nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a	Confirmed	
☐	☒	The exact sample size (*n*) for each experimental group/condition, given as a discrete number and unit of measurement
☐	☒	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
☐	☒	The statistical test(s) used AND whether they are one- or two-sided *Only common tests should be described solely by name; describe more complex techniques in the Methods section.*
☐	☒	A description of all covariates tested
☐	☒	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
☐	☒	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
☐	☒	For null hypothesis testing, the test statistic (e.g. *F*, *t*, *r*) with confidence intervals, effect sizes, degrees of freedom and *P* value noted *Give P values as exact values whenever suitable.*
☒	☐	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
☒	☐	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
☒	☐	Estimates of effect sizes (e.g. Cohen's *d*, Pearson's *r*), indicating how they were calculated

Our web collection on statistics for biologists contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection	MALDI mass spectrometry imaging was performed using a Bruker rapifleX MALDI-TOF/TOF and a timsTOF fleX instrument (Bruker Daltonics). LC-MS analysis was conducted on an Agilent 1290 UHPLC system coupled to a 6550 iFunnel Q-TOF mass spectrometer. RT-qPCR was performed using the QuantStudio 3 Real-Time PCR System (Applied Biosystems). Digital PCR was performed on a QIAcuity Digital PCR system (Qiagen). Behavioral testing was recorded via video and quantified using DeepLabCut. Glycoproteomics data were acquired by LC-MS/MS. Immunofluorescence images were acquired on a Ventana Discovery Ultra automated staining platform.
Data analysis	SCiLS Lab (Bruker) for MALDI imaging data processing. MetaboScape 2022 (Bruker) for metabolite annotation. GraphPad Prism for statistical analyses (ANOVA, t-tests, Kaplan-Meier survival analysis, Cox proportional hazards). GlycReSoft for glycoproteomic data analysis. DeepLabCut for automated behavioral quantification. ImageJ/FIJI for image analysis. R for EHR data analysis and statistical modeling.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Proteomics datasets are available at ProteomeXchange under accession number PXD075438. Due to the large size of MALDI imaging datasets, raw data and imzML files are available from the corresponding author upon reasonable request. Source data are provided with this paper.

Research involving human participants, their data, or biological material

Policy information about studies with human participants or human data. See also policy information about sex, gender (identity/presentation), and sexual orientation and race, ethnicity and racism.

Reporting on sex and gender	Both male and female human brain tissue samples were used. Sex was matched between AD and control groups. For the EHR cohort, sex distribution was reported across all diagnostic categories.
Reporting on race, ethnicity, or other socially relevant groupings	Race and ethnicity data were not specifically analyzed or reported in this study.
Population characteristics	De-identified human brain tissue from the UF-ADRC, including cognitively normal controls and AD patients (Braak stages 0-6). PMIs, age, and sex matched between groups. EHR cohort from UF Health IDR (2012-2024), over 2 million patients.
Recruitment	Human brain tissue obtained from UF-ADRC under IRB-approved protocols. EHR patients identified by ICD-9-CM and ICD-10-CM codes for MCI, AD, and ADRD. Glucosamine users identified through NLP pipeline.
Ethics oversight	UF-ADRC IRB-approved protocols for human tissue. UF IRB (IRB202001888, exempt) for EHR study. All samples de-identified.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

☒ Life sciences ☐ Behavioural & social sciences ☐ Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	No statistical methods were used to predetermine sample sizes for animal experiments. Sample sizes were chosen based on prior experience with similar experimental approaches. For EHR analysis, all patients meeting inclusion criteria were included. Sample sizes are detailed in figure legends.
Data exclusions	All animals and data points are included for statistical analysis. For EHR analysis, records with inconsistencies (death dates preceding birth/diagnosis) were excluded. Transition analyses excluded patients without MCI diagnosis preceding AD/ADRD.
Replication	Hyperglycosylation validated across multiple independent cohorts (fresh-frozen and FFPE human AD brain, Braak stages 0-6) and replicated in two mouse models (5xFAD and PS19). MALDI findings orthogonally validated by LC-MS, glycoproteomics, and RT-qPCR/digital PCR. Behavioral effects replicated across cohorts. All replication attempts were successful.
Randomization	Mice randomly assigned to treatment groups (shPGM3 vs. shScrambled, NGI-1 vs. PBS, glucosamine vs. control). For behavioral testing, mice randomly assigned to testing order. LC-MS samples randomized prior to acquisition. EHR patients categorized by diagnosis and glucosamine use.
Blinding	LC-MS samples randomized and analyzed in a blinded manner. Behavioral testing quantified using automated DeepLabCut software. MALDI imaging processed with automated pipelines. Investigators not blinded during surgeries (not feasible). EHR analyses performed computationally.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study
☐	☒ Antibodies
☐	☒ Eukaryotic cell lines
☐	☒ Palaeontology and archaeology
☒	☐ Animals and other organisms
☒	☐ Clinical data
☐	☒ Dual use research of concern
☐	☒ Plants

Methods

n/a	Involved in the study
☐	☒ ChIP-seq
☐	☒ Flow cytometry
☐	☒ MRI-based neuroimaging

Antibodies

Antibodies used: GFAP (1:100, GTX85454, GeneTex); 6E10 (1:100, 803015, Biolegend); Thr231 (1:300, MN1040, Thermo Scientific); STT3A (1:100, 12034-1-AP, Proteintech); ALG13 (1:100, 28738-1-AP, Proteintech); DPAGT1/GPT (1:100, ab214179, Abcam); O-GlcNAc (1:100, MA1-072, Thermo Scientific). Secondary: Alexa Fluor-conjugated (Invitrogen).

Validation: STT3A and ALG13 validated by preadsorption assay with recombinant proteins (Ag27072 and Ag30299, Proteintech; Supplementary Fig. 17). All other antibodies manufacturer-validated and used at established concentrations from published protocols.

Eukaryotic cell lines

Policy information about cell lines and Sex and Gender in Research

Cell line source(s): Primary cortical neurons from C57BL/6J mouse embryos (E17-18). Not established cell lines.

Authentication: Primary neurons identified by morphology. Authentication not applicable for primary cultures.

Mycoplasma contamination: Mycoplasma testing not performed; short-term primary cultures from embryonic tissue under sterile conditions.

Commonly misidentified lines (See ICLAC register): No commonly misidentified cell lines used.

Palaeontology and Archaeology

Specimen provenance: *Provide provenance information for specimens and describe permits that were obtained for the work (including the name of the issuing authority, the date of issue, and any identifying information). Permits should encompass collection and, where applicable, export.*

Specimen deposition: *Indicate where the specimens have been deposited to permit free access by other researchers.*

Dating methods: *If new dates are provided, describe how they were obtained (e.g. collection, storage, sample pretreatment and measurement), where they were obtained (i.e. lab name), the calibration program and the protocol for quality assurance OR state that no new dates are provided.*

☐ Tick this box to confirm that the raw and calibrated dates are available in the paper or in Supplementary Information.

Ethics oversight: *Identify the organization(s) that approved or provided guidance on the study protocol, OR state that no ethical approval or guidance was required and explain why not.*

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Animals and other research organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research, and Sex and Gender in Research

Laboratory animals: 5xFAD (B6SJL-Tg(APPSwFlLon,PSEN1*M146L*L286V)6799Vas/Mmjax), PS19 (B6;C3-Tg(Prnp-MAPT*P301S)PS19Vle/J), and C57BL/6J mice. Both sexes, 8 months old. Housed 14h light/10h dark, 18-23C, 50-60% humidity. Teklad #2018 diet ad libitum.

Wild animals: No wild animals were used.

Reporting on sex	All procedures approved by University of Florida IACUC.
Field-collected samples	No field-collected samples used.
Ethics oversight	University of Florida IACUC.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Clinical data

Policy information about clinical studies

All manuscripts should comply with the ICMJE guidelines for publication of clinical research and a completed CONSORT checklist must be included with all submissions.

Clinical trial registration	Retrospective observational cohort study using EHR; clinical trial registration not applicable.
Study protocol	UF IRB (IRB202001888, exempt). Patients with MCI/AD/ADRD identified from UF Health IDR (2012-2024), stratified by glucosamine use via NLP extraction.
Data collection	Retrospective EHR data from UF Health IDR (over 1 billion facts, 2+ million patients). MCI/AD/ADRD identified by ICD codes. Glucosamine users identified by NLP from clinical notes and medication records.
Outcomes	Kaplan-Meier survival by glucosamine use within each diagnostic category. MCI-to-AD and MCI-to-ADRD transition rates. Cox proportional hazards modeling.

Dual use research of concern

Policy information about dual use research of concern

Hazards

Could the accidental, deliberate or reckless misuse of agents or technologies generated in the work, or the application of information presented in the manuscript, pose a threat to:

No	Yes	
☐	☐	Public health
☐	☐	National security
☐	☐	Crops and/or livestock
☐	☐	Ecosystems
☐	☐	Any other significant area

Experiments of concern

Does the work involve any of these experiments of concern:

No	Yes	
☐	☐	Demonstrate how to render a vaccine ineffective
☐	☐	Confer resistance to therapeutically useful antibiotics or antiviral agents
☐	☐	Enhance the virulence of a pathogen or render a nonpathogen virulent
☐	☐	Increase transmissibility of a pathogen
☐	☐	Alter the host range of a pathogen
☐	☐	Enable evasion of diagnostic/detection modalities
☐	☐	Enable the weaponization of a biological agent or toxin
☐	☐	Any other potentially harmful combination of experiments and agents

Plants

Seed stocks — *Report on the source of all seed stocks or other plant material used. If applicable, state the seed stock centre and catalogue number. If plant specimens were collected from the field, describe the collection location, date and sampling procedures.*

Novel plant genotypes — *Describe the methods by which all novel plant genotypes were produced. This includes those generated by transgenic approaches, gene editing, chemical/radiation-based mutagenesis and hybridization. For transgenic lines, describe the transformation method, the number of independent lines analyzed and the generation upon which experiments were performed. For gene-edited lines, describe the editor used, the endogenous sequence targeted for editing, the targeting guide RNA sequence (if applicable) and how the editor was applied.*

Authentication — *Describe any authentication procedures for each seed stock used or novel genotype generated. Describe any experiments used to assess the effect of a mutation and, where applicable, how potential secondary effects (e.g. second site T-DNA insertions, mosiacism, off-target gene editing) were examined.*

ChIP-seq

Data deposition

☐ Confirm that both raw and final processed data have been deposited in a public database such as GEO.

☐ Confirm that you have deposited or provided access to graph files (e.g. BED files) for the called peaks.

Data access links
May remain private before publication. — *For "Initial submission" or "Revised version" documents, provide reviewer access links. For your "Final submission" document, provide a link to the deposited data.*

Files in database submission — *Provide a list of all files available in the database submission.*

Genome browser session
(e.g. UCSC) — *Provide a link to an anonymized genome browser session for "Initial submission" and "Revised version" documents only, to enable peer review. Write "no longer applicable" for "Final submission" documents.*

Methodology

Replicates — *Describe the experimental replicates, specifying number, type and replicate agreement.*

Sequencing depth — *Describe the sequencing depth for each experiment, providing the total number of reads, uniquely mapped reads, length of reads and whether they were paired- or single-end.*

Antibodies — *Describe the antibodies used for the ChIP-seq experiments; as applicable, provide supplier name, catalog number, clone name, and lot number.*

Peak calling parameters — *Specify the command line program and parameters used for read mapping and peak calling, including the ChIP, control and index files used.*

Data quality — *Describe the methods used to ensure data quality in full detail, including how many peaks are at FDR 5% and above 5-fold enrichment.*

Software — *Describe the software used to collect and analyze the ChIP-seq data. For custom code that has been deposited into a community repository, provide accession details.*

Flow Cytometry

Plots

Confirm that:

☐ The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).

☐ The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).

☐ All plots are contour plots with outliers or pseudocolor plots.

☐ A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation — *Describe the sample preparation, detailing the biological source of the cells and any tissue processing steps used.*

Instrument — *Identify the instrument used for data collection, specifying make and model number.*

Software — *Describe the software used to collect and analyze the flow cytometry data. For custom code that has been deposited into a community repository, provide accession details.*

Cell population abundance

Describe the abundance of the relevant cell populations within post-sort fractions, providing details on the purity of the samples and how it was determined.

Gating strategy

Describe the gating strategy used for all relevant experiments, specifying the preliminary FSC/SSC gates of the starting cell population, indicating where boundaries between "positive" and "negative" staining cell populations are defined.

☐ Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.

Magnetic resonance imaging

Experimental design

Design type

Indicate task or resting state; event-related or block design.

Design specifications

Specify the number of blocks, trials or experimental units per session and/or subject, and specify the length of each trial or block (if trials are blocked) and interval between trials.

Behavioral performance measures

State number and/or type of variables recorded (e.g. correct button press, response time) and what statistics were used to establish that the subjects were performing the task as expected (e.g. mean, range, and/or standard deviation across subjects).

Acquisition

Imaging type(s)

Specify: functional, structural, diffusion, perfusion.

Field strength

Specify in Tesla

Sequence & imaging parameters

Specify the pulse sequence type (gradient echo, spin echo, etc.), imaging type (EPI, spiral, etc.), field of view, matrix size, slice thickness, orientation and TE/TR/flip angle.

Area of acquisition

State whether a whole brain scan was used OR define the area of acquisition, describing how the region was determined.

Diffusion MRI ☐ Used ☐ Not used

Preprocessing

Preprocessing software

Provide detail on software version and revision number and on specific parameters (model/functions, brain extraction, segmentation, smoothing kernel size, etc.).

Normalization

If data were normalized/standardized, describe the approach(es): specify linear or non-linear and define image types used for transformation OR indicate that data were not normalized and explain rationale for lack of normalization.

Normalization template

Describe the template used for normalization/transformation, specifying subject space or group standardized space (e.g. original Talairach, MNI305, ICBM152) OR indicate that the data were not normalized.

Noise and artifact removal

Describe your procedure(s) for artifact and structured noise removal, specifying motion parameters, tissue signals and physiological signals (heart rate, respiration).

Volume censoring

Define your software and/or method and criteria for volume censoring, and state the extent of such censoring.

Statistical modeling & inference

Model type and settings

Specify type (mass univariate, multivariate, RSA, predictive, etc.) and describe essential details of the model at the first and second levels (e.g. fixed, random or mixed effects; drift or auto-correlation).

Effect(s) tested

Define precise effect in terms of the task or stimulus conditions instead of psychological concepts and indicate whether ANOVA or factorial designs were used.

Specify type of analysis: ☐ Whole brain ☐ ROI-based ☐ Both

Statistic type for inference

Specify voxel-wise or cluster-wise and report all relevant parameters for cluster-wise methods.

(See Eklund et al. 2016)

Correction

Describe the type of correction and how it is obtained for multiple comparisons (e.g. FWE, FDR, permutation or Monte Carlo).

Models & analysis

n/a	Involved in the study
☐	☐ Functional and/or effective connectivity
☐	☐ Graph analysis
☐	☐ Multivariate modeling or predictive analysis

Functional and/or effective connectivity

Report the measures of dependence used and the model details (e.g. Pearson correlation, partial correlation, mutual information).

Graph analysis

Report the dependent variable and connectivity measure, specifying weighted graph or binarized graph, subject- or group-level, and the global and/or node summaries used (e.g. clustering coefficient, efficiency, etc.).

Multivariate modeling and predictive analysis

Specify independent variables, features extraction and dimension reduction, model, training and evaluation metrics.