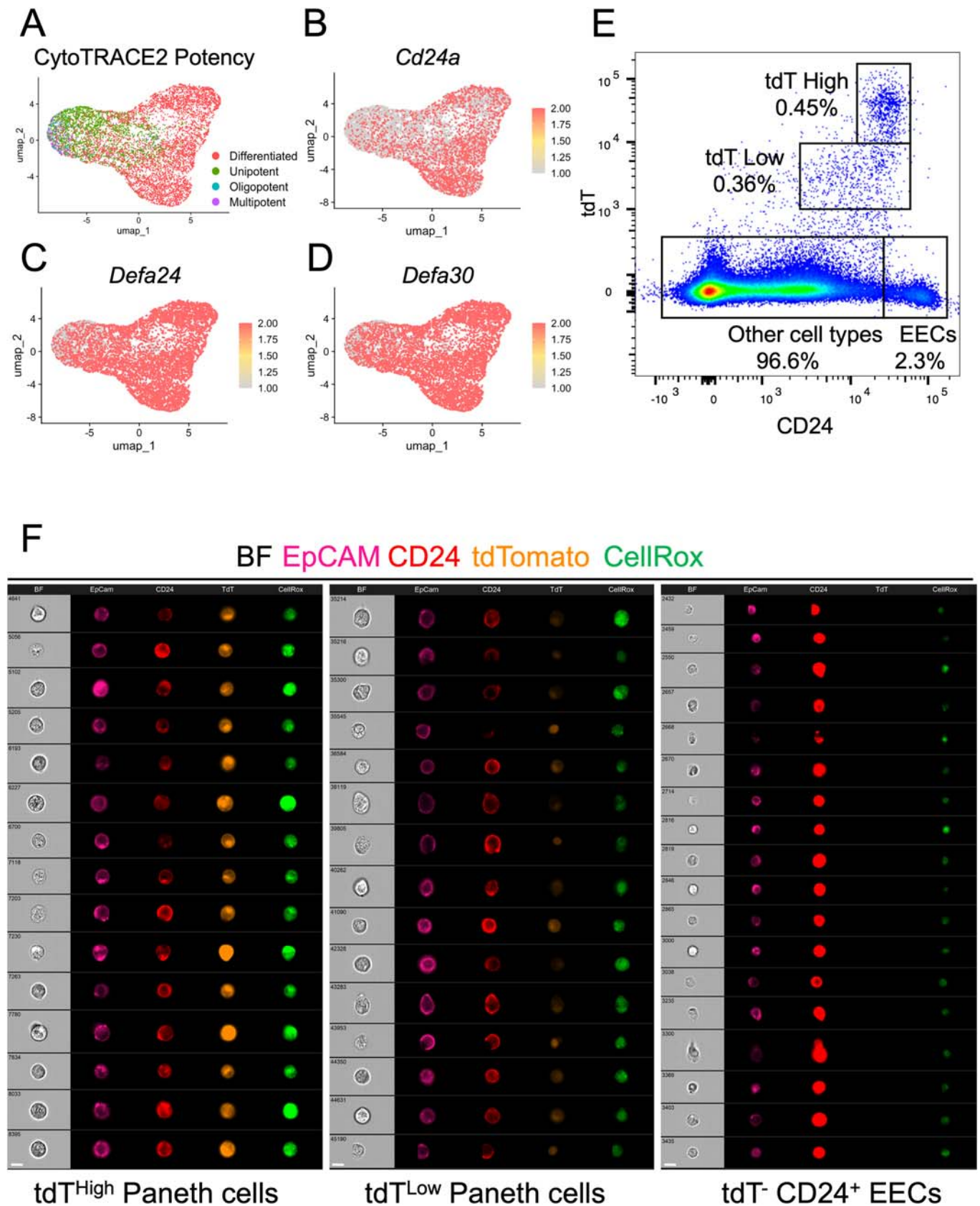


Expanded View Figures

Figure EV1. Intracellular ROS levels vary across Paneth cell populations.

(A) CytoTRACE potency was calculated for individual Paneth cells. (B–D) UMAP for Cd24a, Defa24, and Defa30. (E) Live intestinal epithelial cells were resolved by the expression of tdT and CD24. (F) Imagestream analysis for CellROX was performed for tdT^{High} and tdT^{Low} Paneth cells, tdT⁺CD24⁺ EECs and tdT⁺CD24⁺ IECs. Scale bar, 10 μ m.



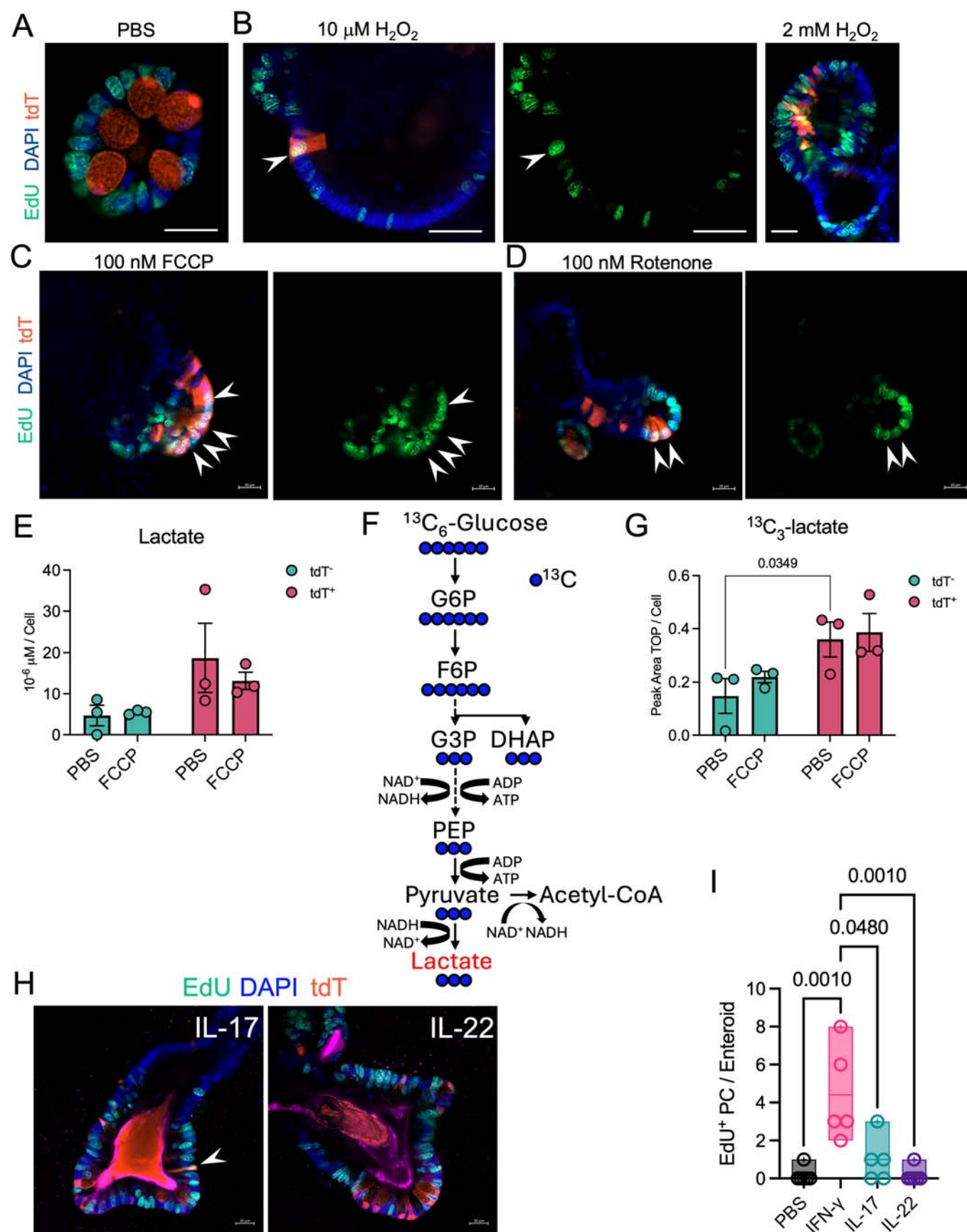


Figure EV2. Impact of oxidative stress on cell-cycle entry and metabolism in Paneth cells.

(A–D) Enteroids were grown from PC reporter mouse duodenum, induced for tdT expression on day 3, and treated with various ROS for 24 h. Enteroids were labeled with EdU for 1 h before fixation and imaging. Scale bar, 20 μ m. Arrowheads point to EdU⁺tdT⁺ cells. Scale bar, 20 μ m. (E) Approximately 1000 duodenal enteroids from PC reporter mice ($n = 3$) were treated with FCCP for 24 h, washed out, and 17.5 mM $^{13}\text{C}_6$ -glucose was added to the medium for a duration of 17 h before washing out and proceeding to FACS sorting. tdT⁺ and tdT⁻ IECs were isolated for LC-MS analysis. Lactate concentrations were normalized against the cell number of individual samples and plotted. Data are presented as mean $\pm$ SEM. (F) Schematic diagram illustrating the glycolysis of glucose to pyruvate, which is either converted to lactate through anaerobic respiration or enters mitochondria to fuel the TCA cycle. (G) Using samples from the experiment described in (E), $^{13}\text{C}_3$ -lactate abundance was quantified from individual samples treated with PBS or FCCP. A two-way ANOVA followed by Fisher's LSD test was performed to compare the difference. Exact p -value is provided above the comparison bar. Data are presented as mean $\pm$ SEM. (H) Enteroids were grown from PC reporter mouse duodenum, induced for tdT expression on day 3, and treated with 50 ng/mL IL-17 or 4 ng/mL IL-22 for 24 h. Enteroids were labeled with EdU for 1 h before fixation and imaging. Scale bar, 20 μ m. Arrowheads point to an EdU⁺tdT⁺ cell in the IL-17-treated condition. (I) The number of EdU⁺tdT⁺ cells was calculated for individual enteroids treated with PBS ($n = 6$), IFN- γ ($n = 5$), IL-17 ($n = 6$), or IL-22 ($n = 5$). Statistical comparisons between groups were performed using a Kruskal-Wallis test followed by an uncorrected Dunn's multiple comparisons test. Exact p -values are provided above the comparison bars. Data are presented as floating bars (min to max) with the central line indicating the mean.

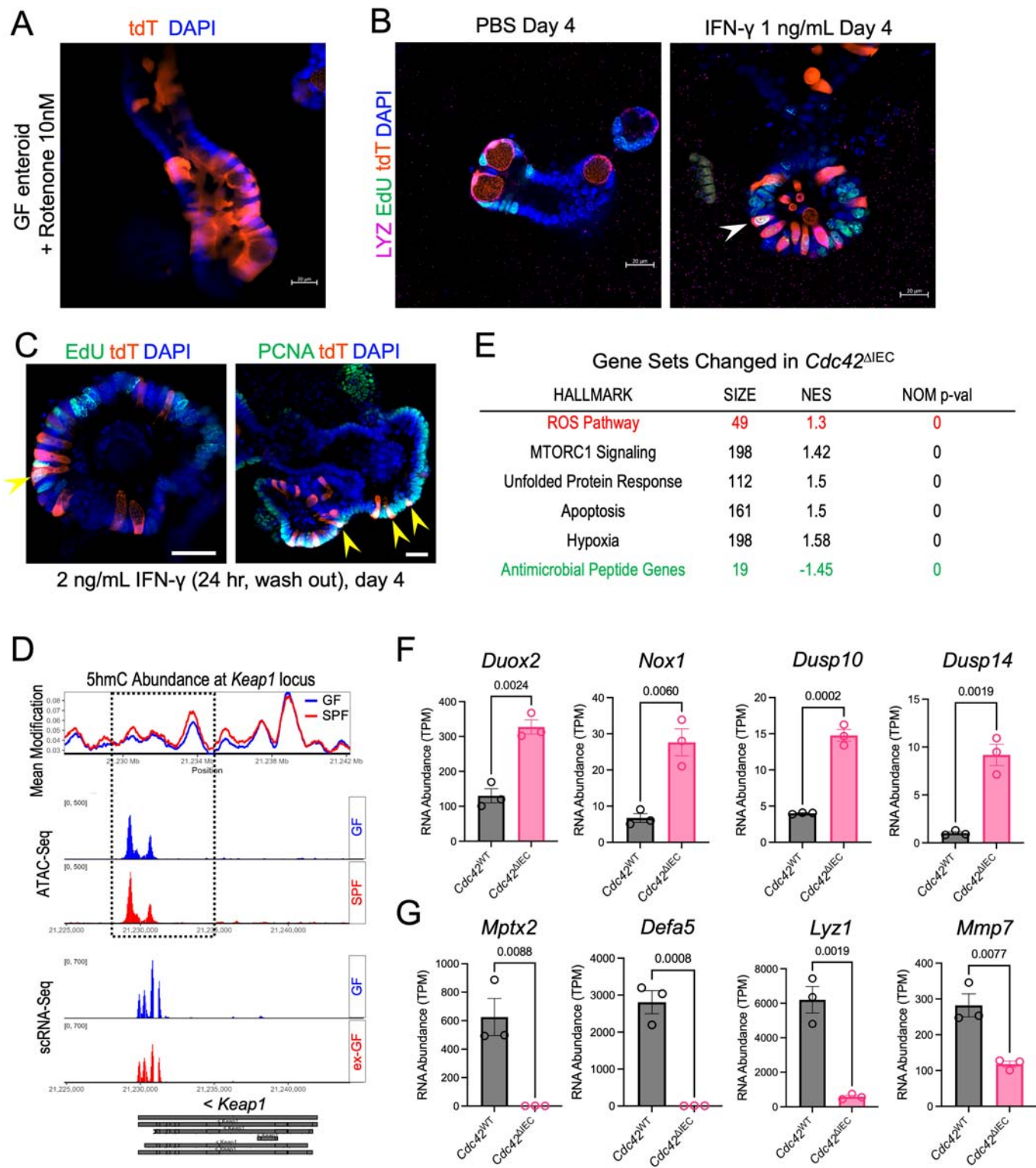


Figure EV3. *Cdc42* deficiency upregulates the ROS pathway and suppresses the AMP gene program.

(A) Enteroids were grown from GF Paneth cell reporter mice. On day 3, 4-OHT was added, and on the following day enteroids were treated with 10 nM rotenone. The image was taken of an enteroid that showed a tdT⁺ stripe. Scale bar, 20 μ m. (B) Enteroids from Paneth cell reporter mouse duodenum were continuously treated with 1 ng/mL IFN- γ or PBS for 4 days, labeled with EdU 1 h before fixation, and stained for lysozyme, EdU, tdT, and DAPI. The white arrow indicates a LYZ⁺EdU⁺tdT⁺ cell. Scale bar, 20 μ m. (C) Enteroids from Paneth cell reporter mouse duodenum were treated with 2 ng/mL IFN- γ for 24 h, washed out, then cultured to day 4, labeled with EdU for 1 h, fixed and stained for lysozyme, EdU, tdT, and DAPI; or PCNA, tdT, and DAPI. Yellow arrows indicate PCNA⁺tdT⁺ cells. Scale bar, 20 μ m. (D) Paneth cell specific multi-modality omics for 5hmC DNA nanopore-sequencing, ATAC-seq and scRNA-seq of Paneth cells from GF and SPF PC reporter mice. (E) Bulk RNA-seq was performed using WT vs. *Cdc42* ^{Δ IEC} mouse small intestinal epithelia ($n = 3$ per condition). Hallmark gene sets for reactive oxygen species (red) and antimicrobial peptides (green) are significantly increased and decreased in *Cdc42* ^{Δ IEC} mice, respectively. (F) RNA abundances of ROS production and regulatory enzymes in *Cdc42*^{WT} ($n = 3$) and *Cdc42* ^{Δ IEC} ($n = 3$) mice are shown. A two-tailed Student's *t*-test was used to compare the differences. Exact *p*-values are provided above the comparison bars. Data are presented as mean $\pm$ SEM. (G) RNA abundances of AMP genes in *Cdc42*^{WT} ($n = 3$) and *Cdc42* ^{Δ IEC} ($n = 3$) mice. A two-tailed Student's *t*-test was used to compare the differences. Exact *p*-values are provided above the comparison bars. Data are presented as mean $\pm$ SEM.

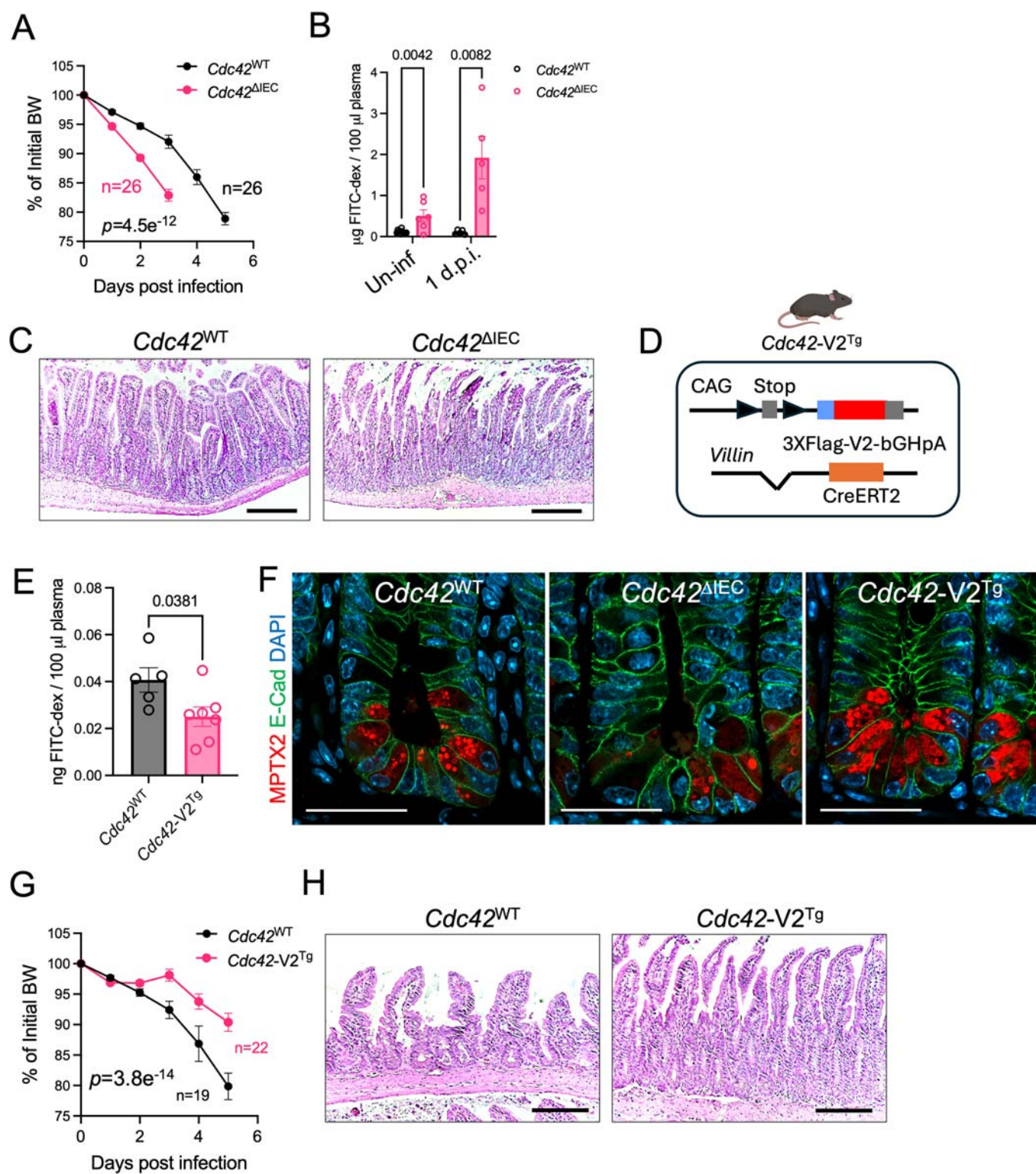


Figure EV4. Cdc42 deficiency increases susceptibility to *S. Typhimurium* infection.

(A) Body weight changes of *Cdc42^{WT}* and *Cdc42^{ΔIEC}* littermates following the *S. Typhimurium* oral inoculation. $n = 26$ for each condition. Curves were modeled using exponential (Malthusian) growth regression. Curves were compared using the Extra sum-of-squares F test. Data are presented as mean $\pm$ SEM. (B) An in vivo gut permeability assay was performed before the mice were sacrificed. Two hours after oral dosing of FITC-conjugated 4 kDa dextran, nanograms of dextran circulating in the plasma of uninfected mice and mice infected for one day were detected, for *Cdc42^{WT}* and *Cdc42^{ΔIEC}* mice. Statistical comparisons were performed using multiple unpaired *t*-tests with the two-stage step-up method of Benjamini, Krieger, and Yekutieli. Exact *p*-values are provided above the comparison bars. Data are presented as mean $\pm$ SEM. (C) Histology of infected *Cdc42^{WT}* and *Cdc42^{ΔIEC}* mouse ileum at 1 d.p.i. Scale bar, 100 μ m. (D) Schematic diagram showing that *Cdc42-V2^{Tg}*; Villin-CreER^{T2} mice overexpress the CDC42 isoform. (E) Mice were orally gavaged with FITC-conjugated 4 kDa dextran 2 h before sacrifice. Nanograms of fluorescent dextran were measured in plasma of *Cdc42^{WT}* ($n = 5$) and *Cdc42-V2^{Tg}* ($n = 7$) mice. A Student's *t*-test was used to compare the difference. Exact *p*-value is provided above the comparison bar. Data are presented as mean $\pm$ SEM. (F) Immunofluorescent staining was performed for E-Cad and MPTX2 in *Cdc42^{WT}*, *Cdc42^{ΔIEC}*, and *Cdc42-V2^{Tg}* mice. Scale bar, 50 μ m. (G) Body weights were measured from infected *Cdc42-V2^{Tg}* ($n = 22$) and *Cdc42^{WT}* ($n = 19$) littermates. Curves were modeled using exponential (Malthusian) growth regression. Curves were compared using the Extra sum-of-squares F test. Data are presented as mean $\pm$ SEM. (H) Histology of infected *Cdc42^{WT}* and *Cdc42-V2^{Tg}* mouse ileum. Scale bar, 100 μ m.

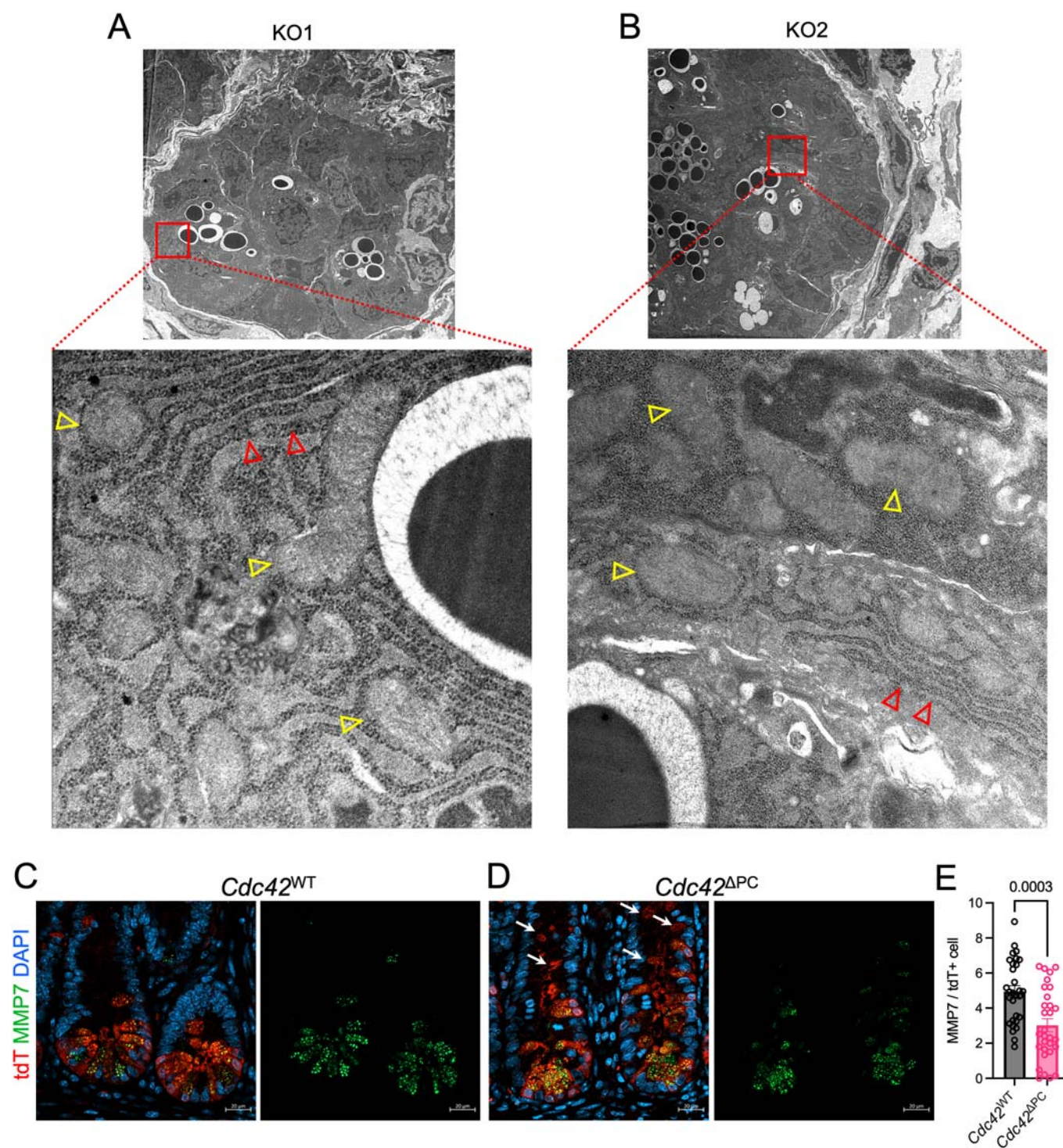


Figure EV5. PC-specific deletion of *Cdc42* impairs mitochondrial integrity and MMP7 production.

(A, B) TEM micrographs of *Cdc42*^{ΔPC} mouse ilea. These representative images illustrate the severe mitochondrial cristolysis (yellow arrows) and pronounced ER lumen dilation (red arrows) in virtually all the PCs of *Cdc42*^{ΔPC} mice. (C, D) Confocal immunofluorescence analysis of MMP7 and tdT in *S. Typhimurium* infected *Cdc42*^{WT} and *Cdc42*^{ΔPC} mice. Arrows in panel F point to up-migrating tdT⁺ cells that do not express MMP7. (E) MMP7 abundances in individual tdT⁺ cells were quantified from *Cdc42*^{WT} and *Cdc42*^{ΔPC} mice, and statistically compared by a Student's *t*-test. Exact *p*-value is provided above the comparison bar. Data are presented as mean ± SEM.

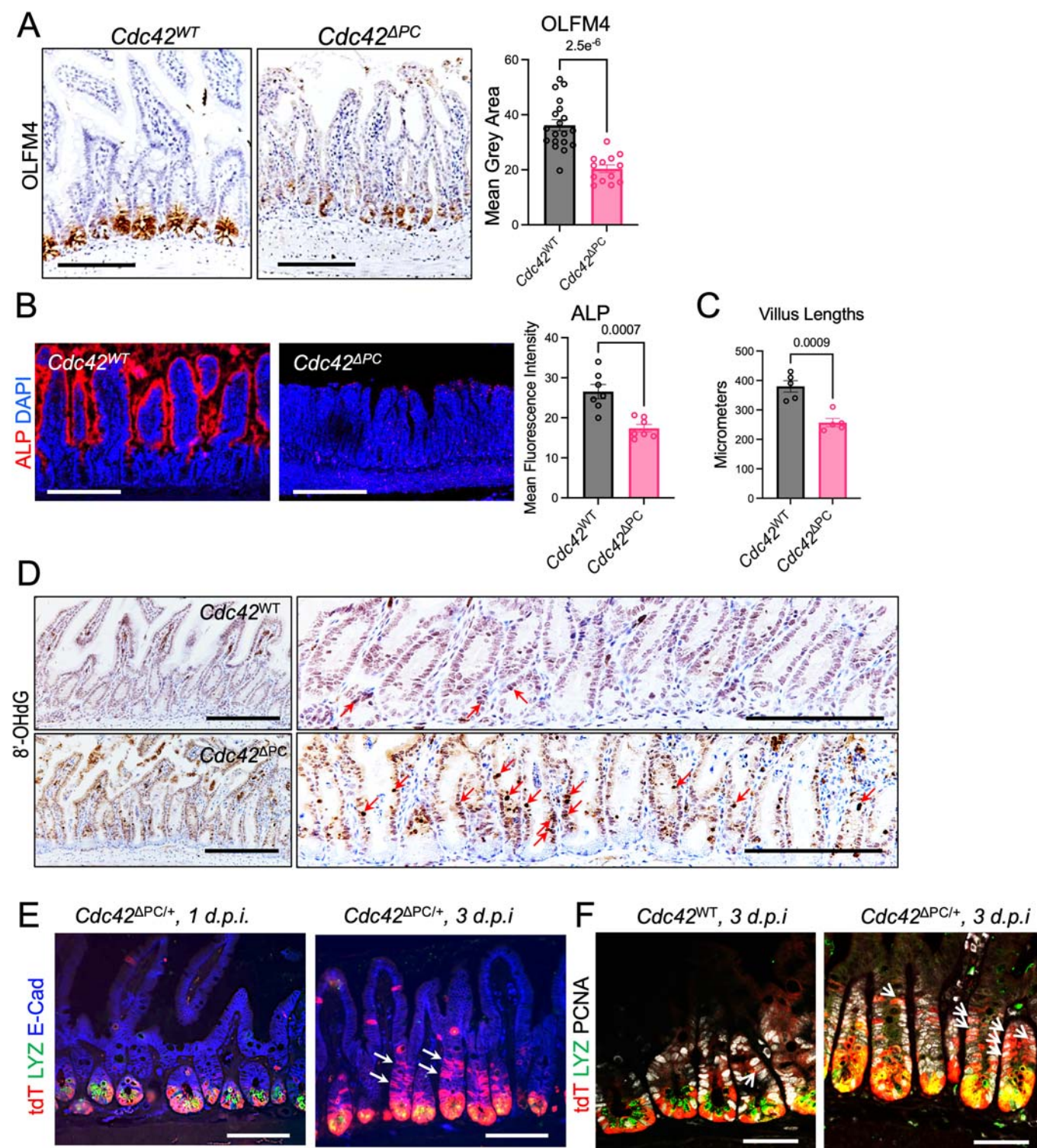


Figure EV6. PC-specific *Cdc42* knockout reduces stem cell and enterocyte populations and elevates DNA oxidation during *S. Typhimurium* infection.

(A) Immunostaining was performed for OLFM4 in infected *Cdc42*^{WT} and *Cdc42*^{ΔPC} mouse intestines. A Student's *t*-test was performed to compare OLFM4 abundance in crypts between the two groups. Exact *p*-value is provided above the comparison bar. Scale bar, 100 μm. Data are presented as mean ± SEM. (B) Immunostaining was performed for alkaline phosphatase (ALP) in infected *Cdc42*^{WT} and *Cdc42*^{ΔPC} intestines. Scale bar, 100 μm. A Student's *t*-test was performed to compare the ALP between the two groups. Exact *p*-value is provided above the comparison bar. Data are presented as mean ± SEM. (C) Villus lengths were measured from the infected *Cdc42*^{WT} and *Cdc42*^{ΔPC} mice at 4 d.p.i. and analyzed by a Student's *t*-test. Exact *p*-value is provided above the comparison bar. Data are presented as mean ± SEM. (D) Immunostaining for 8'-OHdG using infected *Cdc42*^{WT} and *Cdc42*^{ΔPC} intestinal sections. Red arrows point to 8'-OHdG puncta. Scale bar, 100 μm. (E) Tamoxifen was injected into *Cdc42*^{ΔPC/+} mice carrying a PC reporter. Mice were infected with *S. Typhimurium* and sacrificed at 1 and 3 d.p.i. Sections were stained for tdT and lysozyme. Arrows point to up-migrating tdT⁺ cells. Scale bar, 50 μm. (F) Tamoxifen was injected into *Cdc42*^{WT} and *Cdc42*^{ΔPC/+} carrying a PC reporter. Mice were infected with *S. Typhimurium* and sacrificed at 3 d.p.i. Sections were stained for tdT, lysozyme, and PCNA. Arrows point to PCNA⁺tdT⁺ cells. Scale bar, 50 μm.