

Lactylation-driven FTO targets CDK2 to aggravate microvascular anomalies in diabetic retinopathy

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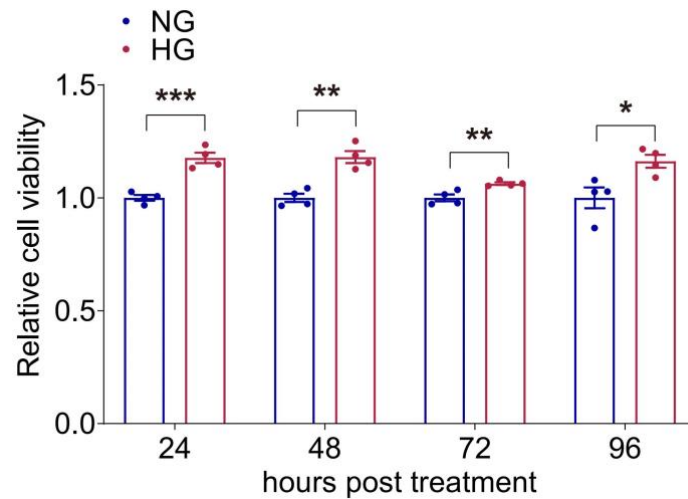
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Appendix Figure S1



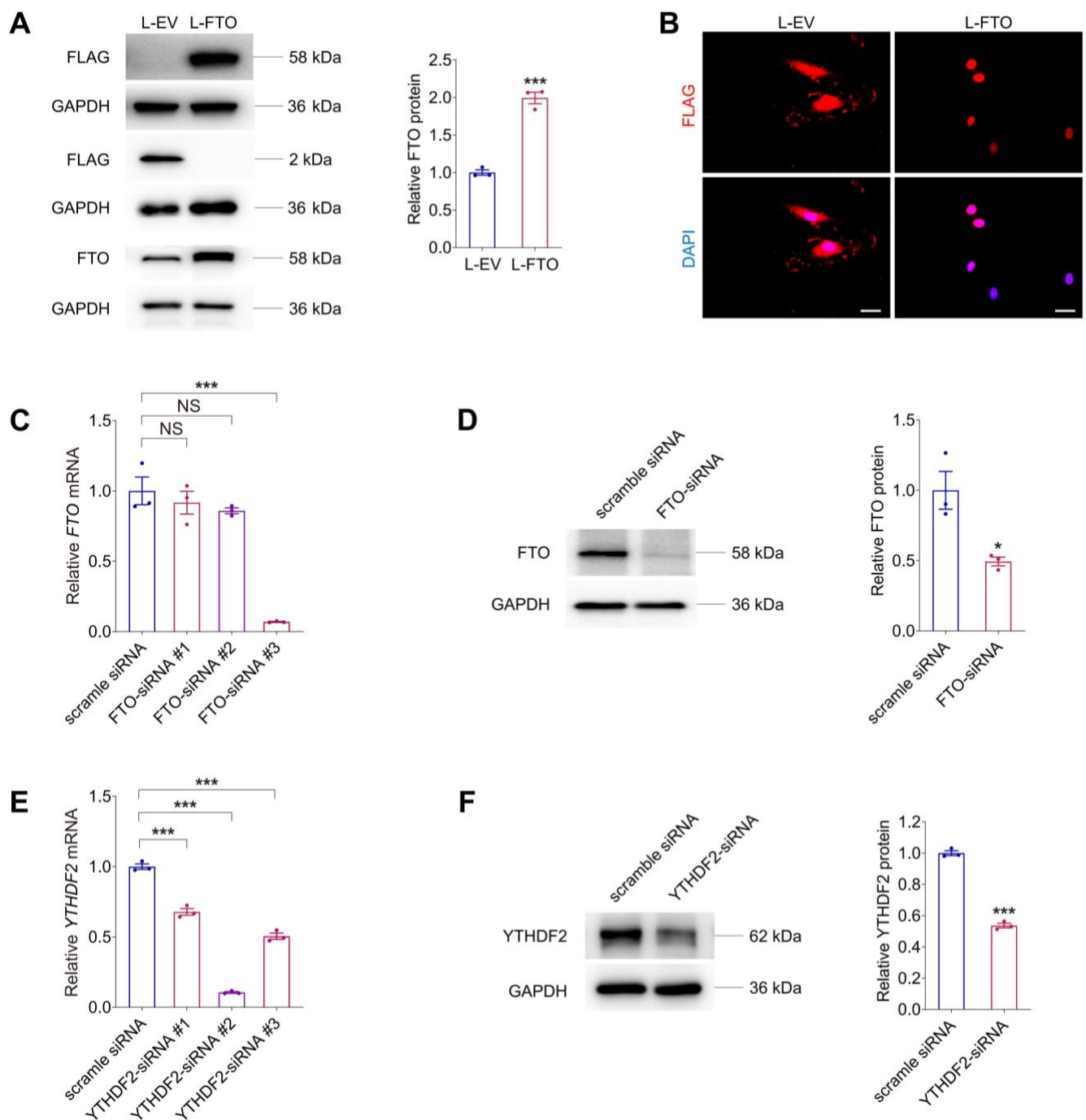
Appendix Figure S1. Cell viability of HUVECs treated with high glucose.

Cell viability of HUVECs at 24, 48, 72 and 96 hours post normal/high glucose treatment was detected by the CCK-8 assay. $n=4$ per group.

Data information: Data represent different numbers (n) of biological replicates. Data are shown as mean $\pm$ SEM. Two-tailed Student's t test is used. * $p<0.05$; ** $p<0.01$; and *** $p<0.001$.

Source data are available online for this figure.

Appendix Figure S2



Appendix Figure S2. Overexpressing efficiency of L-FTO and knocking down efficiency of siRNAs targeting FTO and YTHDF2 in HUVECs.

A. Immunoblotting of FTO and FLAG in HUVECs transduced with L-EV or L-FTO. GAPDH is used as an internal control. $n=3$ per group.

B. Immunofluorescence staining of FLAG in HUVECs transduced with L-EV or L-FTO. Scale bar: 20 μm .

C. *FTO* mRNA level detected by qPCR in HUVECs transfected with distinct siRNAs. $n=3$ per group.

D. Immunoblotting of FTO in HUVECs transfected with scramble siRNA or FTO-siRNA-3. GAPDH is used as an internal control. $n=3$ per group.

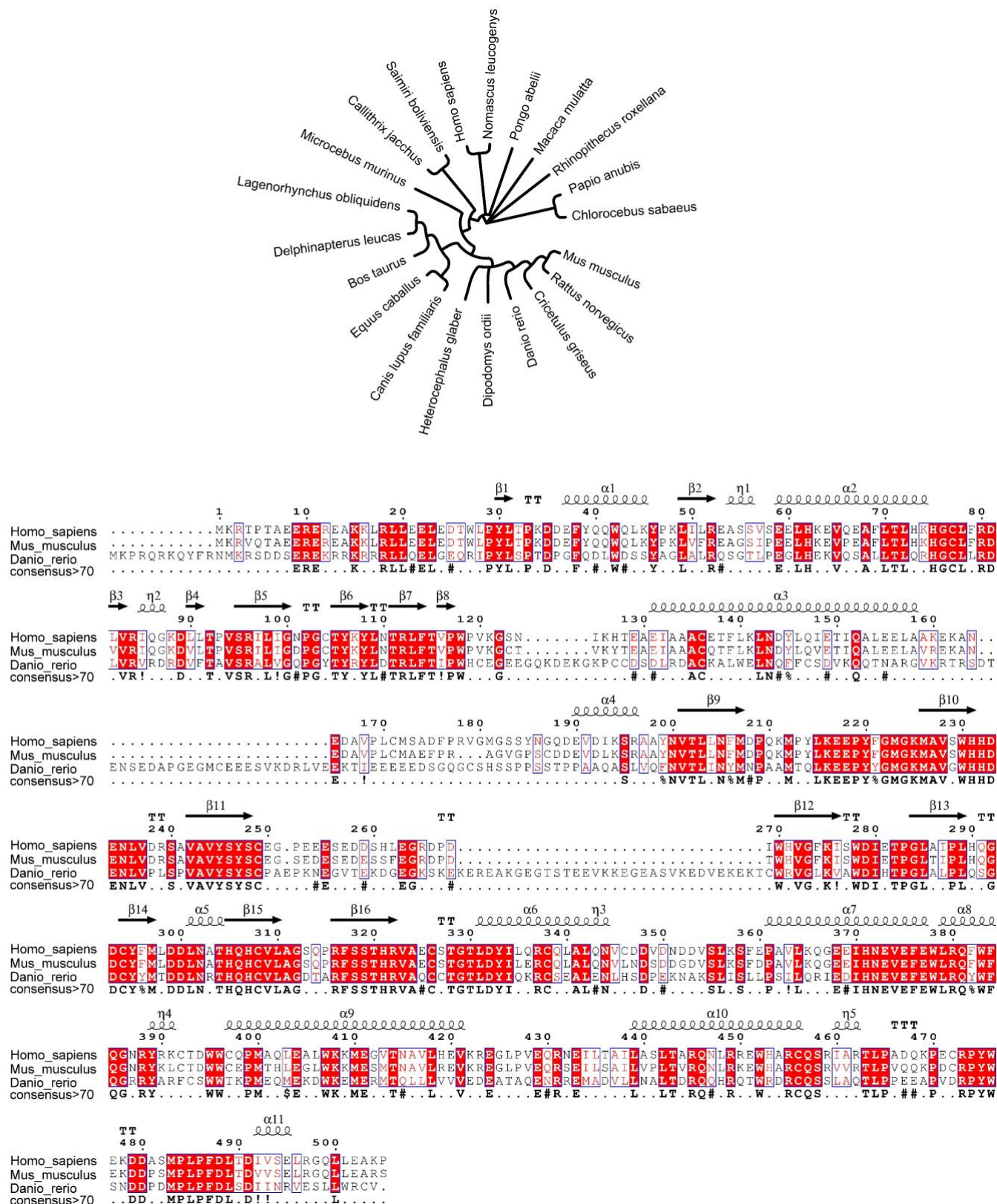
E. *YTHDF2* mRNA level detected by qPCR in HUVECs transfected with distinct siRNAs. $n=3$ per group.

F. Immunoblotting of *YTHDF2* in HUVECs transfected with scramble siRNA or *YTHDF2*-siRNA-2. GAPDH is used as an internal control. $n=3$ per group.

Data information: Data represent different numbers (n) of biological replicates. Data are shown as mean $\pm$ SEM. Two-tailed Student's t test for **A**, **D** and **F**, one-way ANOVA followed by Bonferroni's test for **C** and **E**. NS: not significant ($p>0.05$); * $p<0.05$; and *** $p<0.001$

Source data are available online for this figure.

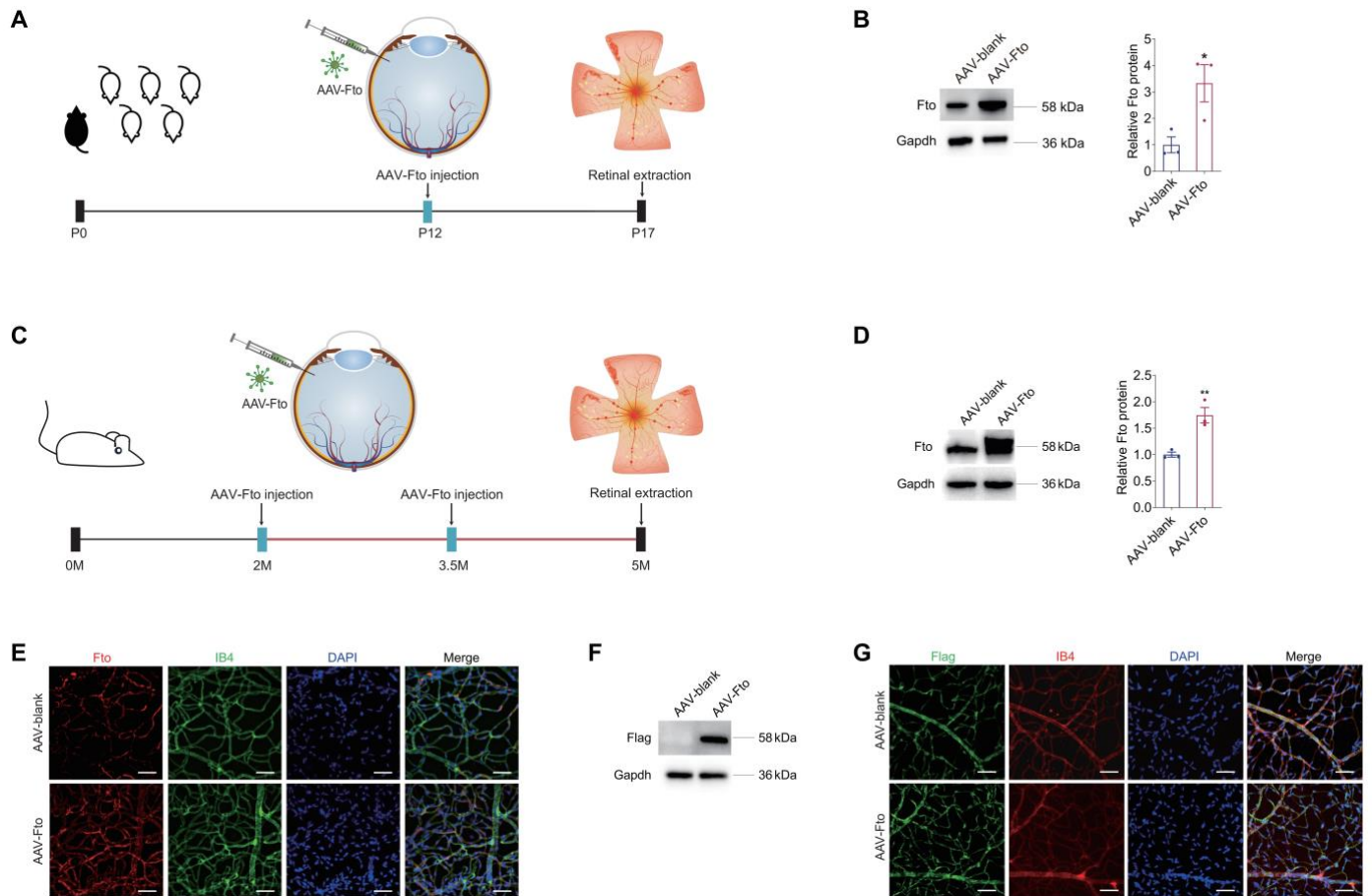
Appendix Figure S3



Appendix Figure S3. FTO is evolutionarily conserved among various species.

The phylogenetic tree is constructed with Evolview. The FTO from different species are clustered and divided into distinct subfamilies.

Appendix Figure S4



Appendix Figure S4. Overexpressing efficiency of AAV-Fto in mouse retinal vascular endothelial cells.

A. Experimental scheme for intra-vitreous AAV-Fto injection in infant mice.

B. Immunoblotting of Fto in neural retinas of P17 mice intra-vitreally injected with AAV-Fto or AAV-blank. Gapdh is used as an internal control. $n=3$ per group.

C. Experimental scheme for intra-vitreous AAV-Fto injection in adult mice.

D. Immunoblotting of Fto in neural retinas of 5-month-old mice intra-vitreally injected with AAV-Fto or AAV-blank. Gapdh is used as an internal control. $n=3$ per group.

E. Immunofluorescence staining of FTO and IB4 in trypsin digested retinal vasculatures originated from mice receiving intra-vitreous AAV-Fto or AAV-blank injection. Cell nuclei are counterstained with DAPI. Scale bar: 50 μ m.

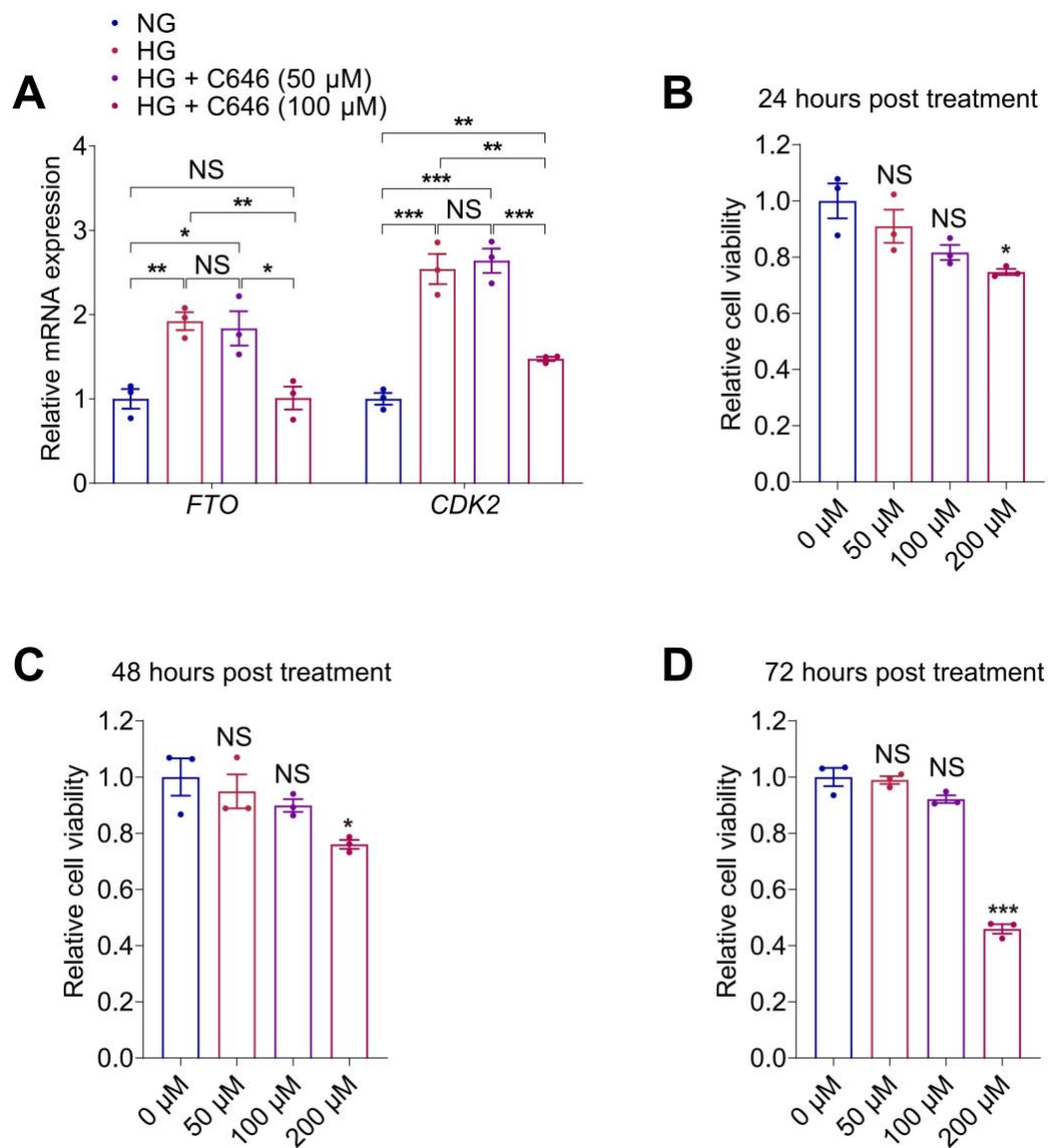
F. Immunoblotting of Flag in neural retinas of mice intra-vitreally injected with AAV-blank or AAV-Fto. Gapdh is used as an internal control.

G. Immunofluorescence staining of Flag and IB4 in trypsin digested retinal vasculatures originated from mice intra-vitreally injected with AAV-Fto or AAV-blank. Cell nuclei are counterstained with DAPI. Scale bar: 50 μ m.

Data information: Data represent different numbers (n) of biological replicates. Data are shown as mean $\pm$ SEM. Two-tailed Student's t test is used. $*p < 0.05$; $**p < 0.01$.

Source data are available online for this figure.

Appendix Figure S5



Appendix Figure S5. The concentration selection of C646 in HUVECs.

A. qPCR presents mRNA levels of *FTO* and *CDK2* in HUVECs receiving distinct treatments. $n=3$ per group.

B-D. CCK-8 assay demonstrates the cell viability of HUVECs added with C646 at distinct concentrations (0, 50, 100 and 200 μ M) for 24 (**B**), 48 (**C**) and 72 (**D**) hours. $n=3$ per group.

Data information: Data represent different numbers (n) of biological replicates. Data are shown as mean $\pm$ SEM. One-way ANOVA followed by Bonferroni's test is used. NS: not significant ($p>0.05$); * $p<0.05$; and *** $p<0.001$.

Source data are available online for this figure.

Table S1. Primers used in this study.

Gene/RNA	Forward primer (5'→3')	Reverse primer (5'→3')
<i>In vitro</i>		
<i>METTL3</i>	TTGTCTCCAACCTTCCGTAGT	CCAGATCAGAGAGGTGGTGTAG
<i>METTL14</i>	TGACATCAGAGAACTAACACCCA	GATCGAGGTGCTGCAATCTC
<i>WTAP</i>	TGAACTGAGTGCCTGGAAGT	CATCTCTTGTTCCCTTGGTTGCT
<i>ALKBH5</i>	TCAGGAAGACAAGATTAGATGCAC	TCCTTGTCCATCTCCAGGAT
<i>FTO</i>	TCATAATGAGGTCGAGTTTGAGTG	TCATGAAGCACAGCATTTGTC
<i>GAPDH</i>	ACAACCTTTGGTATCGTGGAAGG	GCCATCACGCCACAGTTTC
<i>CXCR4</i>	ACTACACCGAGGAAATGGGCT	CCACAATGCCAGTTAAGAAGA
<i>FSCN1</i>	CTGCTACTTTGACATCGAGTGG	GGGCGGTTGATGAGCTTCA
<i>APLN</i>	GTCTCCTCCATAGATTGGTCTGC	GGAATCATCCAACTACAGCCAG
<i>ESM1</i>	ACAGCAGTGAGTGCAAAAGCA	GCGGTAGCAAGTTTCTCCCC
<i>PLAUR</i>	TGTAAGACCAACGGGGATTGC	AGCCAGTCCGATAGCTCAGG
<i>ITGB1</i>	CCTACTTCTGCACGATGTGATG	CCTTTGCTACGGTTGGTTACATT
<i>CDK2</i>	GGCATTCTCTTCCCCTCAT	AGTCTGCTAGCTTGATGGCC
<i>YTHDF2</i>	CCTTACTTGAGTCCACAGGC	TGCATCTGGTAGGAAGTGGG
<i>In vivo</i>		
<i>Mettl3</i>	CTGGGCACTTGGATTTAAGGAA	TGAGAGGTGGTGTAGCAACTT
<i>Mettl14</i>	GACTGGCATCACTGCGAATGA	AGGTCCAATCCTTCCCCAGAA
<i>Wtap</i>	ATGGCACGGGATGAGTTAATTC	TTCCCTTAAACCAGTCACATCG
<i>Alkbh5</i>	TTGCCACCCAGCTATGCTTC	CAGACCGCCGGTTTTCTTCTT
<i>Fto</i>	TTCATGCTGGATGACCTCAATG	GCCAACTGACAGCGTTCTAAG
<i>Gapdh</i>	AGGTCGGTGTGAACGGATTTG	TGTAGACCATGTAGTTGAGGTCA
<i>Apln</i>	CCATGCCTTTCTAAAGCAGGATT	GCGAGACCACGCCATTAGA
<i>Esm1</i>	CTGGAGCGCCAAATATGCG	TGAGACTGTACGGTAGCAGGT
<i>Il1b</i>	GAAATGCCACCTTTTGACAGTG	TGGATGCTCTCATCAGGACAG
<i>Ccl2</i>	TTAAAAACCTGGATCGGAACCAA	GCATTAGCTTCAGATTTACGGGT
<i>Tmem119</i>	TCTTCCGGCAGTACGTGATG	CGGCGCAGACTATGAACATGA
<i>Trem2</i>	CTGGAACCGTCACCATCACTC	CGAAACTCGATGACTCCTCGG
<i>Lgals3</i>	GGAGAGGGAATGATGTTGCCT	TCCTGCTTCGTGTTACACACA
<i>Cd11c</i>	CTGGATAGCCTTTCTTCTGCTG	GCACACTGTGTCCGAAGTCA
<i>Tubb3</i>	TGTTTCAGCAGCTAAGGAAGAG	GCTCTTGGGTCATGCACTTC

Table S2. Antibodies used in this study.

Anti-protein	Host	Dilution and Application	Supplier	Catalog
<i>In vitro</i>				
m ⁶ A	Rabbit	1:1000, dot blot; 1:100, MeRIP-qPCR	Merck	ABE572
m ⁶ A	Rabbit	1 µg antibody/100 µg RNA, MeRIP-Seq	Synaptic system	SS-202 003-50
METTL3	Rabbit	1:1000, Immunoblotting	Invitrogen	PA5-28178
GAPDH	Mouse	1:2000, Immunoblotting	Proteintech	60004-1-Ig
ALKBH5	Rabbit	1:1000, Immunoblotting	Abcam	ab195377
FTO	Rabbit	1:1000, Immunoblotting, 1:50, RIP	Proteintech	27226-1-AP
FLAG	Rabbit	1:1000, Immunoblotting 1:100, Immunofluorescence	Beyotime	AF0036
cleaved caspase-3	Rabbit	1:500, Immunoblotting	Abcam	ab2302
IBA-1	Rabbit	1:150, Immunofluorescence	Abcam	ab178846
TMEM119	Rabbit	1:100, Immunofluorescence	Abcam	ab209064
CDK2	Rabbit	1:1000, Immunoblotting	Abcam	ab32147
β-actin	Rabbit	1:1000, Immunoblotting	Beyotime	AF5003
YTHDF2	Rabbit	1:50, RIP	Proteintech	24744-1-AP
H3K18la	Rabbit	1:1000, Immunoblotting; 1:100, RIP	PTM Bio	PTM-1406RM
H3	Rabbit	1:1000, Immunoblotting	Proteintech	17168-1-AP
<i>In vivo</i>				
Mettl3	Rabbit	1:1000, Immunoblotting	Invitrogen	PA5-28178
Gapdh	Mouse	1:10000, Immunoblotting	Proteintech	60004-1-Ig
Alkbh5	Rabbit	1:1000, Immunoblotting	Abcam	ab195377
Fto	Rabbit	1:1000, Immunoblotting; 1:100, Immunofluorescence	Abcam	27226-1-AP
Flag	Rabbit	1:1000, Immunoblotting 1:100, Immunofluorescence	Beyotime	AF0036
Ib4	/	1:100, Immunofluorescence	Vector Laborato ries	FL-1201-.5& DL-1207-.5
Pdgfrβ	Rabbit	1:100, Immunofluorescence	Cell Signaling Technology	3169

Ve-cadherin	Rat	1:100, Immunofluorescence	Abcam	ab282277
Zo-1	Rabbit	1:1000, Immunoblotting	Invitrogen	#61-7300
Ng2	Rabbit	1:100, Immunofluorescence	Abcam	ab275024
Iba-1	Rabbit	1:150, Immunofluorescence	Abcam	ab178846
Tubb3	Rabbit	1:100, Immunofluorescence	Abcam	ab18207
Cdk2	Rabbit	1:1000, Immunoblotting	Proteintech	10122-1-AP

Table S3. Sequences of siRNAs.

Targeted genes	Sequence (5'→3')
scramble siRNA	sense: UUCUCCGAACGUGUCACGUdTdT antisense: ACGUGACACGUUCGGAGAAdTdT
FTO-siRNA-1	sense: GGACCTGGTTAGGATCCAAdTdT antisense: UUGGAUCCUAACCAGGUCCdTdT
FTO -siRNA-2	sense: GAGCTGGCATCATGATGAAdTdT antisense: UUCAUCAUGAUGCCAGCUCdTdT
FTO -siRNA-3	sense: GCAGAATGTCTGTGACGATdTdT antisense: AUCGUCACAGACAUUCUGCdTdT
YTHDF2-siRNA-1	sense: GACCAAGAATGGCATTGCAdTdT antisense: UGCAAUGCCAUUCUUGGUCdTdT
YTHDF2-siRNA-2	sense: GCACAGAAGTTGCAAGCAAdTdT antisense: UUGCUUGCAACUUCUGUGCdTdT
YTHDF2-siRNA-3	sense: GGTAGCGGGTCCATTACTAdTdT antisense: UAGUAAUGGACCCGCUACCDdTdT
CDK2-siRNA-1	sense: GCACCAAGAUCUCAAGAAAdTdT antisense: CGUGGUUCUAGAGUUCUUUdTdT
CDK2-siRNA-2	sense: GAGUCCCUGUUCGUACUUAdTdT antisense: CUCAGGGACAAGCAUGAAUdTdT
CDK2-siRNA-3	sense: GGAUGUGACCAAGCCAGUAdTdT antisense: CCUACACUGGUUCGGUCAUdTdT