
Protein-responsive gut hormone tachykinin directs food choice and impacts lifespan

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Supplementary Table S1: Fly stocks			
Short name	Full genotype	Source	Reference
<i>Akh-GAL4</i>	<i>Akh-GAL4</i> on 3	BDSC #25684	Lee and Park ¹
<i>Akh^{null}</i>	<i>Akh^{SK1}</i> , a CRISPR-induced frame-shift	Gift of Shu Kondo (Tokyo University of Science)	Hentze <i>et al.</i> ²
<i>Akh^{null} Tk^{Gut}-GAL4</i>	<i>R57C10-GAL80- WPRE[su(Hw)attP8]; +; Akh^{null}, Tk::2A::GAL4</i>	Prepared for this work	<i>Tk</i> > (Deng <i>et al.</i> ³), BDSC #84693. <i>R57C10-GAL80</i> , Harris <i>et al.</i> , 2015 ⁴ , gift of Ryusuke Niwa (University of Tsukuba). <i>Akh^{null}</i> (Hentze <i>et al.</i> ²), gift of Shu Kondo (Tokyo University of Science).
<i>CaLexA</i> system	<i>LexAop-CD8::GFP::2A::CD8::GFP; UAS-mLexA::VP16::NFAT, LexAop-CD2::GFP/TM6B, Tb</i>	BDSC #66542	Masuyama <i>et al.</i> ⁵
<i>CaLexA-Luciferase</i>	<i>LexAop-Luciferase; UAS-mLexA::VP16::NFAT/TM6B, Tb</i>	Gift of Michael Rosbash (Brandeis)	Guo, Chen, and Rosbash ⁶
<i>CaLexA-Luciferase, Tk::2A::GAL4</i>	<i>LexAop-Luciferase; Tk::2A::GAL4, UAS-mLexA::VP16::NFAT/TM6B, Tb</i>	Prepared for this work	<i>Tk</i> > (Deng <i>et al.</i> ³), BDSC #84693. <i>LexAop-Luciferase, UAS-mLexA::VP16::NFAT</i> ⁶ , gift of Michael Rosbash (Brandeis).
<i>Cg^{TS}-GAL4</i>	<i>Cg-GAL4 Tub-GAL80^{TS}</i> on 2	Prepared for this work	<i>Tub-GAL80^{TS}</i> ⁷ , BDSC #7108. <i>Cg-GAL4</i> , BDSC #7011.
<i>ChAT-GAL4</i>	<i>ChAT-GAL4^{19B} UAS-GFP^{S65T}</i> on 2	BDSC #6793	Salvaterra and Kitamoto ⁸
<i>dimm-GAL4</i>	<i>P{w[+mW.hs]=GawB}crc[c929]</i>	BDSC #25373	
<i>Ilp2-GAL4, UAS-GFP</i>	<i>Ilp2-GAL4.R 2/CyO</i>	BDSC #37516	Rulifson ⁹
<i>R34G02-GAL4</i>	<i>R34G02-GAL4[attP2]</i>	BDSC #49797	Jenett <i>et al.</i> ¹⁰ ; Jourjine <i>et al.</i> ¹¹
<i>R57C10-GAL4</i>	<i>R57G02-GAL4[attP2]</i>	BDSC #39171	Jenett <i>et al.</i> ¹⁰
<i>R57C10-GAL80</i>	<i>R57C10-GAL80-WPRE[su(Hw)attP8]</i> on X	Gift of R. Niwa, University of Tsukuba	Harris <i>et al.</i> ⁴
<i>R57C10^{TS}-GAL4</i>	<i>Tub-GAL80^{TS}; R57C10-GAL4[attP2]</i>	BDSC #39171 + #7108	
<i>Tdc2-GAL4</i>	<i>Tdc2-GAL4</i> on 2	BDSC #9313	
<i>TH-GAL4</i>	<i>pale-GAL4/CyO</i>	BDSC #95269	Friggi-Grelin <i>et al.</i> ¹²
<i>tim-GAL4</i>	<i>timeless-GAL4</i> on 2	BDSC #7126	Emery <i>et al.</i> ¹³
<i>Tk::2A::GAL4</i>	<i>TI{2A-GAL4}Tk{2A-GAL4}</i> on 3	BDSC #84693	Deng <i>et al.</i> ³
<i>Tk^{Gut}-GAL4 (Tk^{Gut}>)</i>	<i>R57C10-GAL80-WPRE[su(Hw)attP8]; +; Tk::2A::GAL4</i>	Prepared for this work	<i>Tk</i> > (Deng <i>et al.</i> ³), BDSC #84693. <i>R57C10-GAL80</i> (Harris <i>et al.</i> ⁴), gift of Ryusuke Niwa (Tsukuba).
<i>Tkr99D::2A::GAL4</i>	<i>Mi{Trojan-GAL4.2}Tkr99D[MI10336-TG4.2]</i>	BDSC #76208	Lee <i>et al.</i> ¹⁴
<i>UAS-AKH-RNAi</i>	<i>UAS-Akh-RNAi^{v105063}</i> on 2	VDRC #105063	Dietzl <i>et al.</i> ¹⁵
<i>UAS-AkhR-RNAi^{KK}</i>	<i>UAS-AkhR-RNAi^{v109300}</i> on 2	VDRC #109300	Dietzl <i>et al.</i> ¹⁵
<i>UAS-AkhR-RNAi^{TRIP}</i>	<i>UAS-AkhR-RNAi^{TRIP.JF03256}[attP2]</i>	BDSC #29577	Ni <i>et al.</i> ¹⁶ ; Perkins <i>et al.</i> ¹⁷
<i>UAS-CG16700-RNAi</i>	<i>UAS-CG16700-RNAi^{v45188}</i> on 3	VDRC #45188	Dietzl <i>et al.</i> ¹⁵
<i>UAS-CG2930-RNAi</i>	<i>UAS-CG2930-RNAi^{v7031}</i> on 3	VDRC #7031	Dietzl <i>et al.</i> ¹⁵
<i>UAS-CG31248-RNAi</i>	<i>UAS-CG31248-RNAi^{TRIP.JF03189}[attP2]</i>	BDSC #28761	Ni <i>et al.</i> ¹⁶ ; Perkins <i>et al.</i> ¹⁷
<i>UAS-Eaat1-RNAi</i>	<i>UAS-Eaat1-RNAi^{TRIP.HMS02659}[attP40]/CyO</i>	BDSC #43287	Ni <i>et al.</i> ¹⁶ ; Perkins <i>et al.</i> ¹⁷
<i>UAS-gb-RNAi</i>	<i>UAS-gb-RNAi^{v1262}</i> on 2	VDRC #1262	Dietzl <i>et al.</i> ¹⁵
<i>UAS-GCaMP6s</i>	<i>20xUAS-IVS-GCaMP6s[VK00005]</i> on 3	BDSC #42749	Chen <i>et al.</i> ¹⁸
<i>UAS-Gcn2-RNAi</i>	<i>UAS-Gcn2-RNAi^{TRIP.GL00267}[attP2]</i>	BDSC #35355	Ni <i>et al.</i> ¹⁶ ; Perkins <i>et al.</i> ¹⁷
<i>UAS-mCD8::GFP</i>	<i>UAS-mCD8::GFP.L[LL5], UAS-mCD8::GFP.L[2]</i> on 2	BDSC #5137	Lee and Luo, 1999 ¹⁹
<i>UAS-mnd-RNAi</i>	<i>UAS-mnd-RNAi^{v42485}</i> on 3	VDRC #42485	Dietzl <i>et al.</i> ¹⁵
<i>UAS-Rheb</i>	<i>UAS-Rheb^{EP50.084,w-/TM6B,Tb}</i>	Gift of Ernst Hafen and Hugo Stocker (ETH Zürich).	Stocker <i>et al.</i> ²⁰
<i>UAS-sbm-RNAi</i>	<i>UAS-sbm-RNAi^{TRIP.HMC03347}[attP2]</i>	BDSC #51791	Ni <i>et al.</i> ¹⁶ ; Perkins <i>et al.</i> ¹⁷
<i>UAS-slijf-RNAi^{GD}</i>	<i>UAS-slijf-RNAi^{v45590}</i> on 3	VDRC #45590	Dietzl <i>et al.</i> ¹⁵
<i>UAS-slijf-RNAi^{KK}</i>	<i>UAS-slijf-RNAi^{v110425}/CyO</i>	VDRC #110425	Dietzl <i>et al.</i> ¹⁵
<i>UAS-tdTomato</i>	<i>10xUAS-IVS-myr::tdTomato[su(Hw)attP8]</i> on X	BDSC #32223	Pfeiffer <i>et al.</i> ^{21,22}
<i>UAS-tdTomato; CaLexA</i>	<i>10xUAS-IVS-myr::tdTomato[su(Hw)attP8]; LexAop-CD8::GFP::2A::CD8::GFP; UAS-mLexA-VP16-NFAT, lexAop-rCD2::GFP/TM6B, Tb</i>	BDSC #32223 combined with BDSC #66542	Malita <i>et al.</i> ²³ ; <i>tdTomato</i> (Pfeiffer <i>et al.</i> ^{21,22}), BDSC #32223. <i>CaLexA</i> system (Masuyama <i>et al.</i> ⁵), BDSC #66542.

UAS-tdTomato; CaLexA, Tk::2A::GAL4	10xUAS-IVS-myr::tdTomato[su(Hw)attP8]; LexAop-CD8::GFP::2A::CD8::GFP; Tk::2A::GAL4, UAS-mLexA-VP16- NFAT/TM6B, Tb	Prepared for this work	tdTomato (Pfeiffer <i>et al.</i> ^{21,22}) BDSC #32223. CaLexA system (Masuyama <i>et al.</i> ⁵), BDSC #66542. Tk ^{>3} , BDSC #84693.
UAS-TkR99D-RNAi ^{TRIP1}	UAS-TkR99D-RNAi ^{TRIP.JF02663} [attP2]	BDSC #27513	Ni <i>et al.</i> ¹⁶ ; Perkins <i>et al.</i> ¹⁷
UAS-TkR99D-RNAi ^{TRIP2}	UAS-TkR99D-RNAi ^{TRIP.HMC03749} [attP40]	BDSC #55732	Ni <i>et al.</i> ¹⁶ ; Perkins <i>et al.</i> ¹⁷
UAS-Tk-RNAi ^{KK}	UAS-Tk-RNAi ^{v103662} on 2	VDRC #103662	Dietzl <i>et al.</i> ¹⁵
UAS-Tk-RNAi ^{sh}	UAS-Tk-RNAi ^{v330743} [attP40]	VDRC #330743	Dietzl <i>et al.</i> ¹⁵
UAS-Tk-RNAi ^{sh} ; Akh ^{null}	UAS-Tk-RNAi ^{v330743} [attP40]; Akh ^{null}	Prepared for this work	Tk-RNAi ^{sh} , VDRC #330743. Akh ^{null} (Hentze ²), gift of Shu Kondo.
UAS-Tk-RNAi ^{sh} ; UAS- TrpA1	UAS-Tk-RNAi ^{v330743} [attP40]; UAS- TrpA1/TM6B, Tb	Prepared for this work	Tk-RNAi ^{sh} , VDRC #330743. UAS-TrpA1 on 3, gift of Christian Wegener (Würzburg).
UAS-TOR-RNAi	UAS-TOR-RNAi ^{TRIP.HMS01114} [attP2]	BDSC #34639	Ni <i>et al.</i> ¹⁶ ; Perkins <i>et al.</i> ¹⁷
UAS-TrpA1 (II)	UAS-TrpA1	BDSC #26263	Hamada <i>et al.</i> ²⁴
UAS-TrpA1 (III)	UAS-TrpA1 [attP2]/TM6B, Tb	Gift of Christian Wegener (Würzburg)	
UAS-TrpA1; UAS-TOR- RNAi	UAS-TrpA1(B).K [attP16]; UAS-TOR- RNAi ^{TRIP.HMS01114} [attP2]	Prepared for this work	TrpA1 on 2 (Hamada <i>et al.</i> ²⁴), BDSC #26263. TOR-RNAi, BDSC #34639.
UAS-TrpA1; UAS- Tsc1/2	UAS-TrpA1(B).K [attP16]; UAS-Tsc1, UAS- Tsc2	Prepared for this work	TrpA1 on 2 (Hamada <i>et al.</i> ²⁴), BDSC #26263. UAS-Tsc1, UAS-Tsc2 (Tapon <i>et al.</i> ²⁵), gift of Ernst Hafen and Hugo Stocker, also available as BDSC #80576.
UAS-TrpA1-RNAi	UAS-TrpA1-RNAi ^{TRIP.HMS05348} [attP2]	BDSC #66905	Ni <i>et al.</i> ¹⁶ ; Perkins <i>et al.</i> ¹⁷
TrpA1::GFP	TrpA1::GFP on 3	Gift of Yang Xiang	Gong <i>et al.</i> ²⁶
UAS-Tsc1/2	UAS-Tsc1, UAS-Tsc2 on 3	BDSC #80576	UAS-Tsc1, UAS-Tsc2 (Tapon <i>et al.</i> ²⁵) is also available as BDSC #80576.
VGlut1-GAL4	VGlut1-GAL4 ^{OK371} on 2	BDSC #26160	Mahr and Aberle ²⁷
w ¹¹¹⁸	w ¹¹¹⁸ ; +; +	VDRC #60000	
w ¹¹¹⁸	w ¹¹¹⁸ ; +; +	Gift of Christen Mirth (Monash University)	

References

- Lee, G. & Park, J. H. Hemolymph sugar homeostasis and starvation-induced hyperactivity affected by genetic manipulations of the adipokinetic hormone-encoding gene in *Drosophila melanogaster*. *Genetics* **167**, 311-323 (2004).
- Hentze, J. L., Carlsson, M. A., Kondo, S., Nassel, D. R. & Rewitz, K. F. The Neuropeptide Allatostatin A Regulates Metabolism and Feeding Decisions in *Drosophila*. *Sci Rep* **5**, 11680 (2015). <https://doi.org/10.1038/srep11680>
- Deng, B. *et al.* Chemoconnectomics: Mapping Chemical Transmission in *Drosophila*. *Neuron* **101**, 876-893 e874 (2019). <https://doi.org/10.1016/j.neuron.2019.01.045>
- Harris, R. M., Pfeiffer, B. D., Rubin, G. M. & Truman, J. W. Neuron hemilineages provide the functional ground plan for the *Drosophila* ventral nervous system. *Elife* **4** (2015). <https://doi.org/10.7554/eLife.04493>
- Masuyama, K., Zhang, Y., Rao, Y. & Wang, J. W. Mapping neural circuits with activity-dependent nuclear import of a transcription factor. *Journal of neurogenetics* **26**, 89-102 (2012). <https://doi.org/10.3109/01677063.2011.642910>
- Guo, F., Chen, X. & Rosbash, M. Temporal calcium profiling of specific circadian neurons in freely moving flies. *Proc Natl Acad Sci U S A* **114**, E8780-E8787 (2017). <https://doi.org/10.1073/pnas.1706608114>
- McGuire, S. E., Mao, Z. & Davis, R. L. Spatiotemporal gene expression targeting with the TARGET and gene-switch systems in *Drosophila*. *Sci STKE* **2004**, pl6 (2004). <https://doi.org/10.1126/stke.2202004pl6>

- 8 Salvaterra, P. M. & Kitamoto, T. Drosophila cholinergic neurons and processes visualized with Gal4/UAS-GFP. *Brain research. Gene expression patterns* **1**, 73-82 (2001).
- 9 Rulifson, E. J., Kim, S. K. & Nusse, R. Ablation of insulin-producing neurons in flies: growth and diabetic phenotypes. *Science* **296**, 1118-1120 (2002).
<https://doi.org/10.1126/science.1070058>
- 10 Jenett, A. *et al.* A GAL4-driver line resource for Drosophila neurobiology. *Cell Rep* **2**, 991-1001 (2012). <https://doi.org/10.1016/j.celrep.2012.09.011>
- 11 Jourjine, N., Mullaney, B. C., Mann, K. & Scott, K. Coupled Sensing of Hunger and Thirst Signals Balances Sugar and Water Consumption. *Cell* **166**, 855-866 (2016).
<https://doi.org/10.1016/j.cell.2016.06.046>
- 12 Friggi-Grelin, F. *et al.* Targeted gene expression in Drosophila dopaminergic cells using regulatory sequences from tyrosine hydroxylase. *J Neurobiol* **54**, 618-627 (2003). <https://doi.org/10.1002/neu.10185>
- 13 Emery, P., So, W. V., Kaneko, M., Hall, J. C. & Rosbash, M. CRY, a Drosophila clock and light-regulated cryptochrome, is a major contributor to circadian rhythm resetting and photosensitivity. *Cell* **95**, 669-679 (1998).
[https://doi.org/10.1016/s0092-8674\(00\)81637-2](https://doi.org/10.1016/s0092-8674(00)81637-2)
- 14 Lee, P. T. *et al.* A gene-specific T2A-GAL4 library for Drosophila. *eLife* **7** (2018).
<https://doi.org/10.7554/eLife.35574>
- 15 Dietzl, G. *et al.* A genome-wide transgenic RNAi library for conditional gene inactivation in Drosophila. *Nature* **448**, 151-156 (2007). <https://doi.org/nature05954>
- 16 Ni, J. Q. *et al.* A genome-scale shRNA resource for transgenic RNAi in Drosophila. *Nature methods* **8**, 405-407 (2011). <https://doi.org/10.1038/nmeth.1592>
- 17 Perkins, L. A. *et al.* The Transgenic RNAi Project at Harvard Medical School: Resources and Validation. *Genetics* **201**, 843-852 (2015).
<https://doi.org/10.1534/genetics.115.180208>
- 18 Chen, T. W. *et al.* Ultrasensitive fluorescent proteins for imaging neuronal activity. *Nature* **499**, 295-300 (2013). <https://doi.org/10.1038/nature12354>
- 19 Lee, T. & Luo, L. Mosaic analysis with a repressible cell marker for studies of gene function in neuronal morphogenesis. *Neuron* **22**, 451-461 (1999).
[https://doi.org/10.1016/s0896-6273\(00\)80701-1](https://doi.org/10.1016/s0896-6273(00)80701-1)
- 20 Stocker, H. *et al.* Rheb is an essential regulator of S6K in controlling cell growth in Drosophila. *Nat Cell Biol* **5**, 559-565 (2003). <https://doi.org/10.1038/ncb995>
- 21 Pfeiffer, B. D. *et al.* Tools for neuroanatomy and neurogenetics in Drosophila. *Proc Natl Acad Sci U S A* **105**, 9715-9720 (2008).
<https://doi.org/10.1073/pnas.0803697105>
- 22 Pfeiffer, B. D. *et al.* Refinement of tools for targeted gene expression in Drosophila. *Genetics* **186**, 735-755 (2010). <https://doi.org/10.1534/genetics.110.119917>
- 23 Malita, A. *et al.* A gut-derived hormone suppresses sugar appetite and regulates food choice in Drosophila. *Nat Metab* **4**, 1532-1550 (2022).
<https://doi.org/10.1038/s42255-022-00672-z>
- 24 Hamada, F. N. *et al.* An internal thermal sensor controlling temperature preference in Drosophila. *Nature* **454**, 217-220 (2008). <https://doi.org/10.1038/nature07001>
- 25 Tapon, N., Ito, N., Dickson, B. J., Treisman, J. E. & Hariharan, I. K. The Drosophila tuberous sclerosis complex gene homologs restrict cell growth and cell proliferation. *Cell* **105**, 345-355 (2001).

- 26 Gong, J. *et al.* Shear stress activates nociceptors to drive Drosophila mechanical nociception. *Neuron* **110**, 3727-3742 e3728 (2022).
<https://doi.org:10.1016/j.neuron.2022.08.015>
- 27 Mahr, A. & Aberle, H. The expression pattern of the Drosophila vesicular glutamate transporter: a marker protein for motoneurons and glutamatergic centers in the brain. *Gene expression patterns : GEP* **6**, 299-309 (2006).
<https://doi.org:10.1016/j.modgep.2005.07.006>

Supplementary Table S2: Oligo sequences used for qPCR

Oligos for <i>Drosophila</i> genes	Forward	Reverse
<i>Rp49</i>	AGTATCTGATGCCCAACATCG	CAATCTCCTTGCGCTTCTTG
<i>AKH</i>	TCCCAAGAGCGAAGTCCTCA	CCAGAAAGAGCTGTGCCTGA
<i>slif</i>	CTGACGGCTTTGGGAGTGG	AATCAGGAAGGAAATGGTCACC
<i>Tk</i>	CAATTCCTTTGTGGGGATGCG	CTGCTGTTTTCTCTCAAGTCAT
Oligos for mouse genes	Forward	Reverse
<i>RPLP0</i>	AGTATCTGATGCCCAACATCG	CAATCTCCTTGCGCTTCTTG
<i>TAC1</i>	ATTCCTTTGTTGGACTAATGGGC	ACGTCTTCTTTCGTAGTTCTGC

Supplementary Table S3: Reagents used in intestinal organoid culture

Compound	Company	Stock	Usage Dilution
2 mM EDTA in PBS	Sigma Aldrich #03692		
DMEM	Sigma Aldrich #D6546		Foundation of "2-D medium"
Advanced DMEM/F12	Gibco #12634010		Foundation for "basal medium" for 3-D organoids
GlutaMAX	Gibco #31966021	100x	1:100 to make basal medium
HEPES		1M	10 mM in basal medium
Penicillin/Streptomycin (Pen/Strep)		100x	1:100 to make basal medium (0.1 U/mL pen, 0.1 mg/mL strep final)
B27 supplement (minus Vit. A)	Invitrogen #12587-010	50x	1:50 to make ENR
EGF (mouse)	Gibco #PMG8045	50 µg/mL in 0.1% BSA	1:1000 to make ENR (50 ng/mL final)
N2 supplement	Gibco #17502048	100x	1:100 to make ENR
N-Acetyl-L-Cysteine (NAC)	Sigma Aldrich #A96165-5G	500 mM, 500x	1:500 to make ENR (1 mM final)
Nicotinamide	Sigma Aldrich #N0636/N33776	1 M, 100x	1:100 to make ENR (10 mM final)
Noggin (mouse)	PeproTech #250-38	100 µg/mL in 0.1% BSA	1:1000 to make ENR (100 ng/mL final)
ROCK inhibitor Y27632	Tocris #1254	10 mM	1:1000, added just before use (10 µM final)