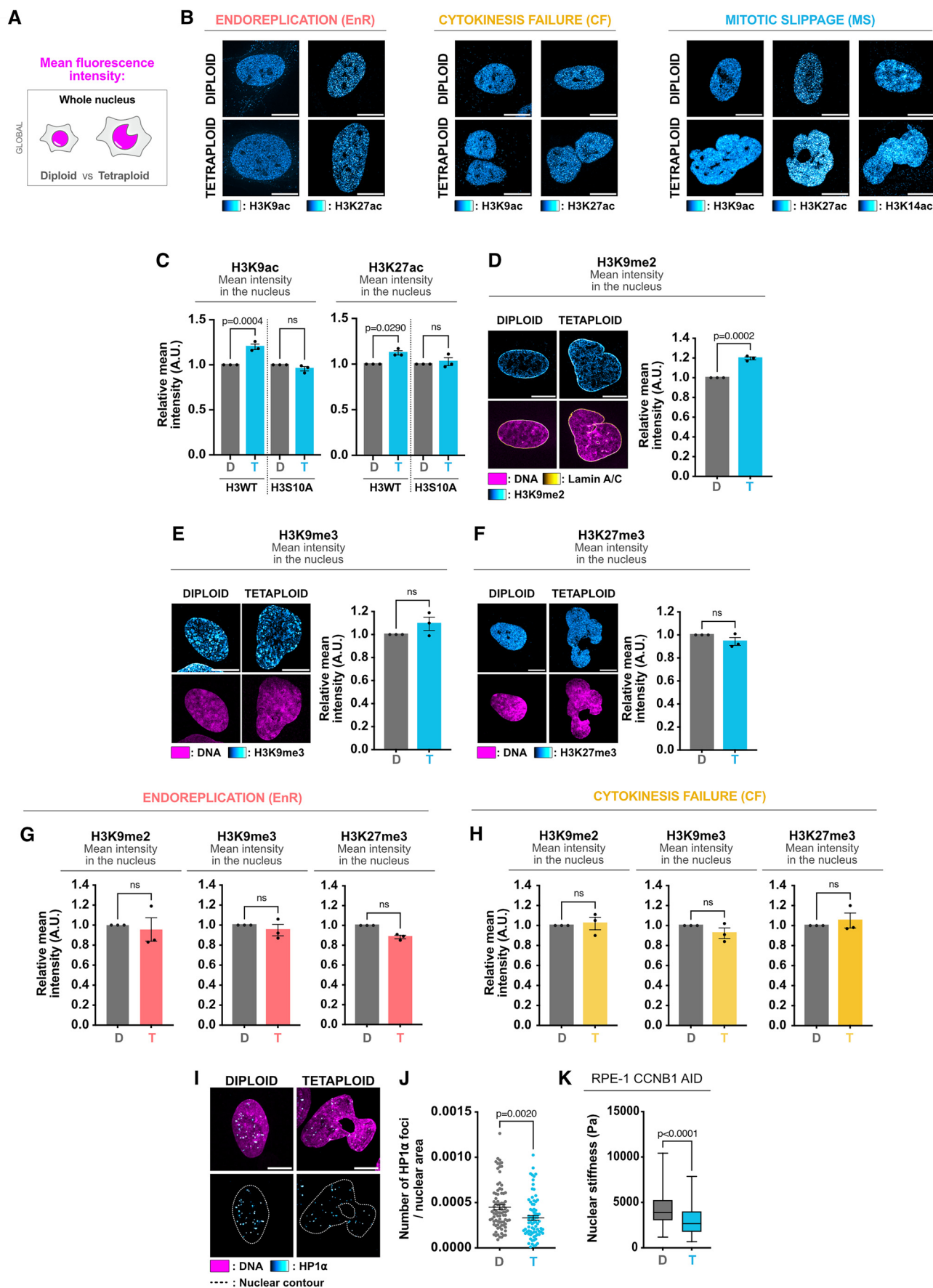


Expanded View Figures

Figure EV1. Mitotic slippage, but not cytokinesis failure and endoreplication, alters histone modification.

(A) Schematic representation of the mean fluorescence measured at the level of the whole nucleus in diploid and tetraploid cells. (B) SIM representative images showing diploid and tetraploid G1 RPE-1 cells generated by EnR (left panel), by CF (central panel) or by MS (right panel). H3K9ac, H3K27ac and H3K14ac in cyan hot. (C) Graphs showing the relative H3K9ac (left panel) or H3K27ac (right panel) mean intensity in the nucleus in diploid (in grey) and in tetraploid (in blue) RPE-1 G1 cells stably expressing H3 WT or H3S10A. Mean $\pm$ SEM, >100 G1 cells analysed per condition, three independent experiments. (D) Left panel—SIM representative images showing diploid and tetraploid G1 RPE-1 cells generated by MS. Lamin A/C in yellow hot, H3K9me2 in cyan hot and DNA in magenta. Right panel—Graph presenting the relative H3K9me2 mean intensity in the nucleus in diploid (in grey) and in tetraploid (in blue) G1 RPE-1 cells. Mean $\pm$ SEM, >100 G1 cells analysed per condition, three independent experiments. (E) Left panel—SIM representative images showing diploid and tetraploid G1 RPE-1 cells generated by MS. Lamin A/C in yellow hot, H3K9me3 in cyan hot and DNA in magenta. Right panel—Graph presenting the relative H3K9me3 mean intensity in the nucleus in RPE-1 diploid (in grey) and in tetraploid (in blue) G1 cells. Mean $\pm$ SEM, >100 G1 cells analysed per condition, three independent experiments. (F) Left panel—SIM representative images showing diploid and tetraploid G1 RPE-1 cells generated by MS. Lamin A/C in yellow hot, H3K27me3 in cyan hot and DNA in magenta. Right panel—Graph presenting the Relative H3K27me2 mean intensity in the nucleus in diploid (in grey) and in tetraploid (in blue) G1 RPE-1 cells. Mean $\pm$ SEM, >100 G1 cells analysed per condition, three independent experiments. (G, H) Graphs showing the relative H3K9me2, H3K9me3 and H3K27me3 mean intensity in the nucleus in diploid (in grey) and in tetraploid G1 RPE-1 cells generated by (G) EnR (in red) or by (H) CF (in yellow). Mean $\pm$ SEM, >100 G1 cells analysed per condition, three independent experiments. (I) SIM representative images showing diploid and tetraploid G1 RPE-1 cells generated by MS. HP1 α in cyan hot and DNA in magenta. (J) Graph showing the number of HP1 foci / nuclear area in diploid (in grey) and in tetraploid (in blue) RPE-1 G1 cells. Mean $\pm$ SEM, >90 G1 cells analysed per condition, at least three independent experiments. (K) Graph showing nuclear stiffness in diploid and in MS-generated tetraploid G1 RPE-1 CENB1 cells (in grey and blue, respectively). MS cells were generated by depleting CENB1. Box and whiskers $\pm$ Min to Max, >175 G1 cells analysed per condition, three independent experiments. Scale bars: 10 μ m. D diploid, T tetraploid, MS mitotic slippage, EnR endoreplication, CF cytokinesis failure, AU arbitrary unit. (C–H, J) t-test (two-sided). (K) Kolmogorov–Smirnov test. ns not significant.



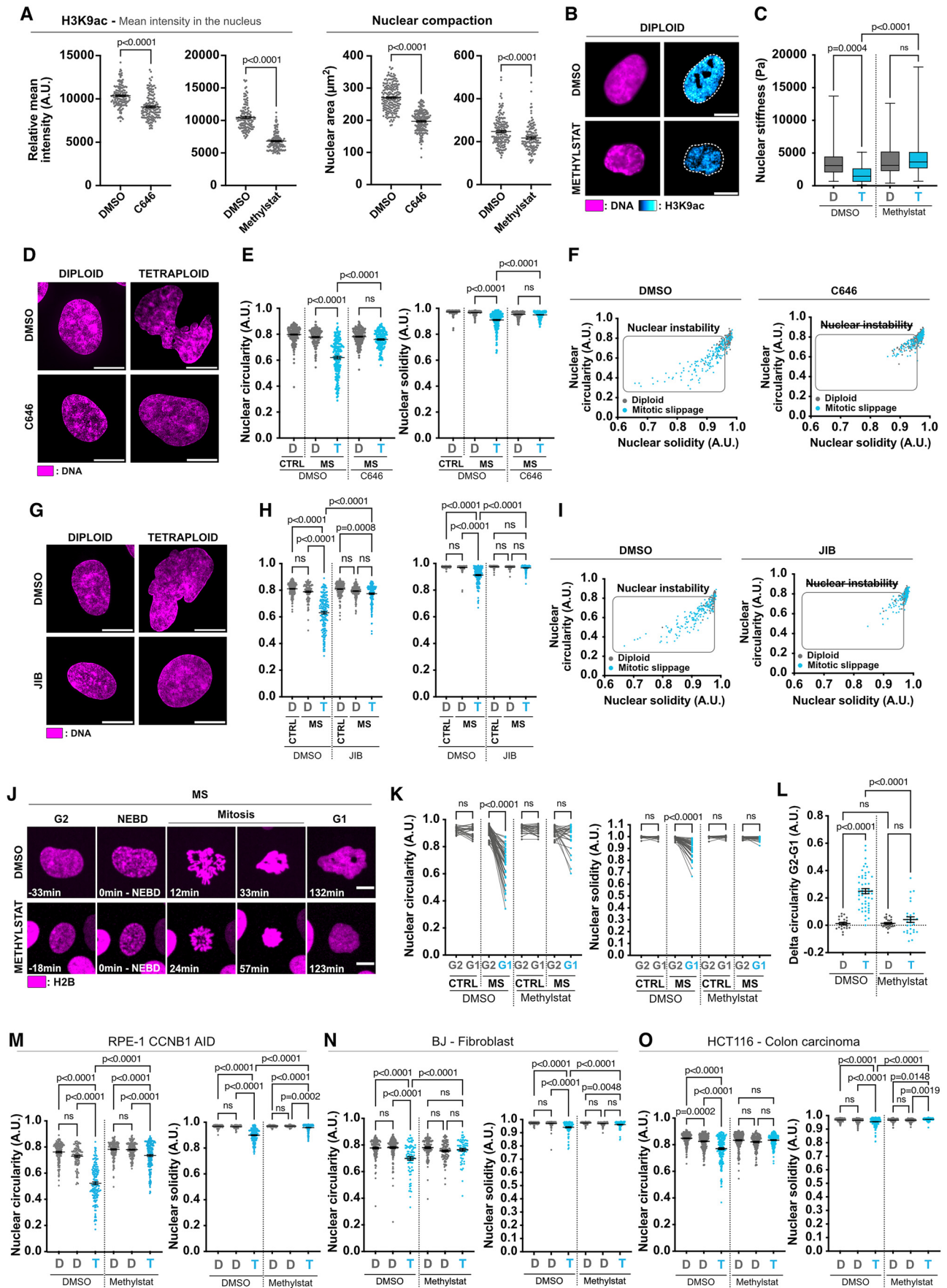


Figure EV2. Promoting nuclear compaction is sufficient to prevent nuclear deformations upon mitotic slippage.

(A) Graphs presenting the H3K9ac mean intensity (left panel) or the nuclear area (right panel) in diploid RPE-1 cells. Mean $\pm$ SEM, >125 cells analysed per condition, at least three independent experiments. (B) Representative images showing RPE-1 diploid cells treated with DMSO or 4 μ M Methylstat. H3K9ac in cyan hot, DNA in magenta. (C) Graph showing the nuclear stiffness in diploid (in grey) and tetraploid (in blue) RPE-1 G1 cells generated by MS treated with DMSO or 4 μ M Methylstat. (D) Representative images showing diploid and tetraploid RPE-1 G1 cells treated with DMSO or 20 μ M C646. DNA in magenta. (E) Graphs showing the nuclear circularity (left panel) and solidity (right panel) indices in diploid (in grey) and tetraploid (in blue) RPE-1 G1 cells generated by MS treated with DMSO or 20 μ M C646. Mean $\pm$ SEM, >125 G1 cells analysed per condition, at least three independent experiments. (F) Graphs presenting the nuclear circularity and solidity from (E) in RPE-1 diploid (in grey) and tetraploid (in blue) G1 cells treated with DMSO or 20 μ M C646. (G) Representative images showing diploid and tetraploid RPE-1 cells treated with DMSO or 5 μ M JIB. DNA in magenta. (H) Graphs presenting the nuclear circularity (left panel) and solidity (right panel) indices in diploid (in grey) and tetraploid (in blue) RPE-1 G1 cells generated by MS treated with DMSO or 5 μ M JIB. Mean $\pm$ SEM, >100 G1 cells analysed per condition, at least three independent experiments. (I) Graphs presenting the nuclear circularity and solidity from (H) in diploid (in grey) and tetraploid (in blue) RPE-1 G1 cells treated with DMSO or 5 μ M JIB. (J) Stills from time-lapse imaging of RPE-1 cells stably expressing mCherry-H2B (in magenta) performing MS and treated with DMSO (upper panel) or 2 μ M Methylstat (lower panel). (K) Graphs showing the nuclear circularity (left panel) and solidity (right panel) measured at the single cell levels in G2 and in G1 upon mitotic division (in grey) or MS (in blue) in RPE-1 cells treated with DMSO or 2 μ M Methylstat. >30 cells analysed per condition, three independent experiments. (L) Graph showing the delta circularity (defined as the difference between nuclear circularity in G2 vs in G1) upon mitotic division (in grey) or MS (in blue) in RPE-1 cells treated with DMSO or 2 μ M Methylstat. Mean $\pm$ SEM, >25 cells analysed per condition, three independent experiments. (M–O) Graphs displaying nuclear circularity and solidity in (M) RPE-CCNB1, (N) BJ or (O) HCT116 cells treated with DMSO or 4 μ M Methylstat. (M) Tetraploid cells were generated by depleting CCNB1. Mean $\pm$ SEM, >65 G1 cells analysed per condition, three independent experiments. Scale bars: 10 μ m. D diploid, T tetraploid, MS mitotic slippage, NEBD nuclear envelope breakdown, AU arbitrary unit. (A, K, L) *t*-test (two-sided). (E, H, M–O) ANOVA test (one-sided). (C) Kolmogorov–Smirnov test. ns not significant.

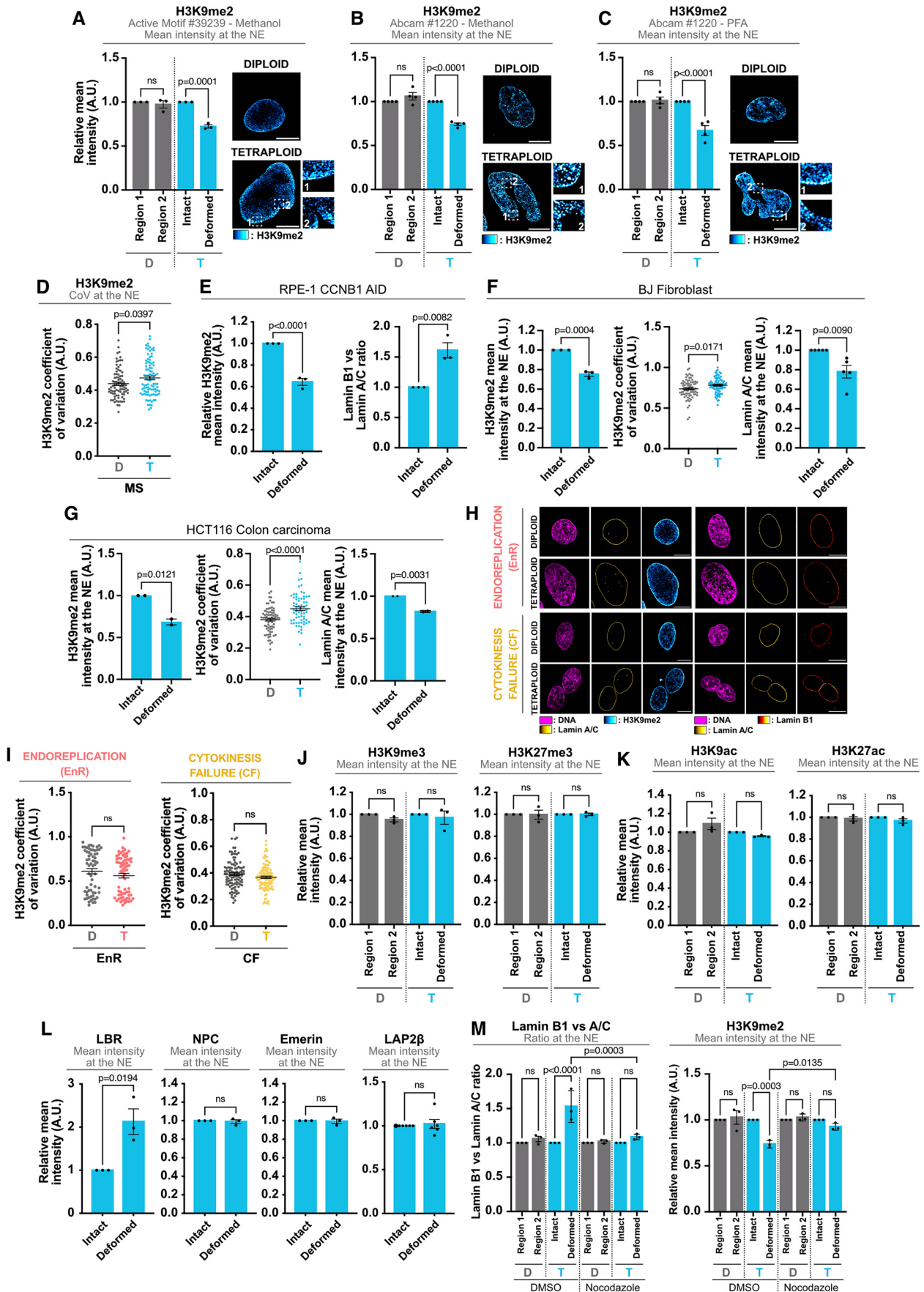


Figure EV3. Mitotic slippage promotes local changes in the Lamina network and in H3K9me2 levels.

(A) Left panel—Graph showing the relative H3K9me2 mean intensity at the nuclear envelope in diploid (in grey) and tetraploid (in blue) G1 RPE-1 cells. Mean $\pm$ SEM, >90 G1 cells analysed per condition, three independent experiments. Right panel—SIM representative images showing diploid and tetraploid G1 RPE-1 cells. H3K9me2 in cyan hot. (B) Left panel—Graph showing the relative H3K9me2 mean intensity at the nuclear envelope in diploid (in grey) and tetraploid (in blue) G1 RPE-1 cells. Mean $\pm$ SEM, >90 G1 cells analysed per condition, three independent experiments. Right panel—SIM representative images showing diploid and tetraploid G1 RPE-1 cells. H3K9me2 in cyan hot. (C) Left panel—Graph showing the relative H3K9me2 mean intensity at the nuclear envelope in diploid (in grey) and tetraploid (in blue) G1 RPE-1 cells. Mean $\pm$ SEM, >90 G1 cells analysed per condition, three independent experiments. Right panel—SIM resolution representative images showing diploid and tetraploid G1 RPE-1 cells. H3K9me2 in cyan hot. (D) Graph presenting the H3K9me2 coefficient of variation at the nuclear envelope in MS-induced tetraploid G1 RPE-1 cells. Mean $\pm$ SEM, >90 G1 cells analysed per condition, three independent experiments. (E) Graphs showing the relative H3K9me2 mean intensity (left panel) or the Lamin B1 vs A/C ratio (right panel) at the nuclear envelope in MS-induced tetraploid RPE-1 CCNB1 cells. MS cells were generated by depleting CCNB1. Mean $\pm$ SEM, >90 G1 cells analysed per condition, three independent experiments. (F, G) Graphs of H3K9me2 mean intensity at the NE (left panel), H3K9me2 coefficient of variation at the NE (central panel) and Lamin A/C mean intensity at the NE (right panel) in diploid and in MS-generated tetraploid (F) BJ and (G) HCT116 cells. Mean $\pm$ SEM, >60 G1 cells analysed per condition, three independent experiments. (H) SIM representative images showing diploid and tetraploid cells generated by EnR or CF. H3K9me2, Lamin A/C and Lamin B1 in cyan hot, yellow hot and red hot, respectively. DNA in magenta. (I) Graphs showing the H3K9me2 coefficient of variation in diploid (in grey) and tetraploid G1 RPE-1 cells generated by EnR (in red) or CF (in yellow). Mean $\pm$ SEM, >90 G1 cells analysed per condition, three independent experiments. (J) Graphs showing the relative H3K9me3 (left panel) or H3K27me3 (right panel) mean intensity in diploid (in grey) and tetraploid (in blue) RPE-1 G1 cells. Mean $\pm$ SEM, >90 G1 cells analysed per condition, three independent experiments. (K) Graphs showing the relative H3K9ac (left panel) or H3K27ac (right panel) mean intensity in diploid (in grey) and tetraploid (in blue) RPE-1 G1 cells. Mean $\pm$ SEM, >90 G1 cells analysed per condition, three independent experiments. (L) Graphs presenting the relative mean intensity at the nuclear envelope of the indicated proteins in tetraploid RPE-1 G1 cells. Mean $\pm$ SEM, >90 G1 cells analysed per condition, at least three independent experiments. (M) Graphs showing the Lamin B1 vs A/C ratio (left panel) or the H3K9me2 mean intensity (right panel) at the nuclear envelope in diploid (in grey) and tetraploid (in blue) RPE-1 G1 cells treated with DMSO or with Nocodazole. The same control was used in Fig. 5E because the experiments were done together. Mean $\pm$ SEM, >90 G1 cells analysed per condition, three independent experiments. Scale bars: 10 μ m. D diploid, T tetraploid, MS mitotic slippage, EnR endoreplication, CF cytokinesis failure, AU arbitrary unit. (M) ANOVA test (one-sided). (A–G, I–L) t-test (two-sided). ns not significant.

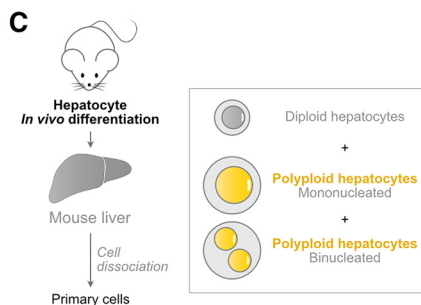
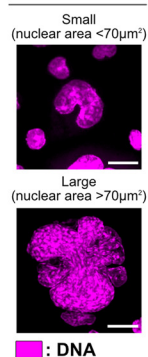
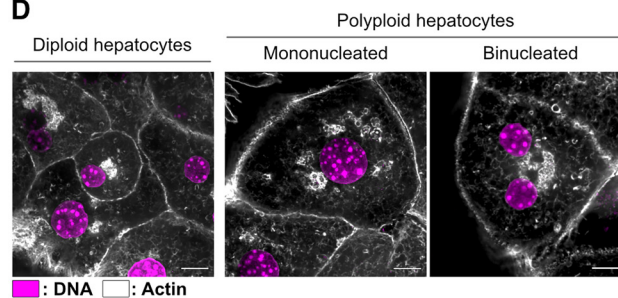
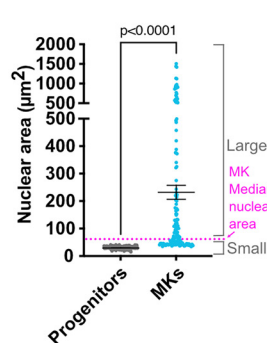
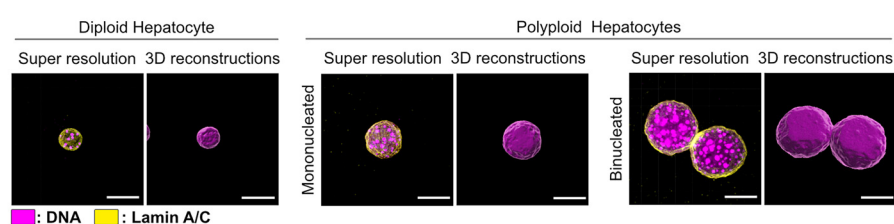
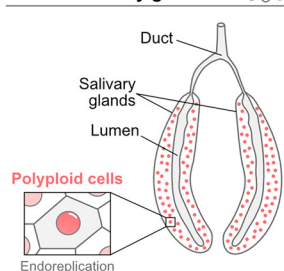
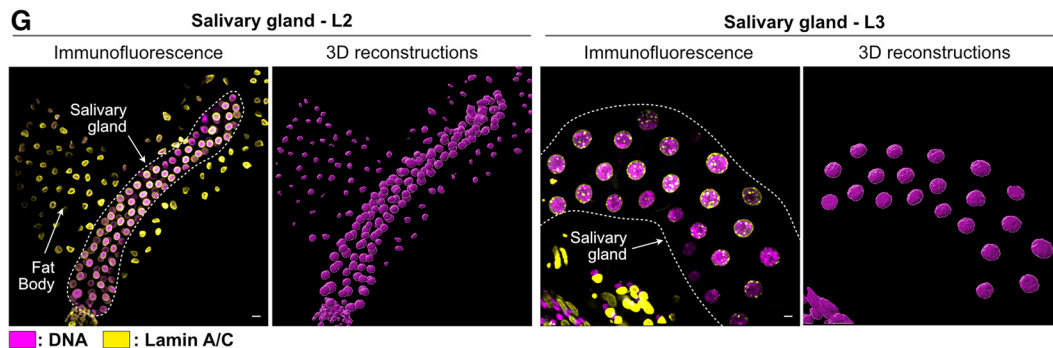
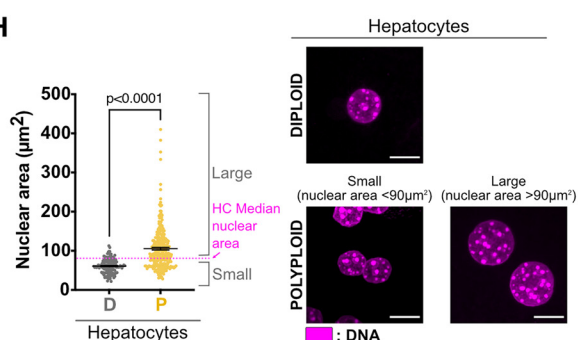
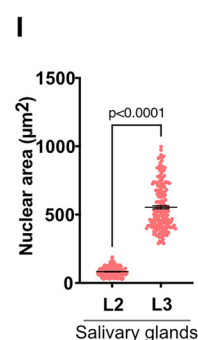
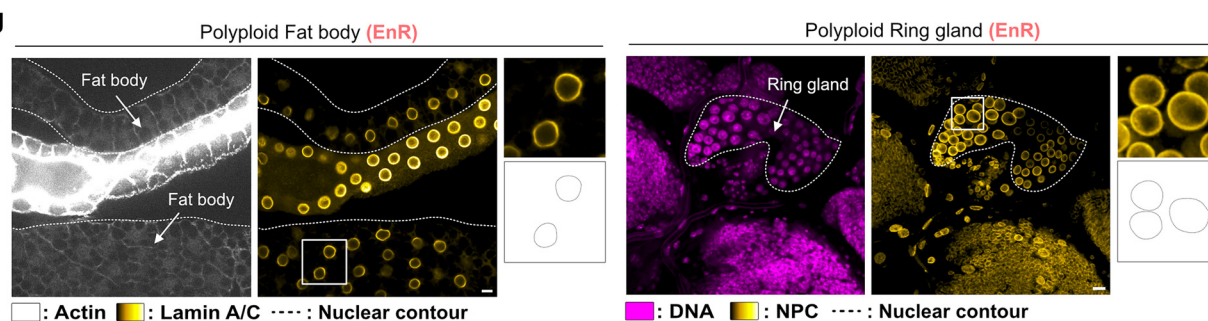
A MEGAKARYOCYTES**D****B****E****F** Larval *Drosophila* Salivary gland**G****H****I****J**

Figure EV4. Physiological polyploid cells generated by cytokinesis failure and endoreplication show homogenous nuclear shape.

(A) Representative images of small and large MKs. DNA in magenta. (B) Graph showing nuclear area in diploid progenitors and in polyploid MKs. The MK median nuclear area was used to distinguish small and large MKs. Mean $\pm$ SEM, >150 interphase cells analysed per condition, three independent experiments. (C) Schematic workflow showing the methods used to obtain mouse hepatocytes. (D) Representative images showing diploid and polyploid hepatocytes. Actin in grey, DNA in magenta. (E) SIM representative images and their corresponding 3D reconstructions showing diploid and polyploid hepatocytes. Lamin A/C in yellow, DNA in magenta. (F) Schematic representation of *Drosophila* salivary glands. (G) Representative images and their corresponding 3D reconstructions showing polyploid salivary glands. Lamin A/C in yellow, DNA in magenta. (H) Left panel—Graph showing the nuclear area in diploid and in polyploid hepatocytes (in grey and yellow, respectively). The hepatocyte median nuclear area was used to distinguish small and large hepatocytes. Mean $\pm$ SEM, >140 interphase cells analysed per condition, three independent experiments. Right panel—Representative images showing diploid and polyploid hepatocytes. DNA in magenta. (I) Graph showing the nuclear area in polyploid salivary glands at two different developmental stages (L2 and L3). Mean $\pm$ SEM, >100 cells analysed, three independent experiments. (J) Representative immunofluorescence images showing *Drosophila* fat body (left panel) and ring gland (right panel). Lamin A/C or Nuclear pore complex (NPC) in yellow hot, Actin in grey, DNA in magenta. Scale bars: 10 μ m. D diploid. P polyploid, MK megakaryocyte, NPC nuclear pore complex. (B, H, I) *t*-test (two-sided). ns not significant.



◀ **Figure EV5. Polyploid hepatocytes and salivary gland cells do not show defects in H3K9me2 and Lamin A/C distribution.**

(A–D) Graphs showing (A) H3K27ac, (B) H3K9me3 and (C) H3K9me2 mean intensity in the nucleus in diploid progenitors (in grey) and in polyploid megakaryocytes (in blue). Mean $\pm$ SEM, >120 cells analysed, three independent experiments and Lamin B1 mean intensity at the nuclear envelope (D) in diploid progenitors (in grey) and in polyploid megakaryocytes (in blue). (E) SIM representative images of diploid progenitors and polyploid megakaryocytes. H3K9me2 in cyan hot, DNA in magenta. (F, G) Graphs showing H3K9me2 (F) mean intensity and (G) coefficient of variation (defined as the ratio of the standard deviation to the mean) in progenitors (in grey) and in MKs (in blue). Mean $\pm$ SEM, >150 interphase cells analysed per condition, four independent experiments. (H) Graph showing H3K9me2 mean intensity in progenitors (in grey) and in MKs (in blue). Mean $\pm$ SEM, >65 interphase cells analysed per condition, four independent experiments. (I) Representative images of microtubules (in cyan) in MKs, DNA in magenta. (J) SIM representative images showing diploid and polyploid hepatocytes. Lamin A/C, Lamin B1 and H3K9me2 in yellow hot, red hot and cyan hot, respectively, DNA in magenta. (K, L) Graphs showing (K) the relative H3K9me2 mean intensity at the NE and (L) the H3K9me2 coefficient of variation (defined as the ratio of the standard deviation to the mean) at the NE in diploid (in grey) and in polyploid (in yellow) hepatocytes. Mean $\pm$ SEM, >90 interphase cells analysed per condition, three independent experiments. (M–O) Graphs presenting (M) Lamin A/C, (N) and Lamin B1 mean intensity at the NE and (O) the Lamin B1 vs Lamin A/C ratio at the NE in hepatocytes. Mean $\pm$ SEM, >90 interphase cells analysed per condition, three independent experiments. (P) Representative images showing *Drosophila* salivary glands at two different developmental stages (L2 and L3). Lamin A/C and Lamin B1 in yellow hot and red hot, respectively, Actin in grey, DNA in magenta. (Q, R) Graphs showing (Q) Lamin A/C and (R) Lamin B1 mean intensity at the NE in *Drosophila* salivary glands. Mean $\pm$ SEM, >150 cells analysed per condition, three independent experiments. (S) Representative images showing (left panel) polyploid hepatocytes and (right panel) *Drosophila* salivary gland cells. Microtubules in cyan. DNA in magenta. (T) Graph presenting the nuclear circularity and solidity index in progenitors (in grey) or megakaryocytes (in blue) treated with DMSO or Nocodazole. Scale bars: 10 μ m. D diploid, P polyploid, HC hepatocytes, MK megakaryocytes, NE nuclear envelope. (D) ANOVA test (one-sided). (F, G, J–O, Q, R) t-test (two-sided). ns not significant.