

◀ **Figure EV1. Related to Fig. 3, MB-specific *Atg5* knockdown does not attenuate H₂O₂ resistance or feeding behavior.**

(A, B) Kaplan–Meier survival curves of male (A) and female (B) flies with MB-specific *Atg5* knockdown (*ok107-Gal4>UAS-Atg5-RNAi*, blue curve) compared to *ok107-Gal4/+* and *UAS-Atg5-RNAi/+* control flies continuously exposed to 2% H₂O₂ from day 2 onwards. *n* = 85–89 flies per genotype and sex. Pairwise log-rank (Mantel–Cox) test comparison. **p* = 0.0180, ***p* = 0.0011, ns not significant. (C, D) Kaplan–Meier survival curves of male (C) and female (D) flies with MB-specific *Atg5* knockdown (*vt30559-Gal4>UAS-Atg5-RNAi*, green curve) compared to *vt30559-Gal4/+* and *UAS-Atg5-RNAi/+* control flies continuously exposed to 2% H₂O₂ from day 2 onwards. *n* = 87–90 flies per genotype and sex. The *UAS-Atg5-RNAi/+* controls in (A, B) are the same as in (C, D). Pairwise log-rank (Mantel–Cox) test comparison. ****p* = 0.0003, *****p* < 0.0001, ns not significant. (E) Number of sips/hour of 5-day-old female *vt30559-Gal4>UAS-Atg5-RNAi* and *ok107-Gal4>UAS-Atg5-RNAi* flies compared to their respective age- and sex-matched controls (*vt30559-Gal4/+* and *ok107-Gal4/+*). Flies were sorted at 3 days of age and transferred 24 h before the experiment onto 2% H₂O₂ containing sucrose solution. During the experiment, consumption of 2% H₂O₂-containing food solution was automatically assessed in FlyPad arenas. *n* = 25–30 flies per genotype pooled from two independent experiments. Mann–Whitney test. ns not significant. (F) Number of sips/hour of 5-day-old female *vt30559-Gal4>UAS-Atg5-RNAi* and *ok107-Gal4>UAS-Atg5-RNAi* flies compared to their respective age- and sex-matched controls (*vt30559-Gal4/+* and *ok107-Gal4/+*). Flies were sorted at 3 days of age and the consumption of food solution containing 1% Agar, 5% yeast and 5% sucrose was automatically assessed in FlyPad arenas. *n* = 29–45 flies per genotype pooled from three independent experiments. Mann–Whitney test. **p* = 0.0270, ns not significant. Data presented as mean ± SD. Source data are available online for this figure.

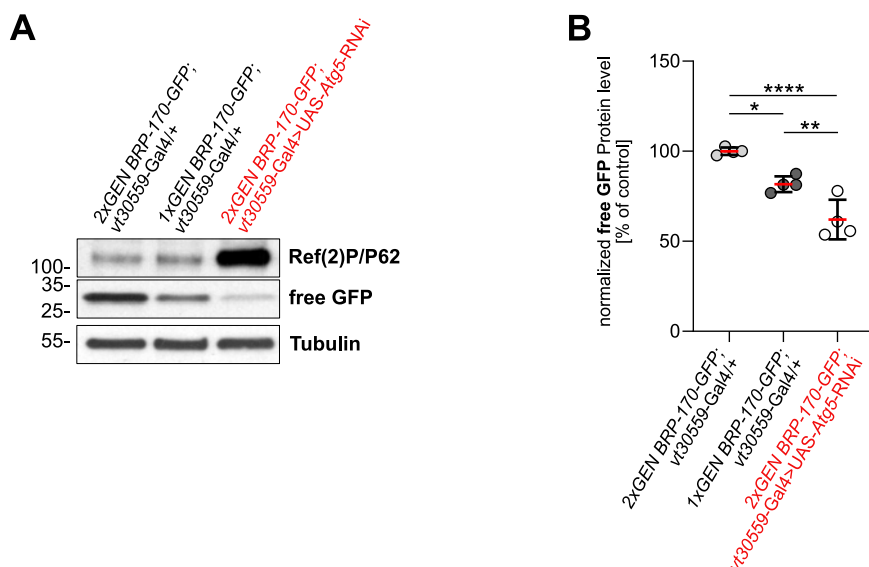


Figure EV2. Related to Fig. 6, lysosomal degradation of a GFP-tagged 170 kDa BRP fusion protein is strongly reduced in fly brains with MB-autophagy impairment.

(A) Representative Western blot of brain homogenates from 5-day-old female flies expressing either one or two additional genomic copies of a GFP-tagged BRP 170 kDa isoform. Control flies additionally carry the vt30559-Gal4/+ transgene alone, while in the effect group, UAS-Atg5-RNAi is driven exclusively in the MB under control of vt30559-Gal4. Membranes were probed against GFP and Ref(2)P/P62, and Tubulin was used as a loading control. An amount equal to two brains was loaded for each lane. (B) Quantification of free GFP protein amounts. Measured band intensities were first normalized to the respective Tubulin band intensity, and values of all groups were normalized to the 2xGen BRP-170-GFP;vt30559-Gal4/+ control group. $n = 4$ independently prepared Western blot samples run on two separate gels. One-way ANOVA with Tukey's post hoc test for multiple comparisons. * $p = 0.0120$, ** $p = 0.0079$, **** $p < 0.0001$. Data presented as mean $\pm$ SD. Source data are available online for this figure.

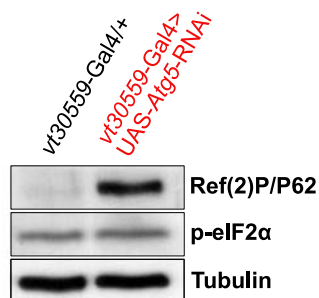
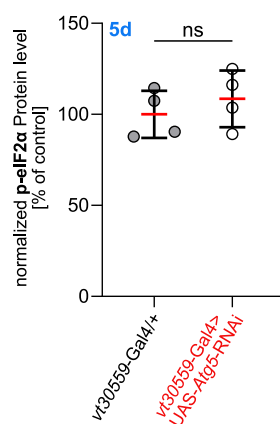
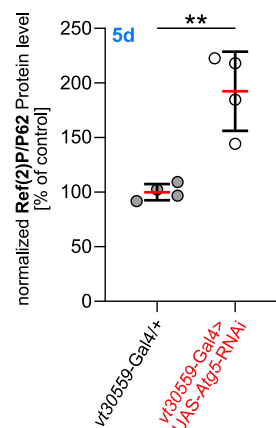
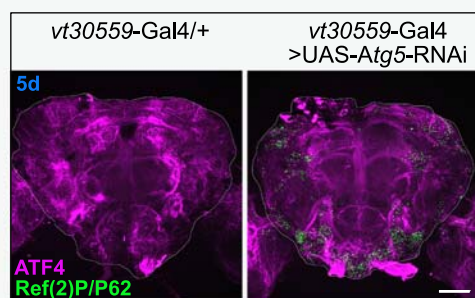
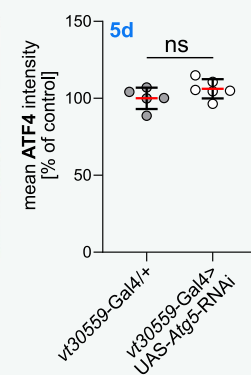
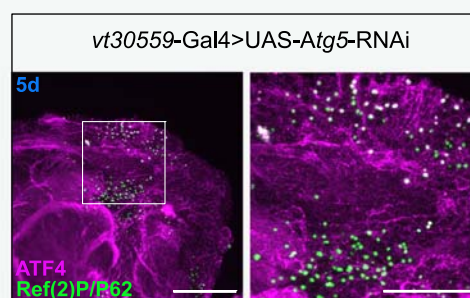
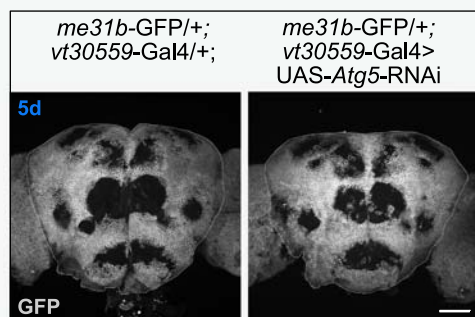
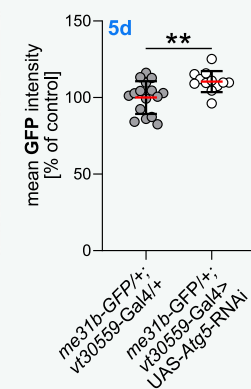
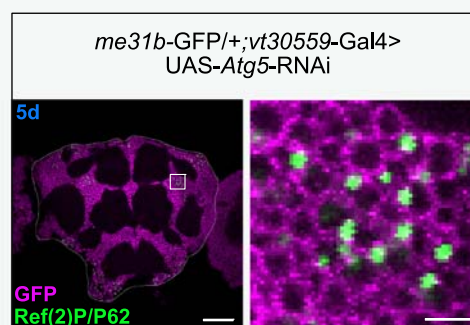
A**B****C****D****E****F****G****H****I**

Figure EV3. Related to Fig. 7, MB-specific *Atg5* knockdown does not induce global activation of the integrated stress response (ISR).

(A–C) Western blot analysis of Ref(2)P/P62, as well as of Ser51-phosphorylation levels of eIF2 α in brain homogenates of 5-day-old female *vt30559-Gal4>UAS-Atg5-RNAi* flies in comparison to age- and sex-matched *vt30559-Gal4/+* controls. (A) Representative Western blot displaying protein bands for Ref(2)P/P62, p-eIF2 α and Tubulin as loading control. An amount equal to 2 brains was loaded for each lane. (B) Quantification of p-eIF2 α levels. Band intensities were first normalized to the respective Tubulin band intensity, and all values were subsequently normalized to the *vt30559-Gal4/+* control group. $n = 4$ independently prepared Western blot samples run together on the same gel; unpaired two-tailed t -test. ns not significant. (C) Quantification of Ref(2)P/P62 levels. Band intensities were first normalized to the respective Tubulin band intensity, and all values were subsequently normalized to the *vt30559-Gal4/+* control group. $n = 4$ independently prepared Western blot samples run together on the same Gel; unpaired two-tailed t -test. $^{**}p = 0.0025$. (D–F) Confocal microscopy analysis of ATF4 and Ref(2)P/P62 in adult brains of 5-day-old female *vt30559-Gal4>UAS-Atg5-RNAi* flies and *vt30559-Gal4/+* controls. (D) Representative confocal max projections of ATF4 and Ref(2)P/P62. (E) Quantification of whole brain mean ATF4 intensities in 5-day-old female flies. Scale bar = 50 μ m; $n = 5$ –6 adult brains per genotype; unpaired two-tailed t -test. ns not significant. (F) Representative confocal max projections of the anterior brain of a *vt30559-Gal4>UAS-Atg5-RNAi* fly, illustrating Ref(2)P/P62 aggregates with and without co-localization of ATF4. Scale bar left panel = 50 μ m; Scale bar right panel = 25 μ m. (G–I) Confocal microscopy analysis of endogenously tagged me31b-GFP in adult brains of 5-day-old female *me31b-GFP/+;vt30559-Gal4>UAS-Atg5-RNAi* flies and *me31b-GFP;vt30559-Gal4/+* controls. (G) Representative confocal max projections of me31b-GFP fluorescent intensity. (H) Quantification of whole brain mean me31b-GFP intensities in 5-day-old female flies. Scale bar = 50 μ m; $n = 12$ –16 adult brains per genotype pooled from two independent experiments; unpaired two-tailed t -test. $^{**}p = 0.0066$. (I) Representative images of a single optical section of the anterior brain of a *vt30559-Gal4>UAS-Atg5-RNAi* fly, illustrating Ref(2)P/P62 aggregates without observable accumulation of me31b-GFP signal. Scale bar left panel = 50 μ m, Scale bar right panel = 5 μ m. Data presented as mean $\pm$ SD. Source data are available online for this figure.

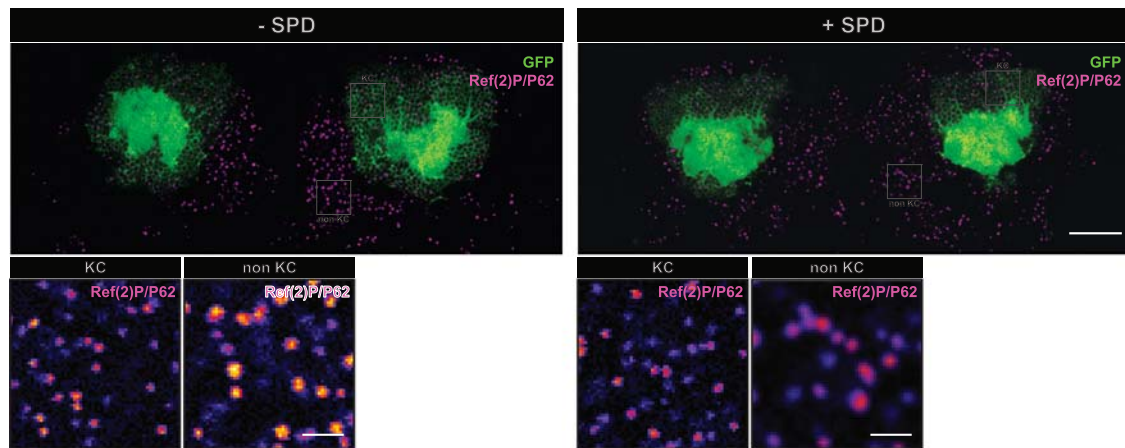
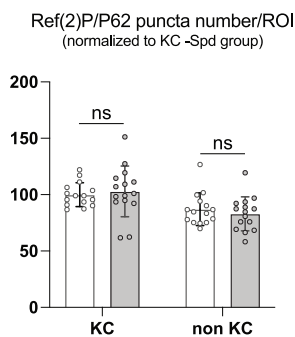
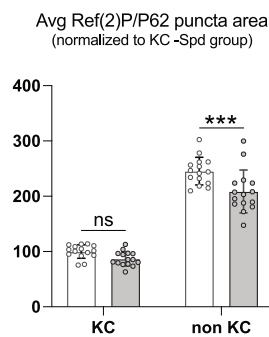
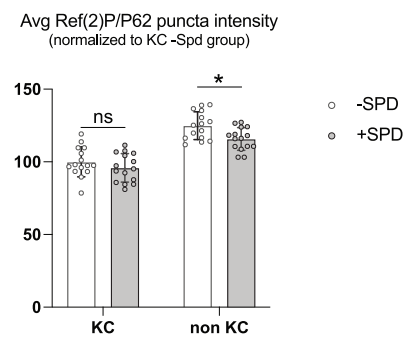
A*vt30559-Gal4 > UAS-mcd8-GFP::UAS-Atg5-RNAi***B****C****D**

Figure EV4. Related to Fig. 8, 5 days spermidine (Spd) supplementation significantly reduces the size and intensity of non-cell autonomous Ref(2)P/P62 aggregates outside the MB KCs.

Freshly eclosed *vt30559-Gal4>UAS-mcd8-GFP::UAS-Atg5-RNAi* flies were split at day 1 and either transferred onto 5 mM spermidine food or maintained on control food for 5 days. Female fly brains were subsequently dissected and immunostained against Ref(2)P/P62 for confocal microscopic analysis. (A) Representative single optical slices of the posterior MB calyx region, exemplifying chosen regions of interest (ROIs) for the analysis of Ref(2)P/P62 puncta in the MB Kenyon cells (KC) and in non-MB neurons (non-KC) of the same optical slice. MB KCs were identified based on the endogenous *mcd8-GFP* signal driven by *vt30559-Gal4*. Scale bar upper panels = 25 μ m; Scale bar lower panels = 5 μ m (B–D) Quantification of Ref(2)P/P62 puncta number (B), area (C), and intensity (D) for 5-day-old spermidine-treated (+Spd) and non-treated (-Spd) *vt30559-Gal4>UAS-mcd8-GFP::UAS-Atg5-RNAi* flies. The average number, area and intensity of Ref(2)P/P62 spots per individual brain was calculated from a total number of 8 ROIs per each brain region (i.e., KC and non-KC), retrieved from two individual optical slices. Measured data from two independent experiments was pooled by normalizing the measured values onto the KC - Spd group. Only relevant statistical comparisons between Spd-supplemented (+Spd) and untreated (-Spd) groups are displayed. The selection of ROIs and suitable optical slices was performed blinded. $n = 15$ adult brains per treatment, pooled from two independent experiments. One-way ANOVA with Tukey's post hoc test for multiple comparisons. * $p = 0.0487$, *** $p = 0.0009$, ns not significant. Data presented as mean $\pm$ SD. Source data are available online for this figure.