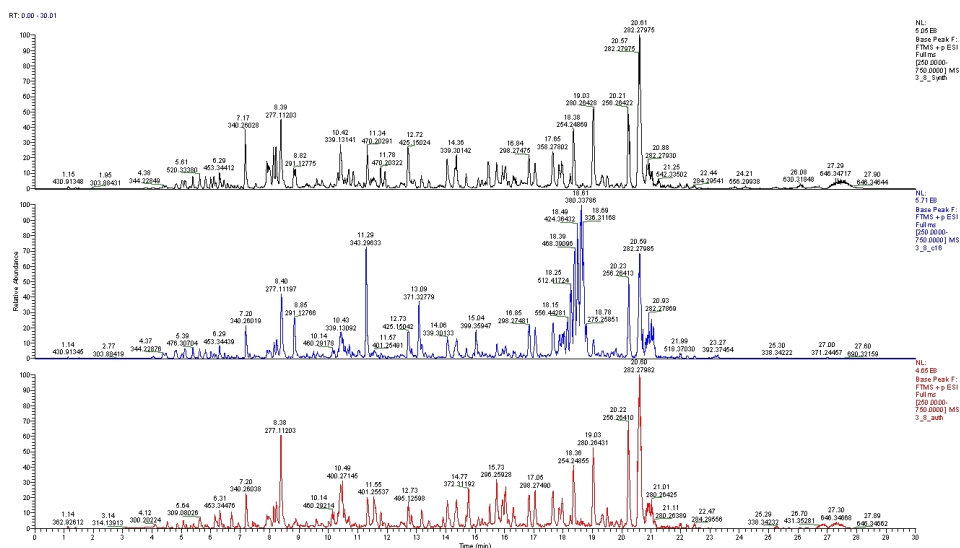


Figure S19. The base peak chromatograms of thioacidolyzed authentic *P. rigida* sporopollenin (red), simplified sporopollenin analogue **15** (blue), and synthetic linker sporopollenin analogue **26** (black)

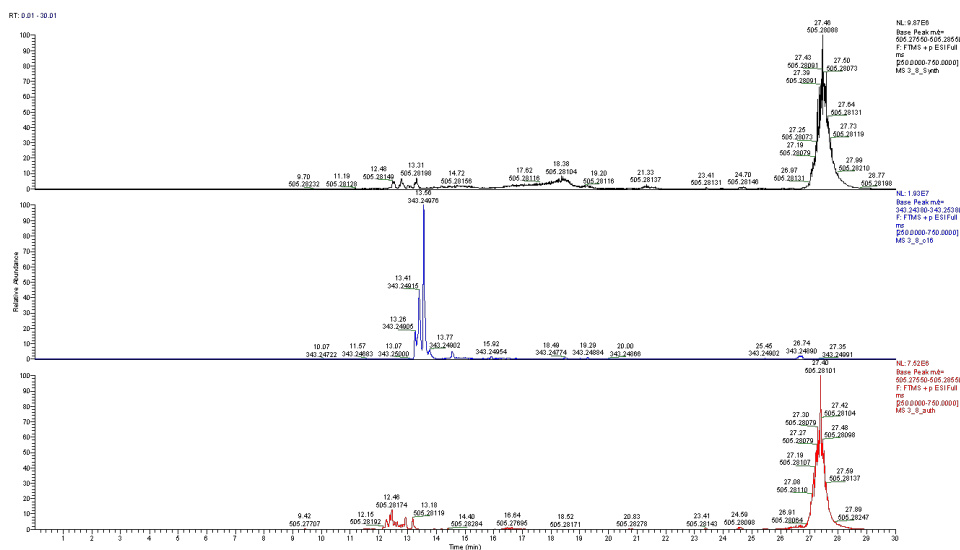


Figure S20. The extracted-ion chromatograms of thioacidolyzed authentic *P. rigida* sporopollenin (red), simplified sporopollenin analogue **15** (blue), and synthetic linker sporopollenin analogue **26** (black)

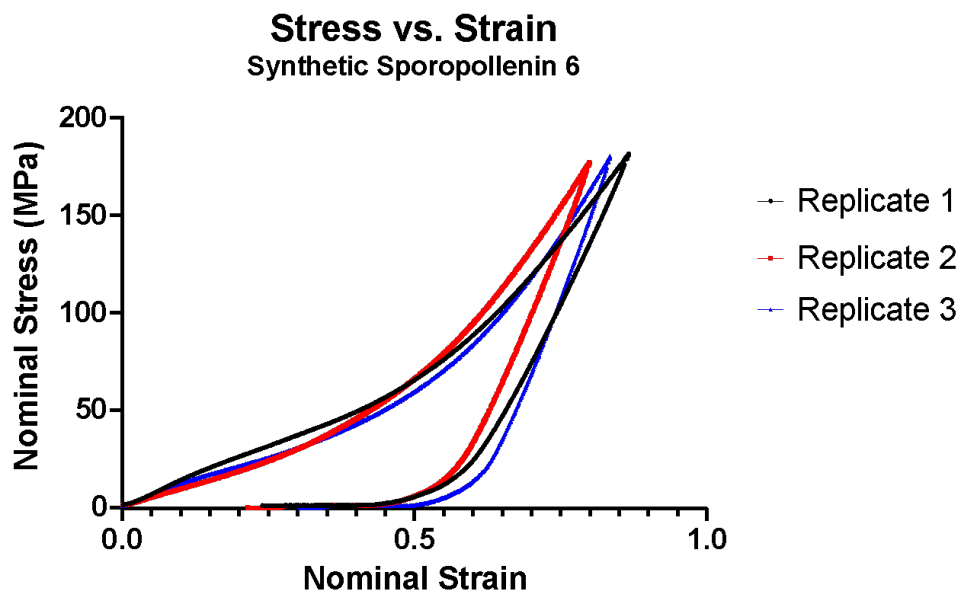


Figure S21. Compressive stress-strain curve for synthetic sporopollenin analogue 6

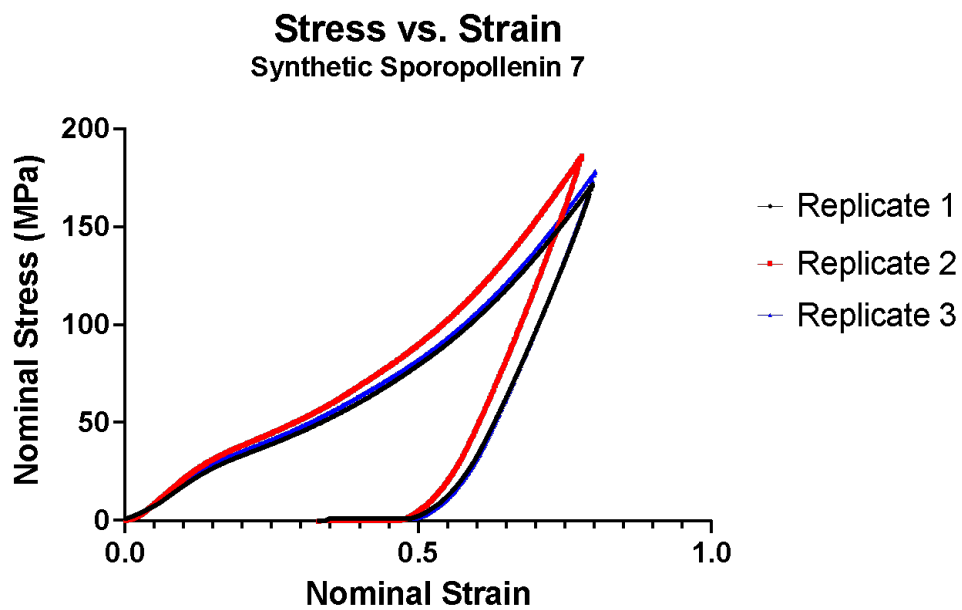


Figure S22. Compressive stress-strain curve for synthetic sporopollenin analogue 7

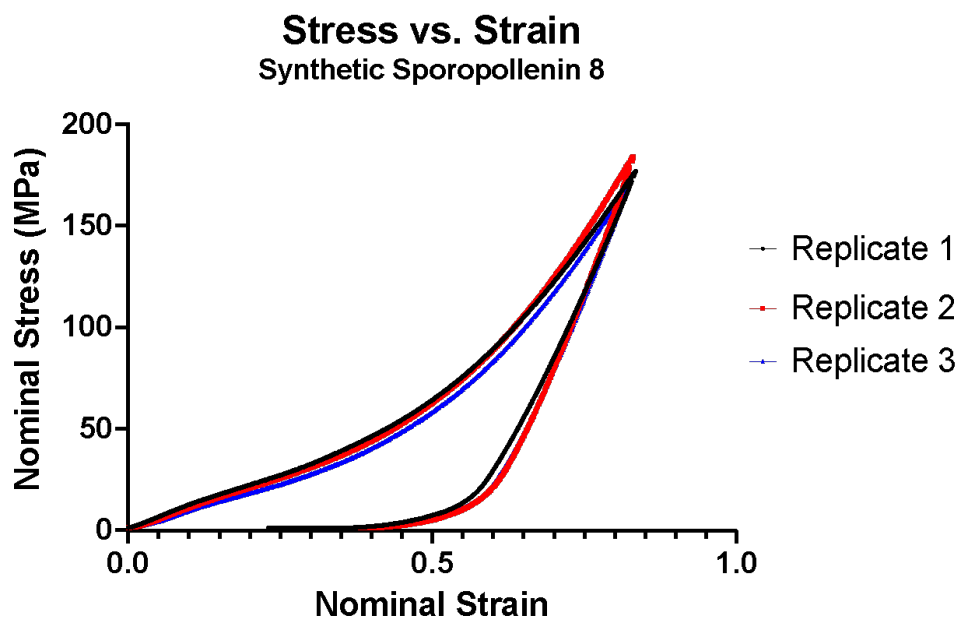


Figure S23. Compressive stress-strain curve for synthetic sporopollenin analogue 8

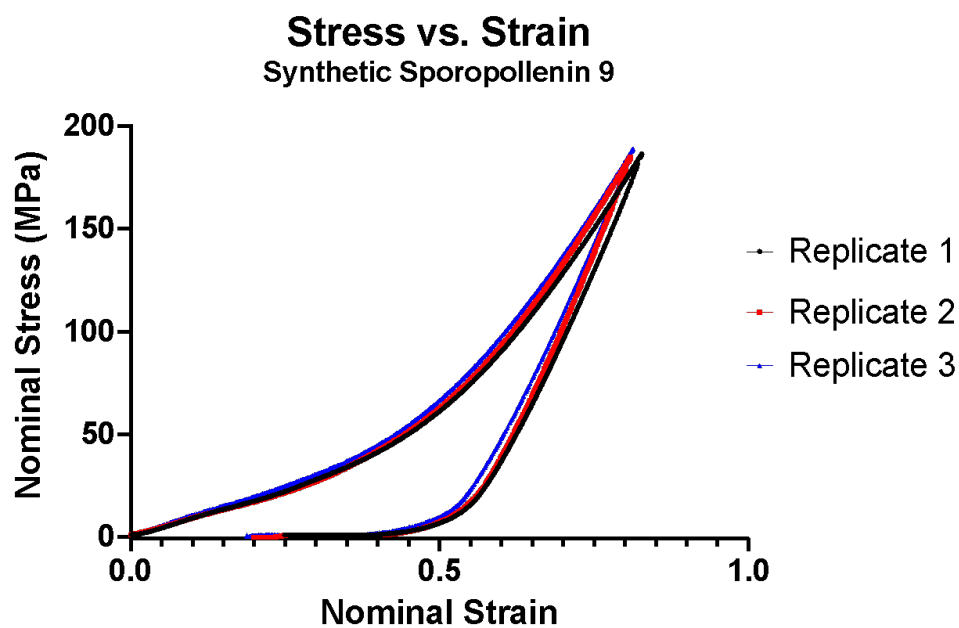


Figure S24. Compressive stress-strain curve for synthetic sporopollenin analogue 9

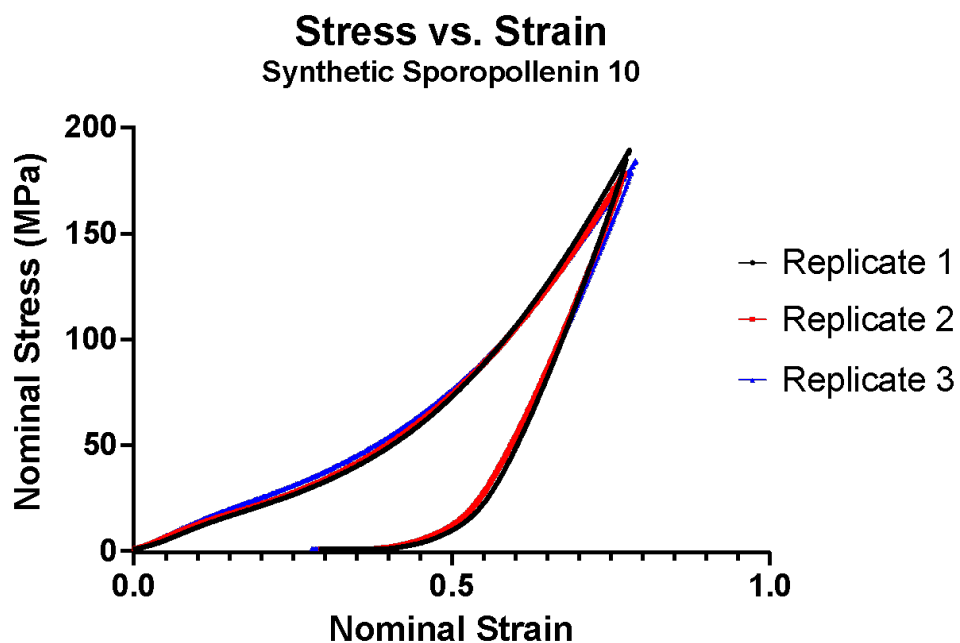


Figure S25. Compressive stress-strain curve for synthetic sporopollenin analogue **10**

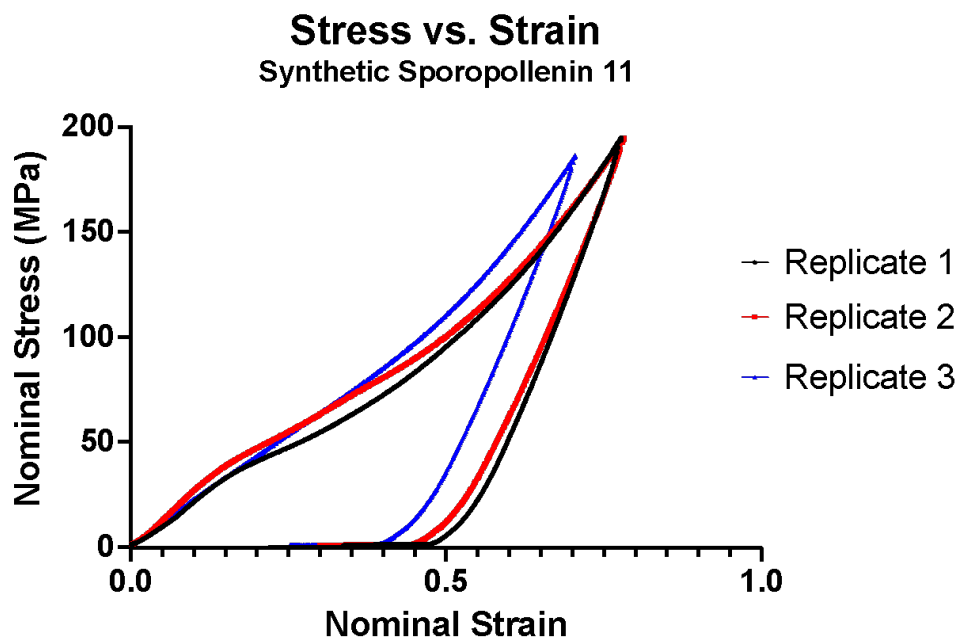


Figure S26. Compressive stress-strain curve for synthetic sporopollenin analogue **11**

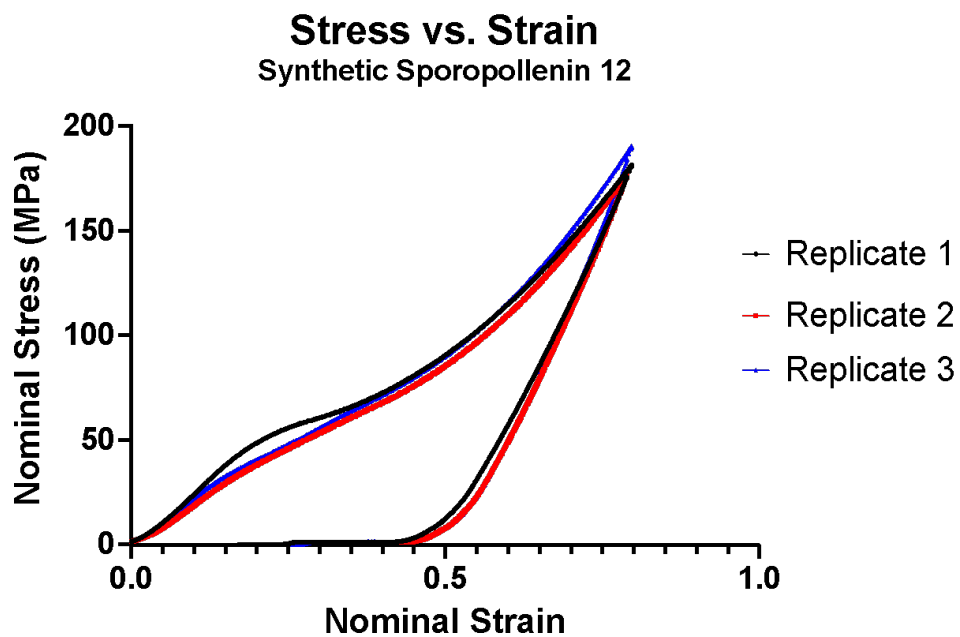


Figure S27. Compressive stress-strain curve for synthetic sporopollenin analogue 12

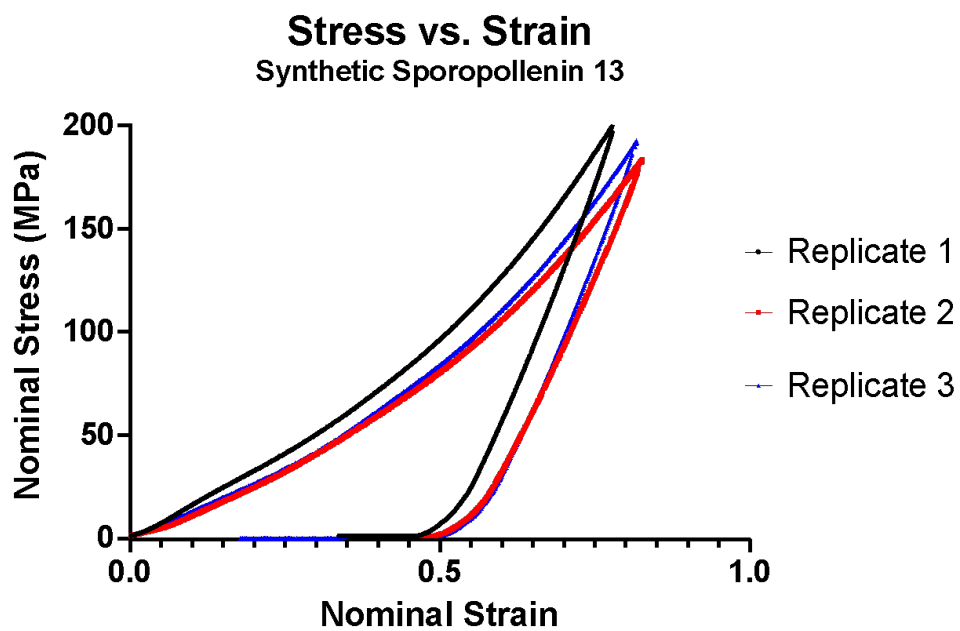


Figure S28. Compressive stress-strain curve for synthetic sporopollenin analogue 13

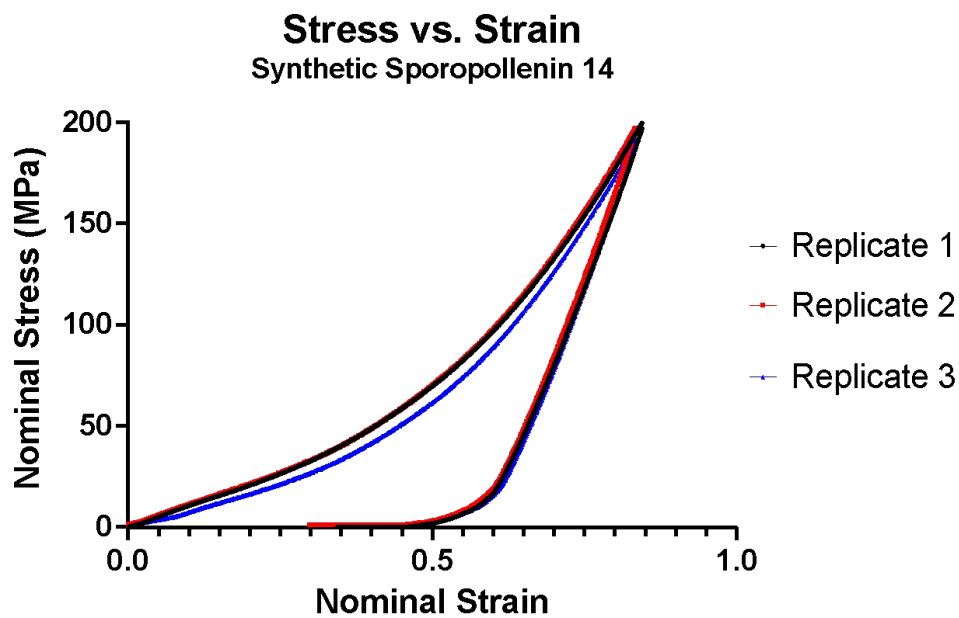


Figure S29. Compressive stress-strain curve for synthetic sporopollenin analogue **14**

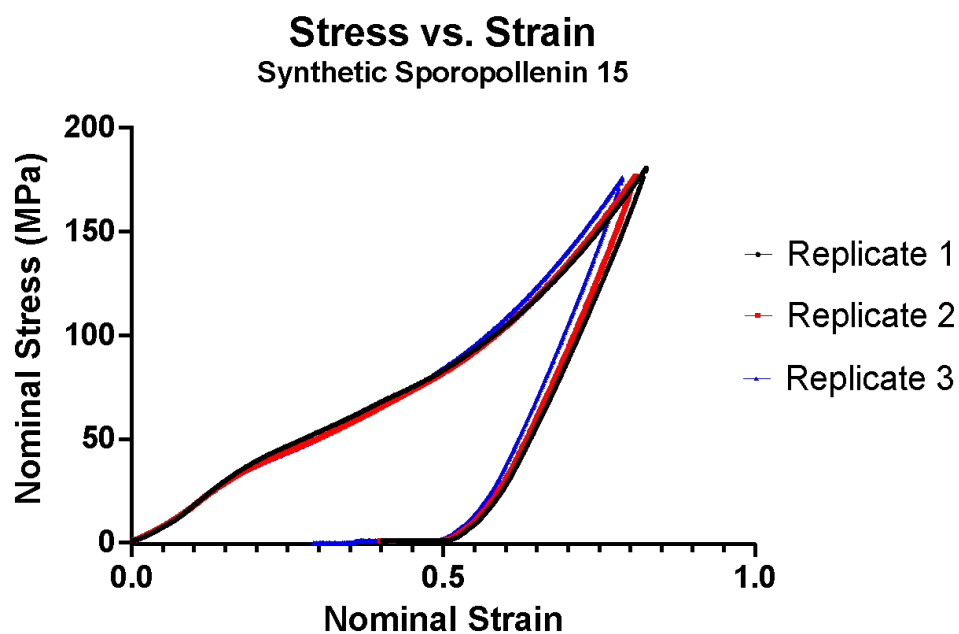


Figure S30. Compressive stress-strain curve for synthetic sporopollenin analogue **15**

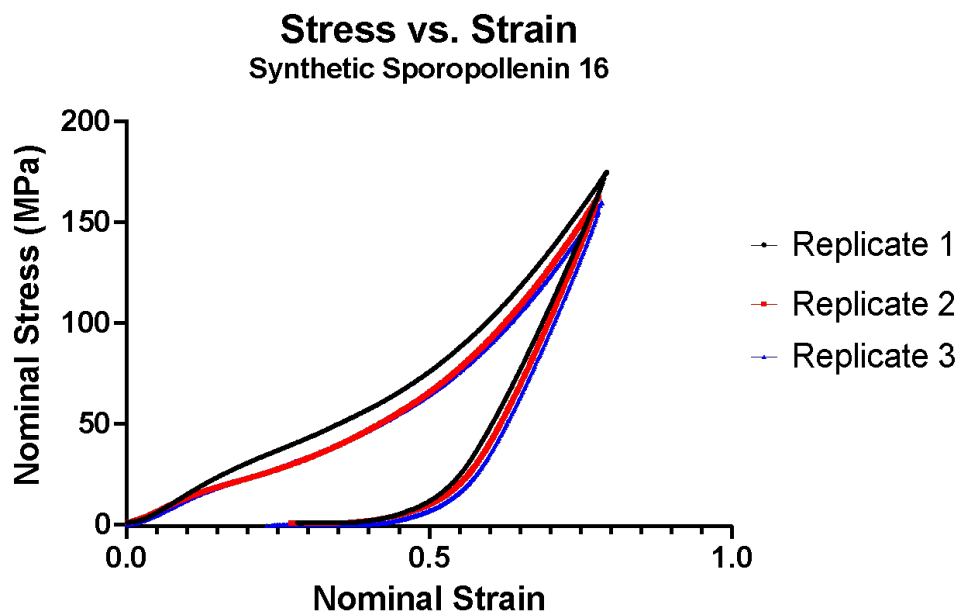


Figure S31. Compressive stress-strain curve for synthetic sporopollenin analogue 16

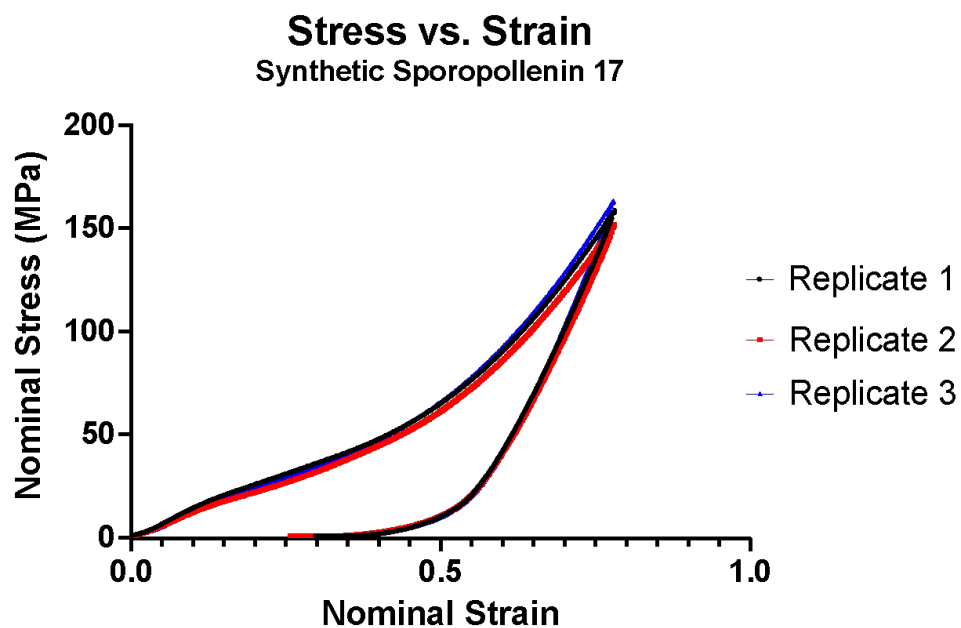


Figure S32. Compressive stress-strain curve for synthetic sporopollenin analogue 17

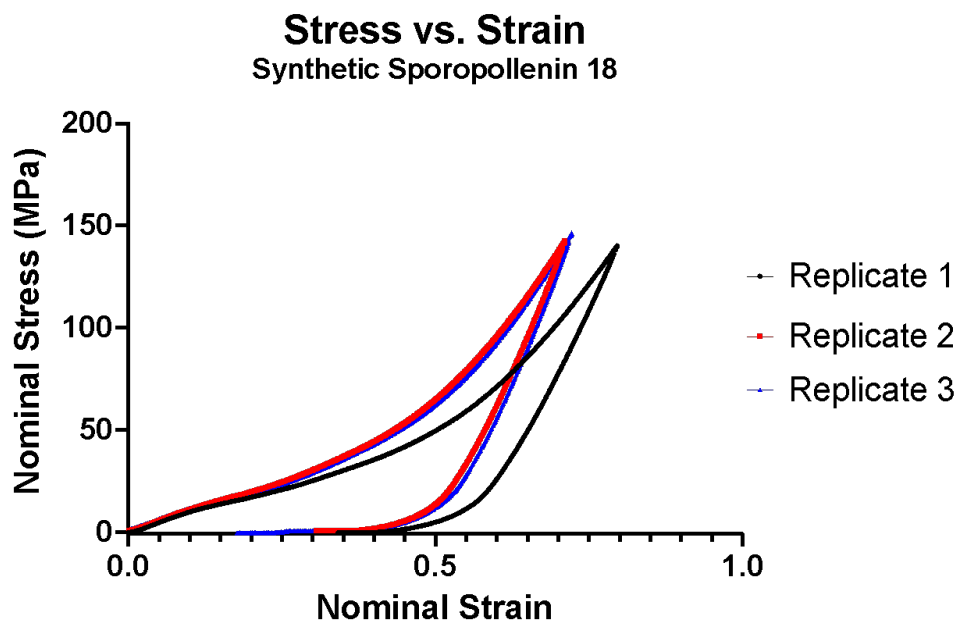


Figure S33. Compressive stress-strain curve for synthetic sporopollenin analogue 18

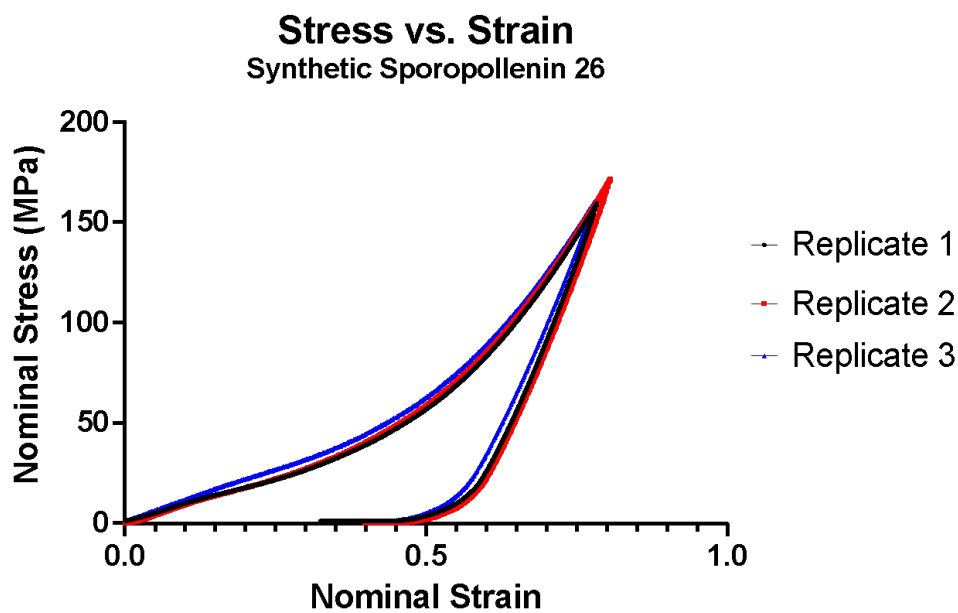


Figure S34. Compressive stress-strain curve for synthetic sporopollenin analogue 26

Modulus vs. Linker Carbon Count

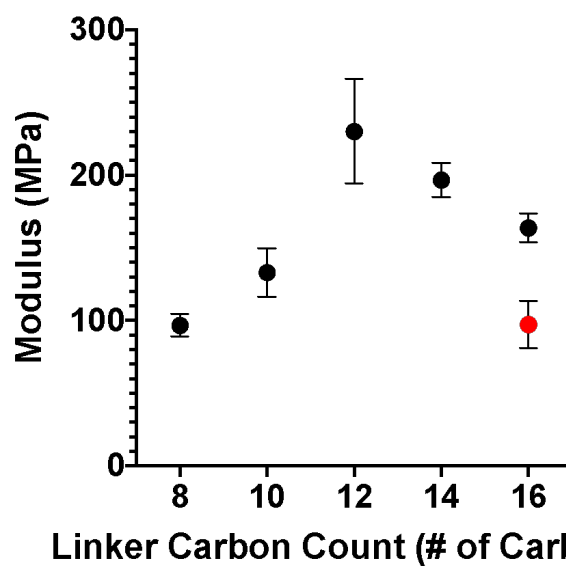


Figure S35. Relationship between compressive modulus and linker carbon count in simplified (black) and substituted (red) synthetic linker sporopollenin analogues (error bars = standard deviation)

Modulus vs. Linker Density

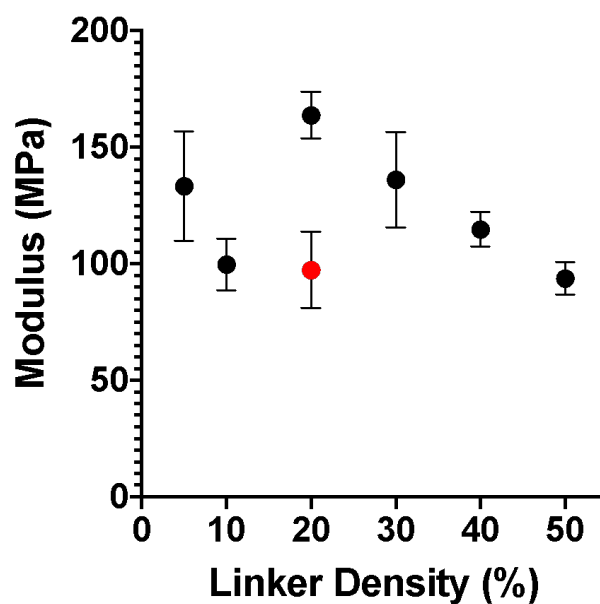


Figure S36. Relationship between compressive modulus and linker density in simplified (black) and substituted (red) synthetic linker sporopollenin analogues (error bars = standard deviation)

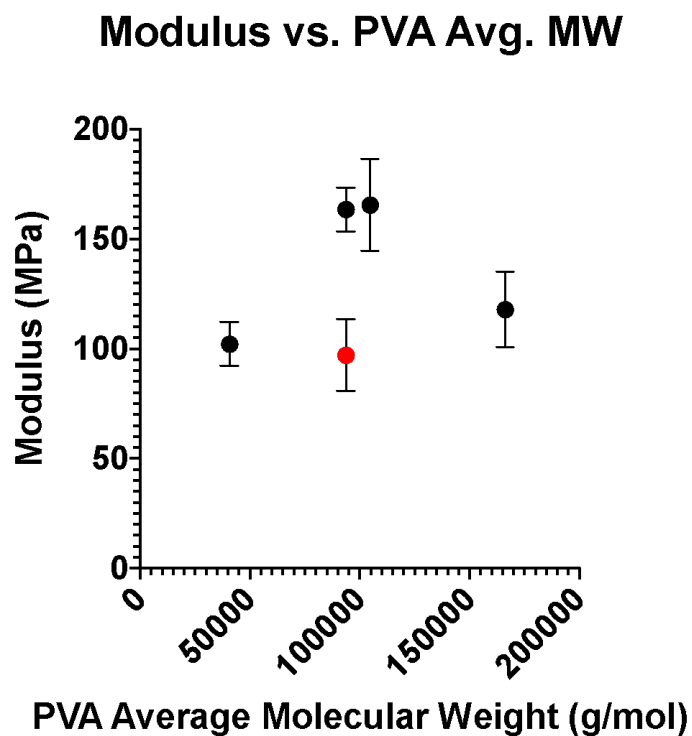


Figure S37. Relationship between compressive modulus and PVA MW_{avg} in simplified (black) and substituted (red) synthetic linker sporopollenin analogues (error bars = standard deviation)

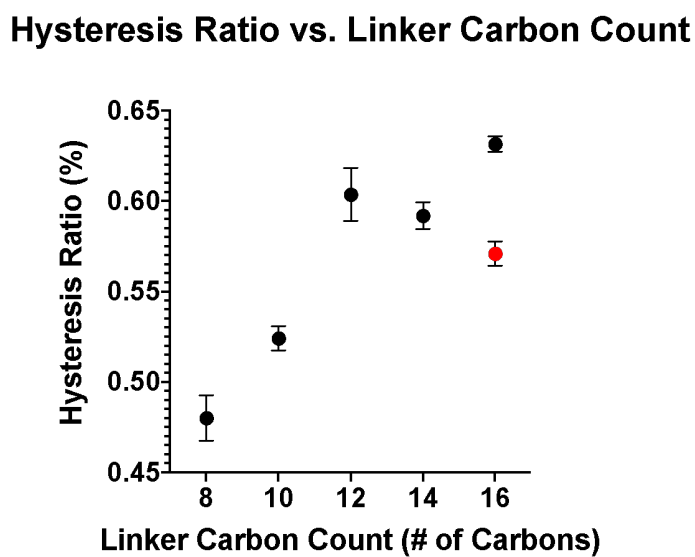


Figure S38. Relationship between compressive hysteresis ratio and linker carbon count in simplified (black) and substituted (red) synthetic linker sporopollenin analogues (error bars = standard deviation)

Hysteresis Ratio vs. Linker Density

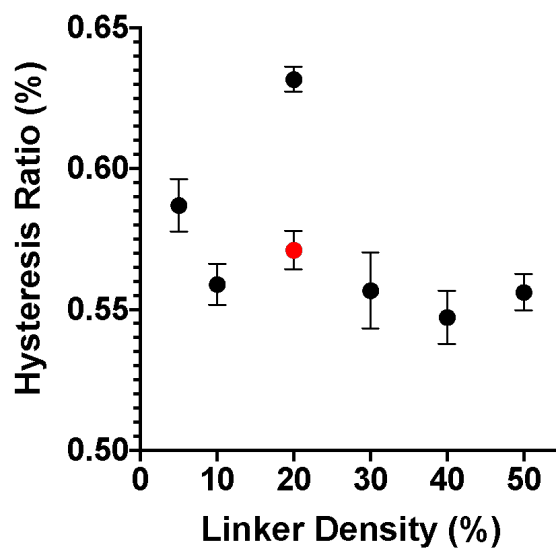


Figure S39. Relationship between compressive hysteresis ratio and linker density in simplified (black) and substituted (red) synthetic linker sporopollenin analogues (error bars = standard deviation)

Hysteresis Ratio vs. PVA Avg. MW

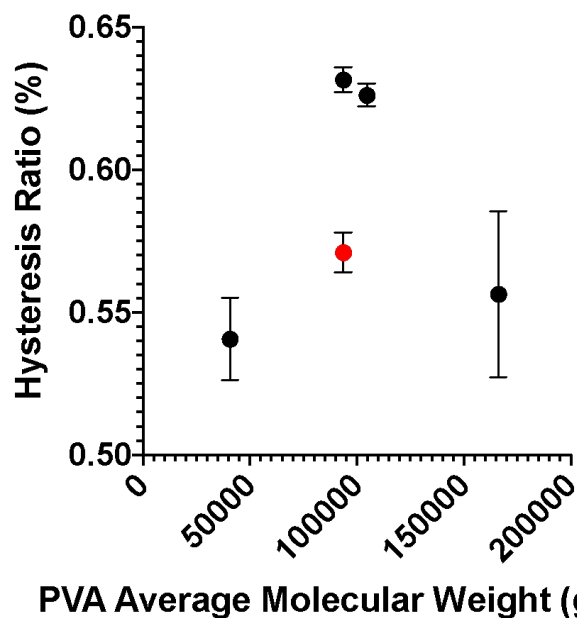


Figure S40. Relationship between compressive hysteresis ratio and PVA MW_{avg} in simplified (black) and substituted (red) synthetic linker sporopollenin analogues (error bars = standard deviation)

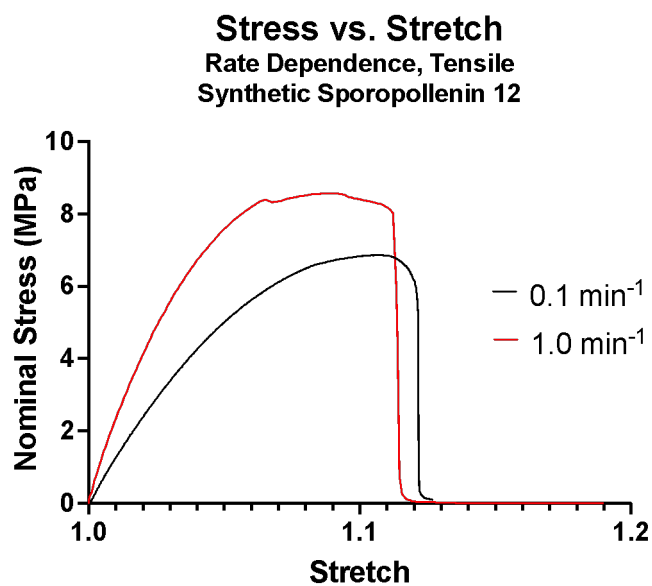


Figure S41. Tensile stress-strain curve for synthetic sporopollenin analogue **12** at 1.0 (red) and 0.1 min⁻¹ (black) loading rates

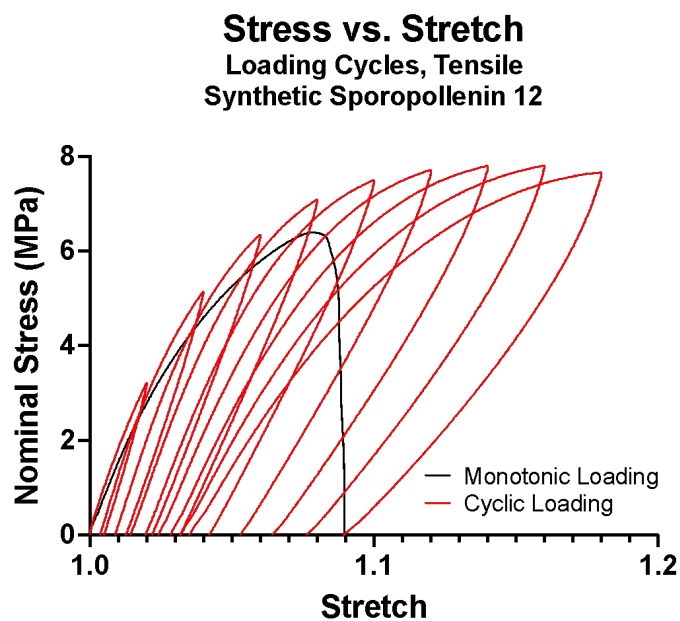


Figure S42. Tensile stress-strain curve for synthetic sporopollenin analogue **12** with cyclic (red) and monotonic (black) loading

Stress vs. Time **Relaxation, Tensile** **Synthetic Sporopollenin 12**

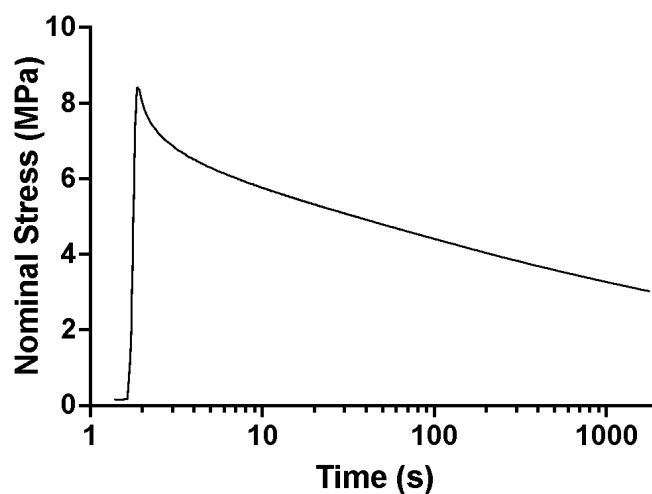


Figure S43. Tensile stress-time relaxation curve for synthetic sporopollenin analogue **12**

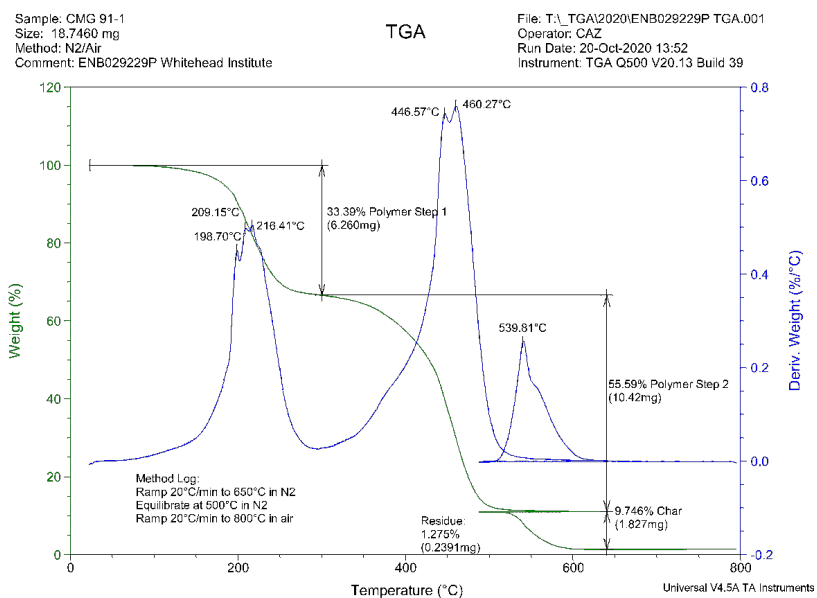


Figure S44. The TGA thermogram for synthetic sporopollenin analogue **6**

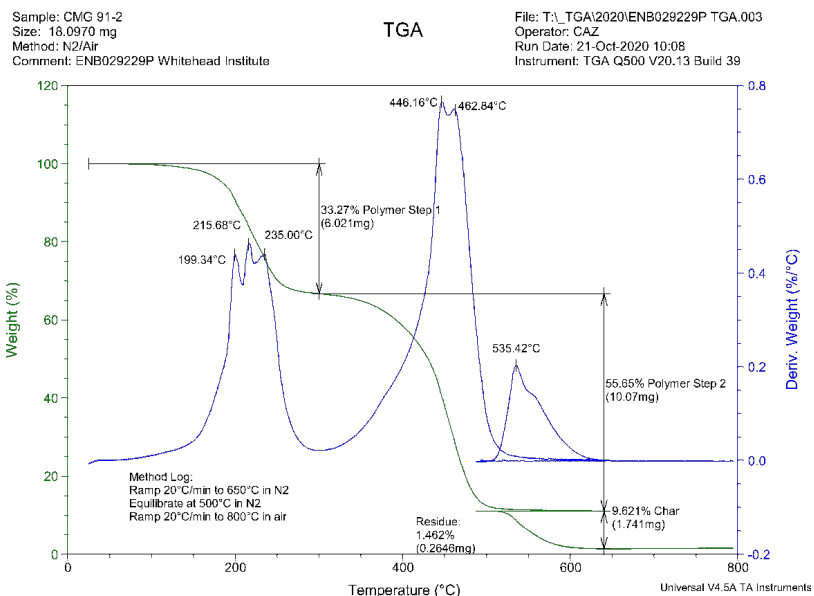


Figure S45. The TGA thermogram for synthetic sporopollenin analogue **7**

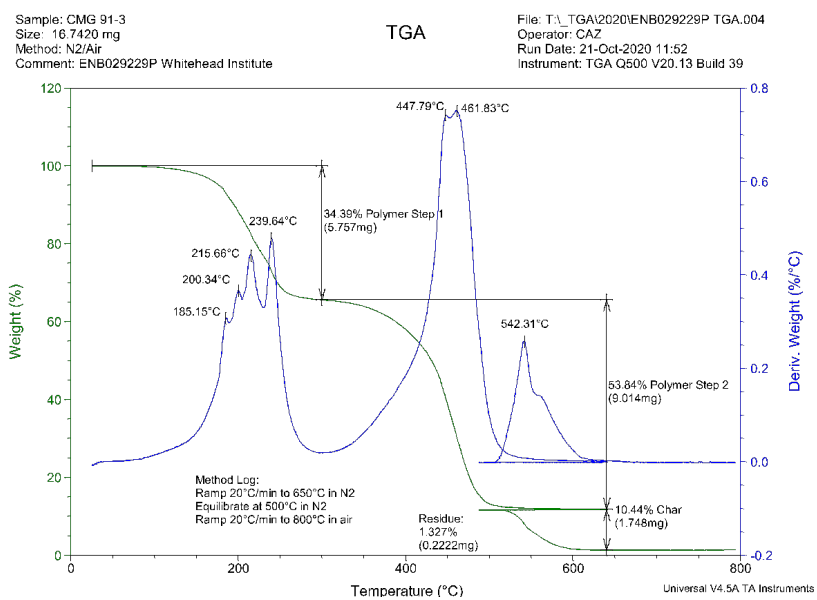


Figure S46. The TGA thermogram for synthetic sporopollenin analogue **8**

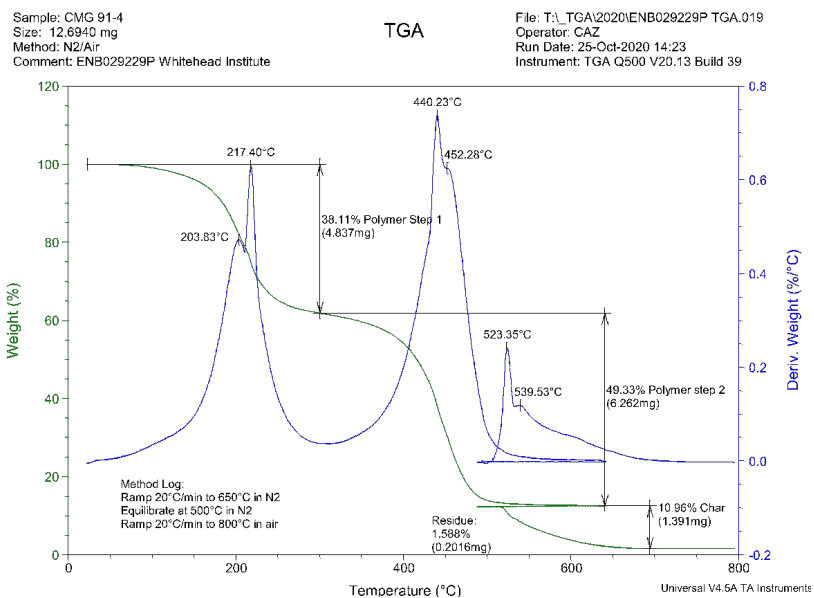


Figure S47. The TGA thermogram for synthetic sporopollenin analogue **9**

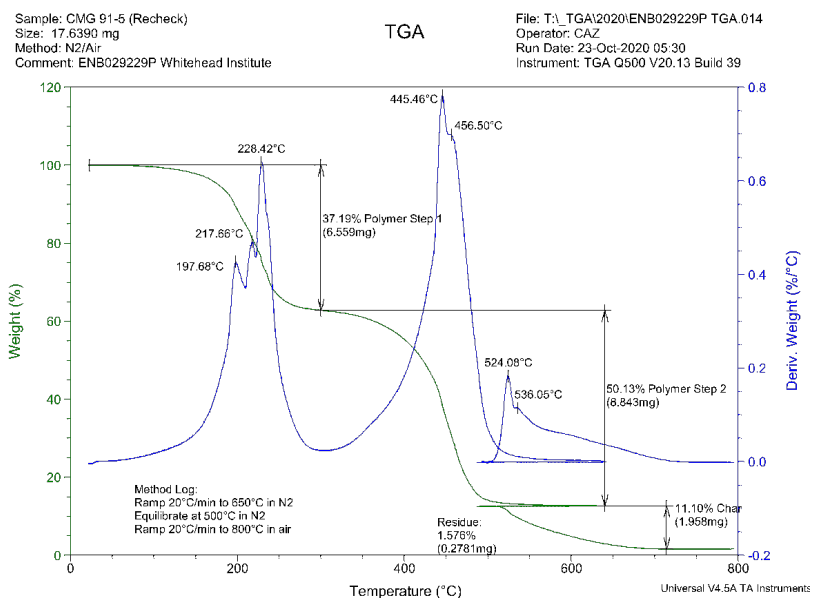


Figure S48. The TGA thermogram for synthetic sporopollenin analogue **10**

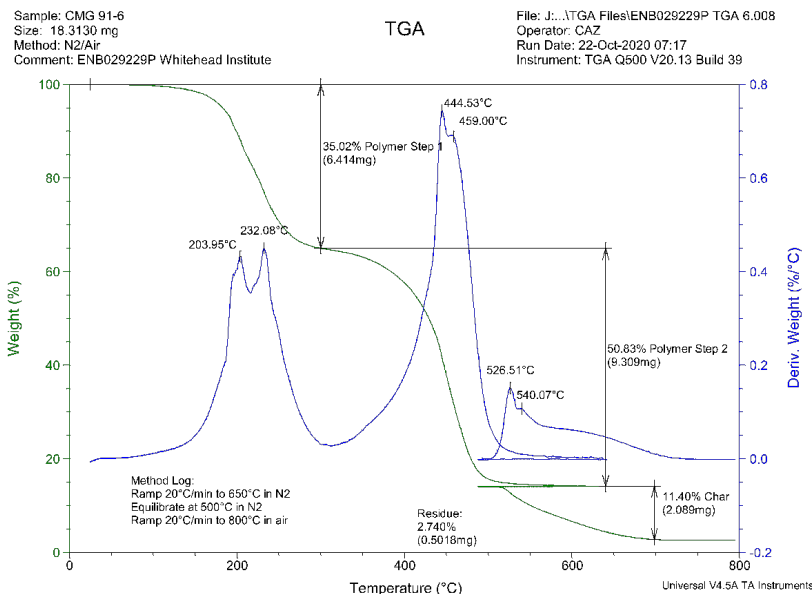


Figure S49. The TGA thermogram for synthetic sporopollenin analogue **11**

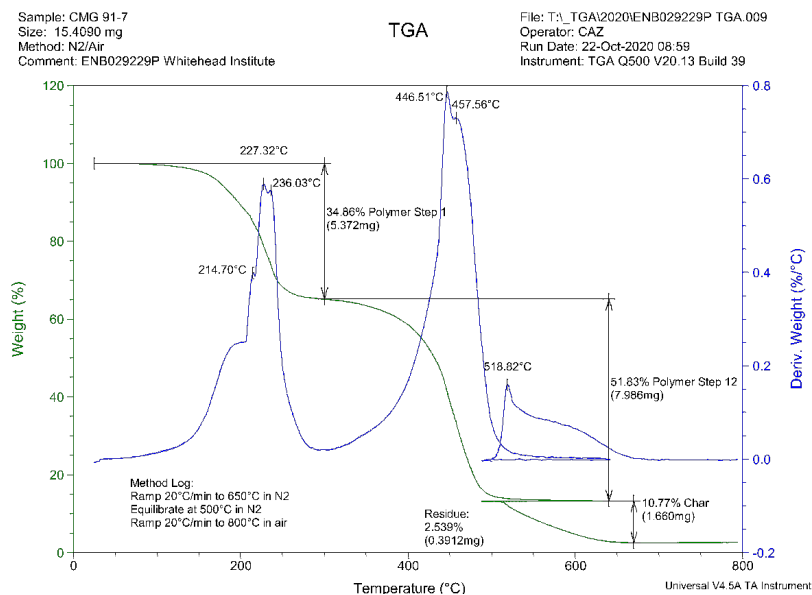


Figure S50. The TGA thermogram for synthetic sporopollenin analogue **12**

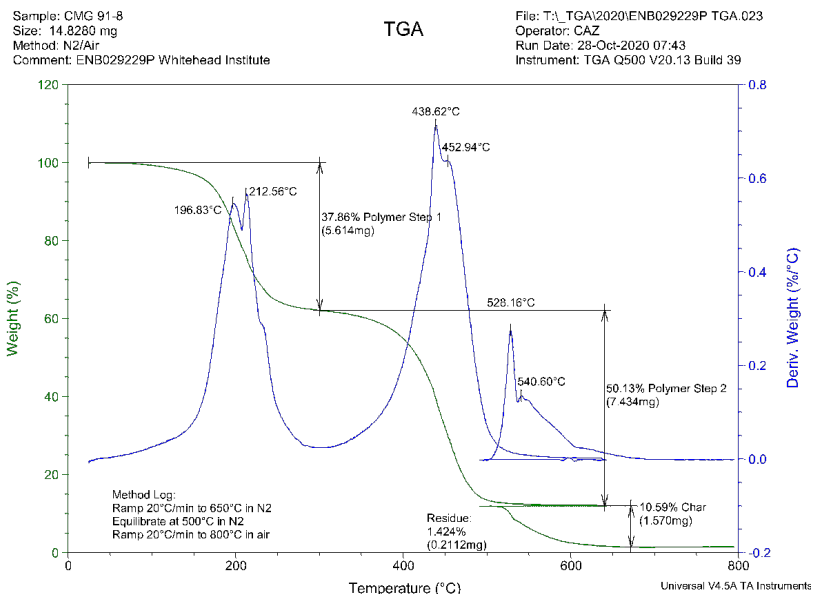


Figure S51. The TGA thermogram for synthetic sporopollenin analogue **13**

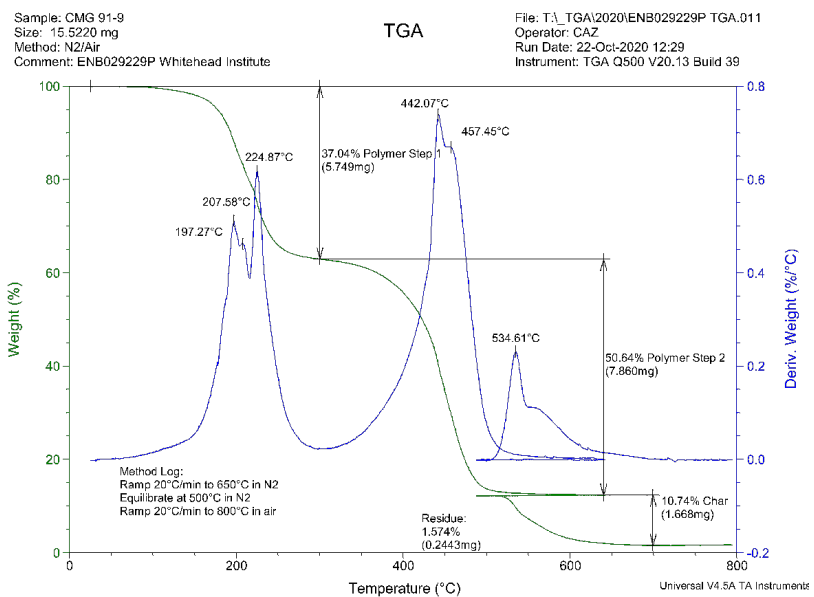


Figure S52. The TGA thermogram for synthetic sporopollenin analogue **14**

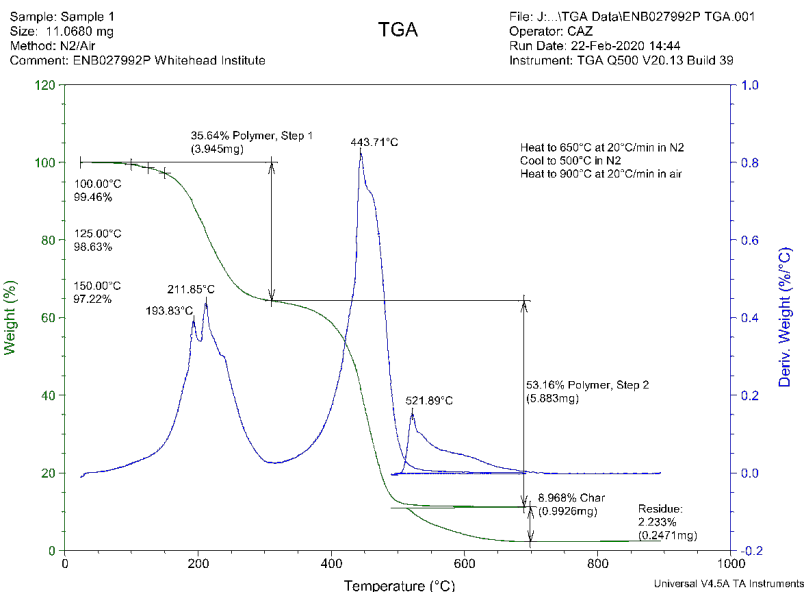


Figure S53. The TGA thermogram for synthetic sporopollenin analogue **15**

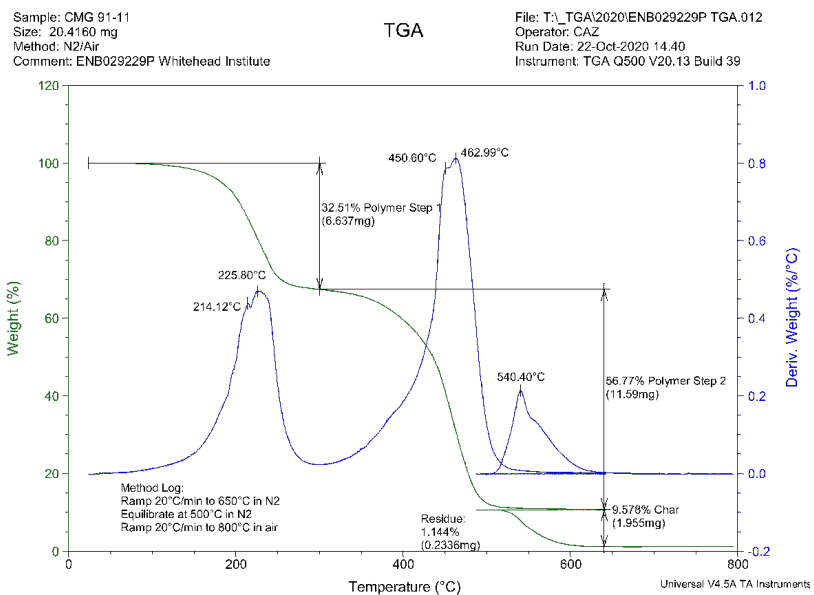


Figure S54. The TGA thermogram for synthetic sporopollenin analogue **16**

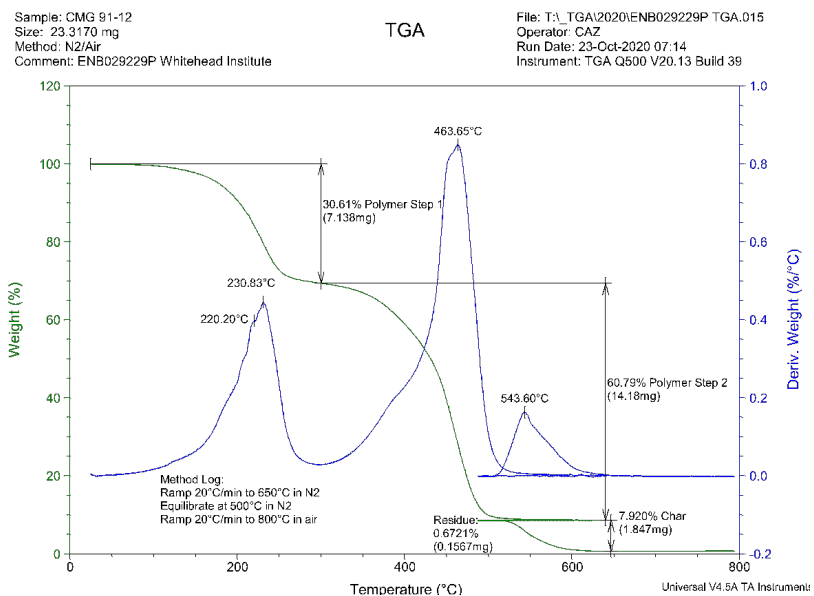


Figure S55. The TGA thermogram for synthetic sporopollenin analogue **17**

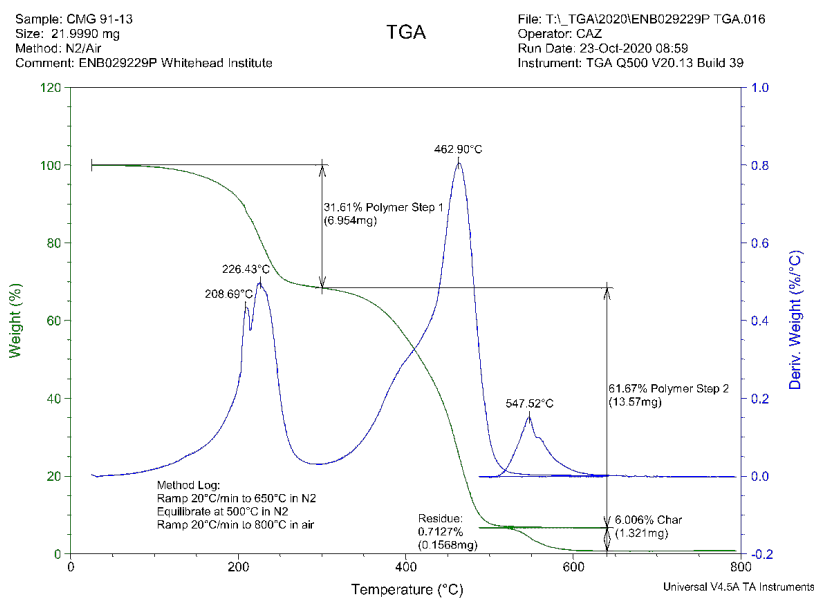


Figure S56. The TGA thermogram for synthetic sporopollenin analogue **18**

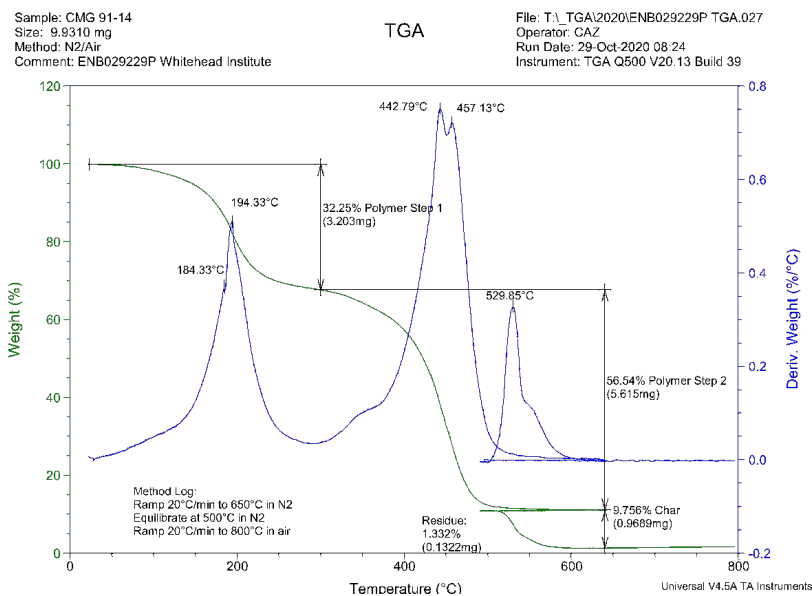


Figure S57. The TGA thermogram for synthetic sporopollenin analogue **26**

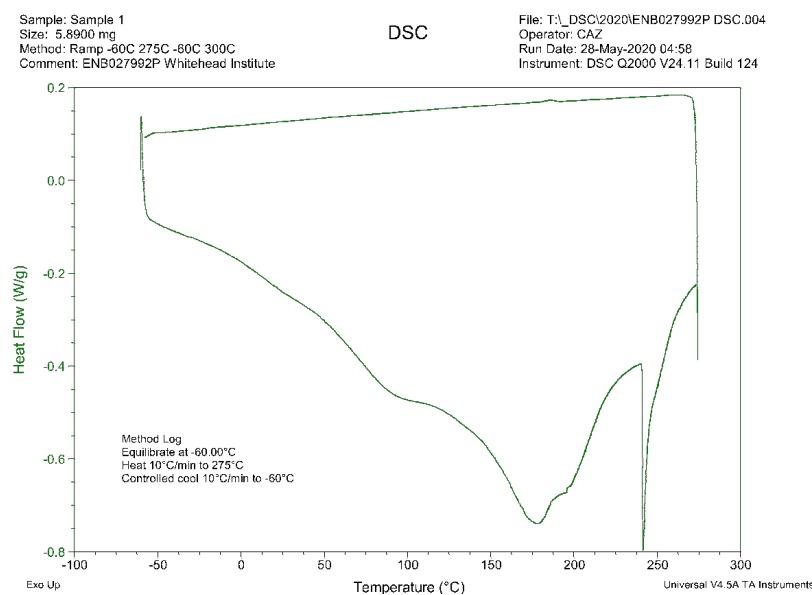


Figure S58. The DSC thermogram for synthetic sporopollenin analogue **15**, first replicate

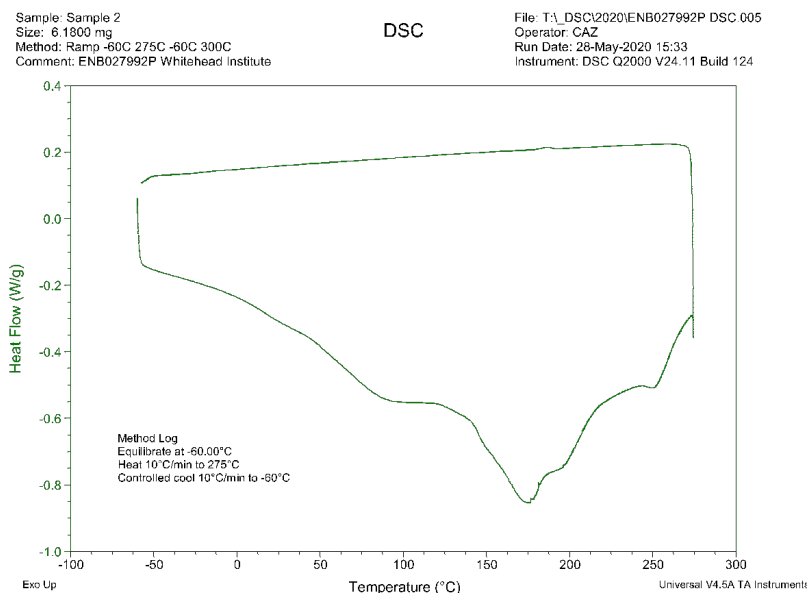


Figure S59. The DSC thermogram for synthetic sporopollenin analogue **15**, second replicate

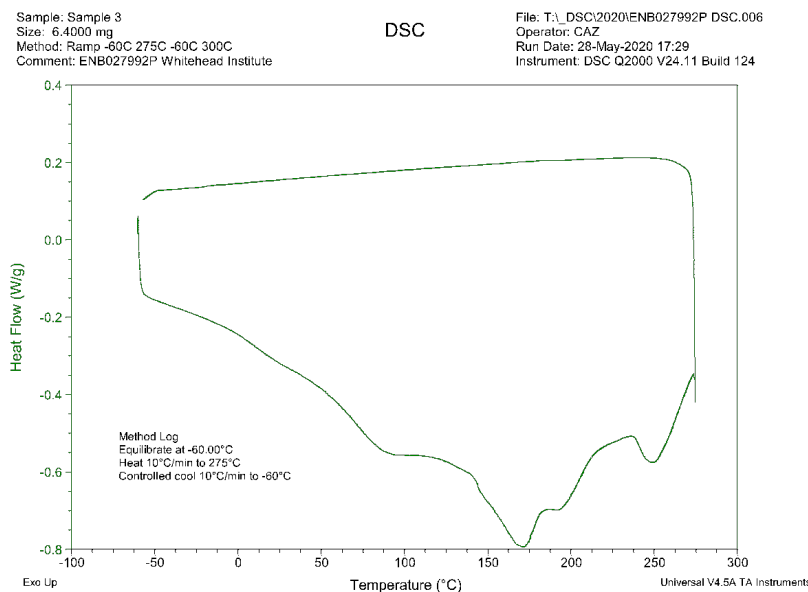


Figure S60. The DSC thermogram for synthetic sporopollenin analogue **15**, third replicate

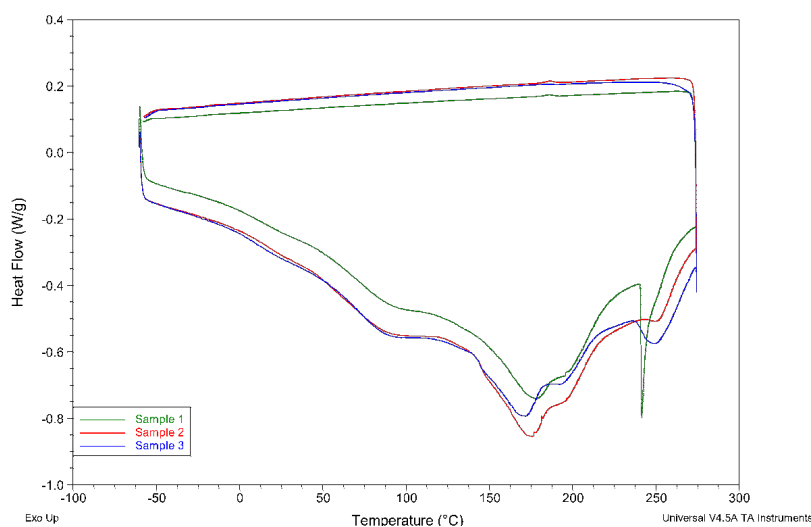


Figure S61. The DSC thermogram for synthetic sporopollenin analogue **15**, all replicates

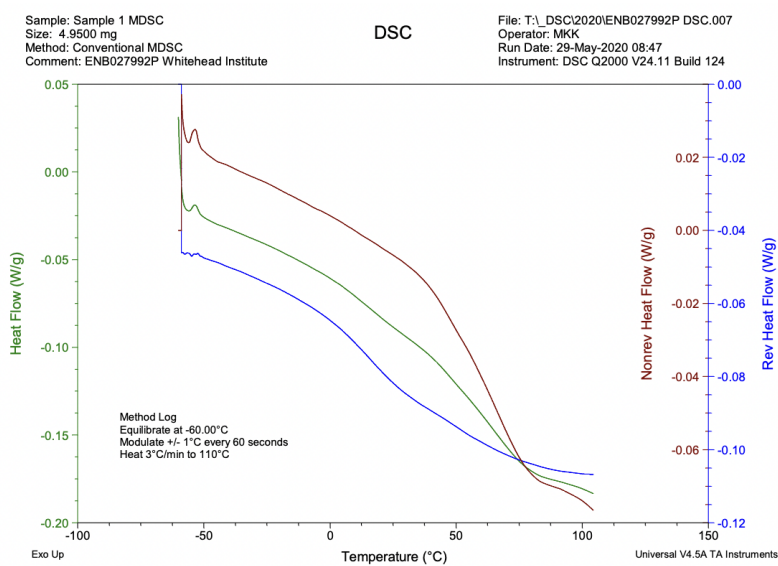


Figure S62. The modulated DSC thermogram for synthetic sporopollenin analogue **15**