

Expanded View Figures

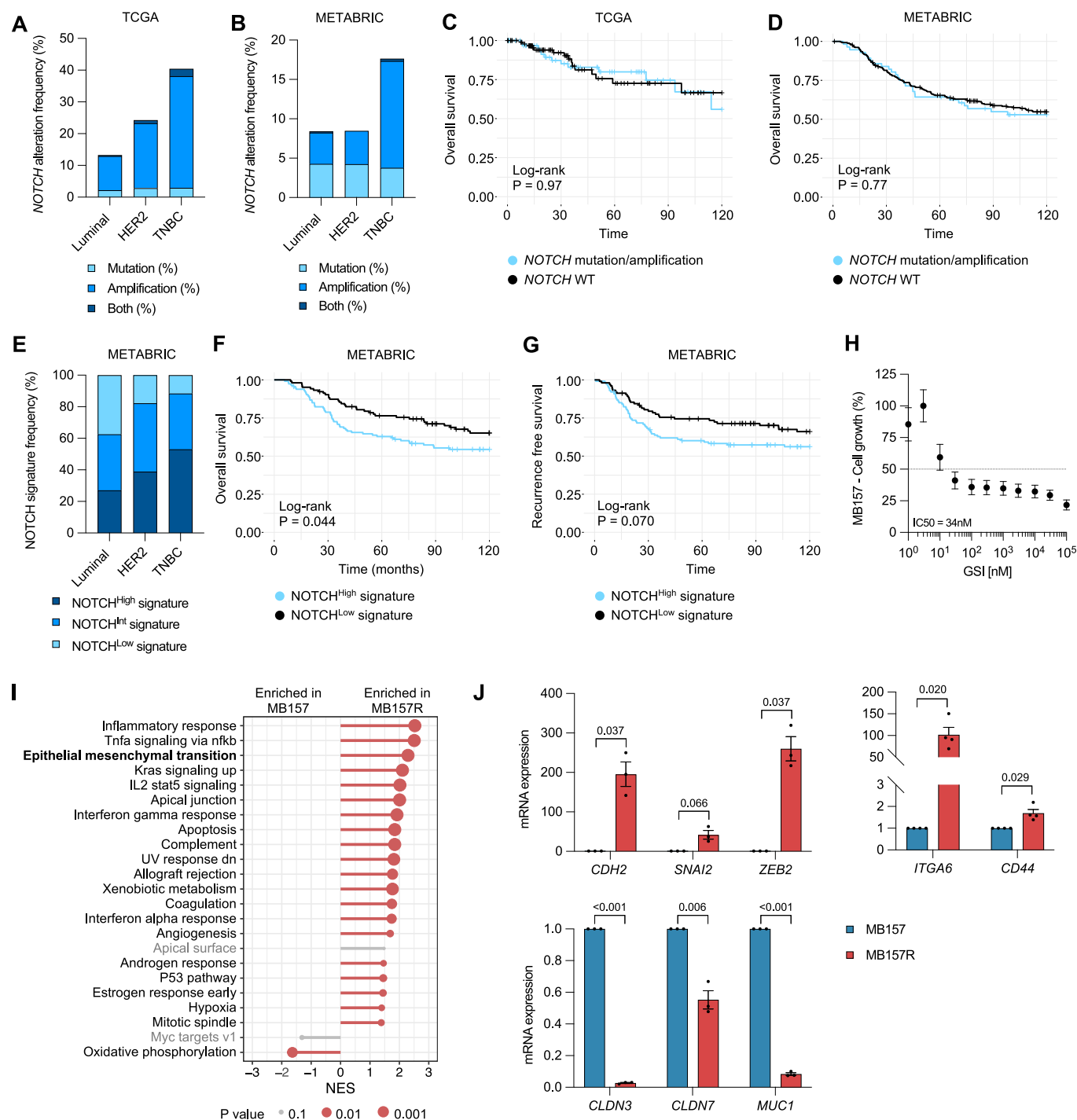


Figure EV1. Chronic exposure of NOTCH-driven TNBC cells to GSI induces drug resistance which is associated with EMT and CSC features.

(A) NOTCH mutation and amplification frequency in luminal ($n = 686$), HER2 ($n = 103$) and TNBC ($n = 168$) patients from TCGA dataset ($n = 957$). (B) NOTCH mutation and amplification frequency in luminal ($n = 1332$), HER2 ($n = 212$) and TNBC ($n = 317$) patients from METABRIC dataset ($n = 1861$). (C) OS of TNBC patients from TCGA dataset with NOTCH wild-type ($n = 100$) or altered ($n = 68$). (D) OS of TNBC patients from METABRIC dataset with NOTCH wild-type ($n = 261$) or altered ($n = 56$). (E) Frequency of NOTCH^{High}, NOTCH^{Int} or NOTCH^{Low} signature in luminal ($n = 1395$), HER2 ($n = 26$) and TNBC ($n = 347$) patients from METABRIC dataset ($n = 1968$). (F) OS of TNBC patients from METABRIC dataset with NOTCH^{High} ($n = 114$) or NOTCH^{Low} ($n = 103$) signature. (G) RFS of TNBC patients from METABRIC dataset with NOTCH^{High} ($n = 114$) or NOTCH^{Low} ($n = 103$) signature. (H) Cell growth inhibition of MB157 cells treated with GSI (1 μM) for 6 days, $n = 4$. (I) Hallmark GSEA from RNAseq analysis of MB157R compared to MB157 cells, $n = 3$. (J) Relative mRNA expression of EMT and stemness markers in MB157 and MB157R cells, $n = 3$. Data from biological replicates are represented as mean ± SEM. Log-rank test (C, D, F, G), permutation test (I) or Student *t* test (J) were used to determine *P* values.

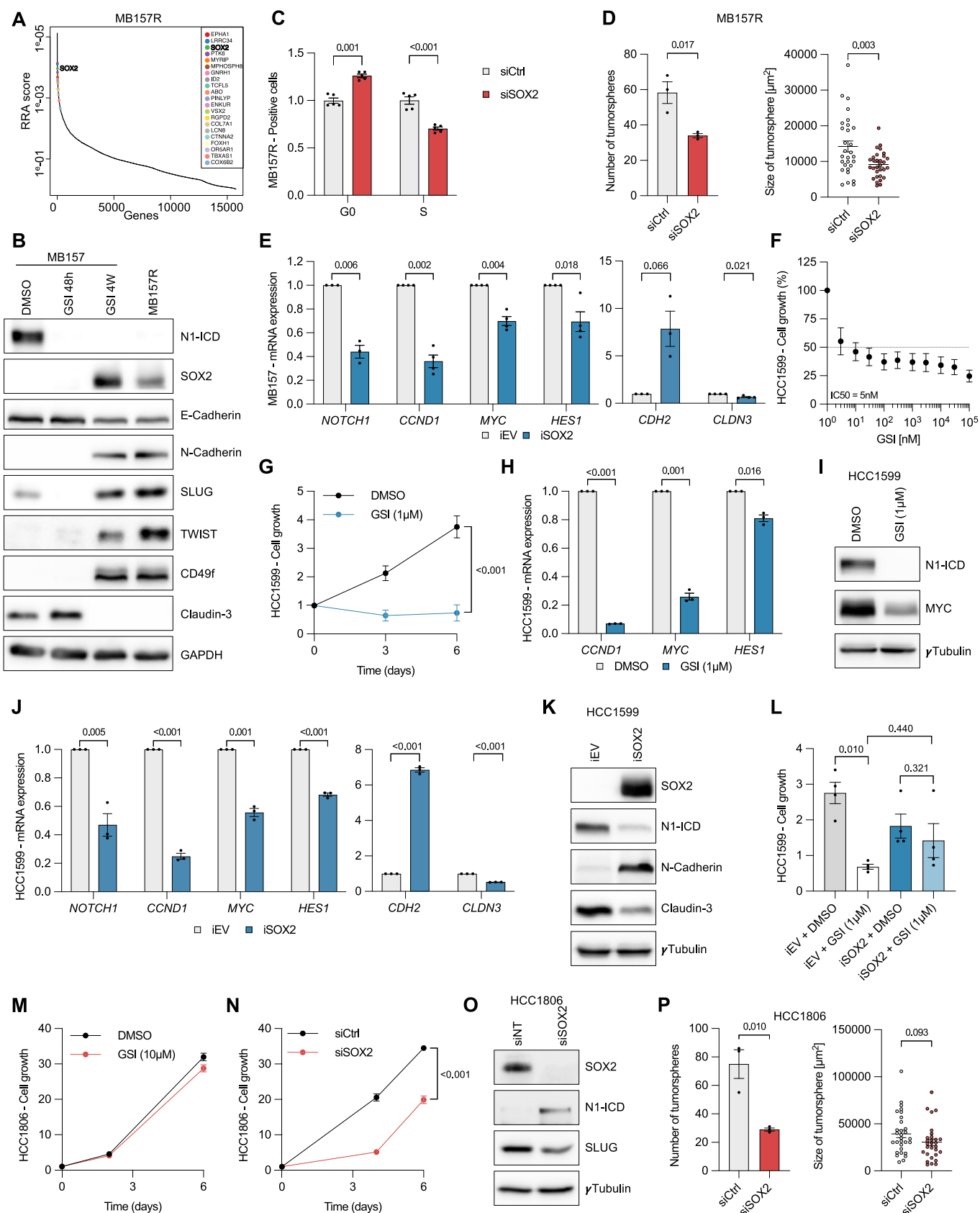


Figure EV2. SOX2 mediates resistance to GSI in TNBC inhibiting Notch signaling, promoting EMT and CSC features.

(A) Robust rank aggregation (RRA) of genes negatively selected in CRISPR-Cas9 screen of MB157R cells treated with GSI (10 μ M) for 14 days. (B) Representative immunoblotting of N1-ICD, SOX2 and EMT/Stemness markers derived from MB157 or MB157R cells as indicated. (C) Cell cycle analysis of MB157R cells 72 h after transfection with siRNA SOX2 or Ctrl, $n = 5$. (D) Number and size of tumorspheres derived from MB157R cells with siRNA SOX2 or Ctrl, $n = 3$. (E) Relative mRNA expression of *NOTCH1* and its target genes (*CCND1*, *MYC*, *HES1*) and EMT/Stemness markers in iSOX2 or iEV control MB157 cells, 72 h after DOX induction, $n = 3$. (F) Cell growth inhibition of HCC1599 treated with GSI (1 μ M) for 6 days, $n = 4$. (G) Cell proliferation assay of HCC1599 cells treated with GSI (1 μ M) for 6 days, normalized to day 0, $n = 3$. (H) Relative mRNA expression of *CCND1*, *MYC* and *HES1* in HCC1599 treated with GSI (1 μ M) for 24 h, $n = 3$. (I) Representative immunoblotting of N1-ICD and MYC derived from HCC1599 cells treated with GSI (1 μ M) for 24 h, $n = 3$. (J) Relative mRNA expression of *NOTCH1* and its target genes (*CCND1*, *MYC*, *HES1*) and EMT/Stemness markers, $n = 3$ and (K) Representative immunoblotting of N1-ICD, SOX2, N-Cadherin and Claudin-3 derived from iSOX2 or iEV control HCC1599 cells, 72 h after DOX induction, $n = 3$. (L) Cell proliferation assay of iSOX2 or iEV control HCC1599 cells treated with GSI (1 μ M) or VHC. Cell growth was assessed 6 days post treatment and normalized to day 0, $n = 4$. (M) Cell proliferation assay of HCC1806 cells treated with GSI (10 μ M) for 6 days, normalized to day 0, $n = 3$. (N) Cell proliferation assay of HCC1806 cells 72 h after transfection with siRNA SOX2 or Ctrl, normalized to day 0, $n = 3$. (O) Representative immunoblotting of N1-ICD, SOX2 and SLUG derived from HCC1806 cells 72 h after transfection with siRNA SOX2 or Ctrl, $n = 3$. (P) Number and size of tumorspheres derived from HCC1806 cells with siRNA SOX2 or Ctrl, $n = 3$. Data from biological replicates are represented as mean $\pm$ SEM. Student *t* test (C-E, H, J, P) two-way ANOVA (G, N) or one-way ANOVA (L) were used to determine *P* value.

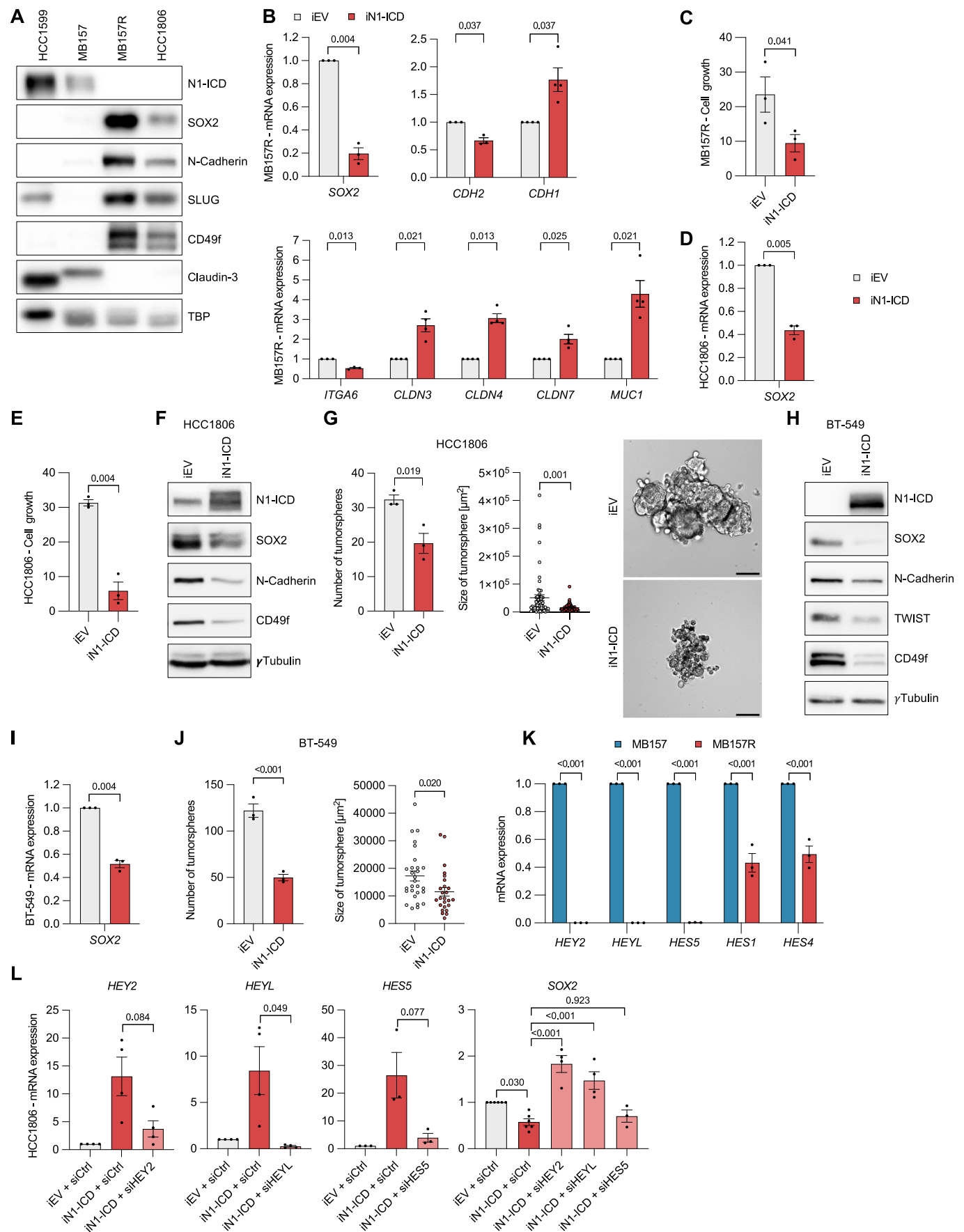
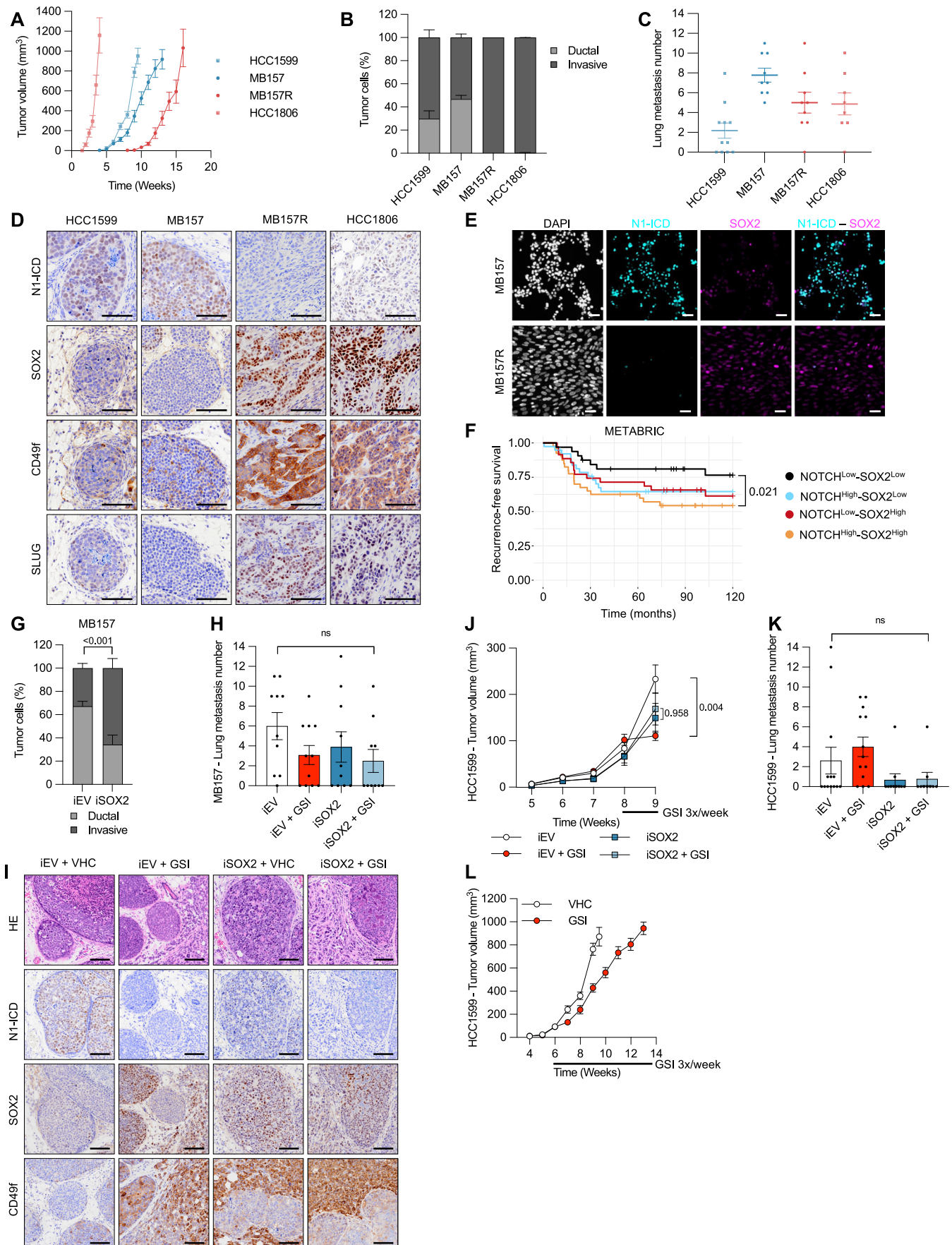


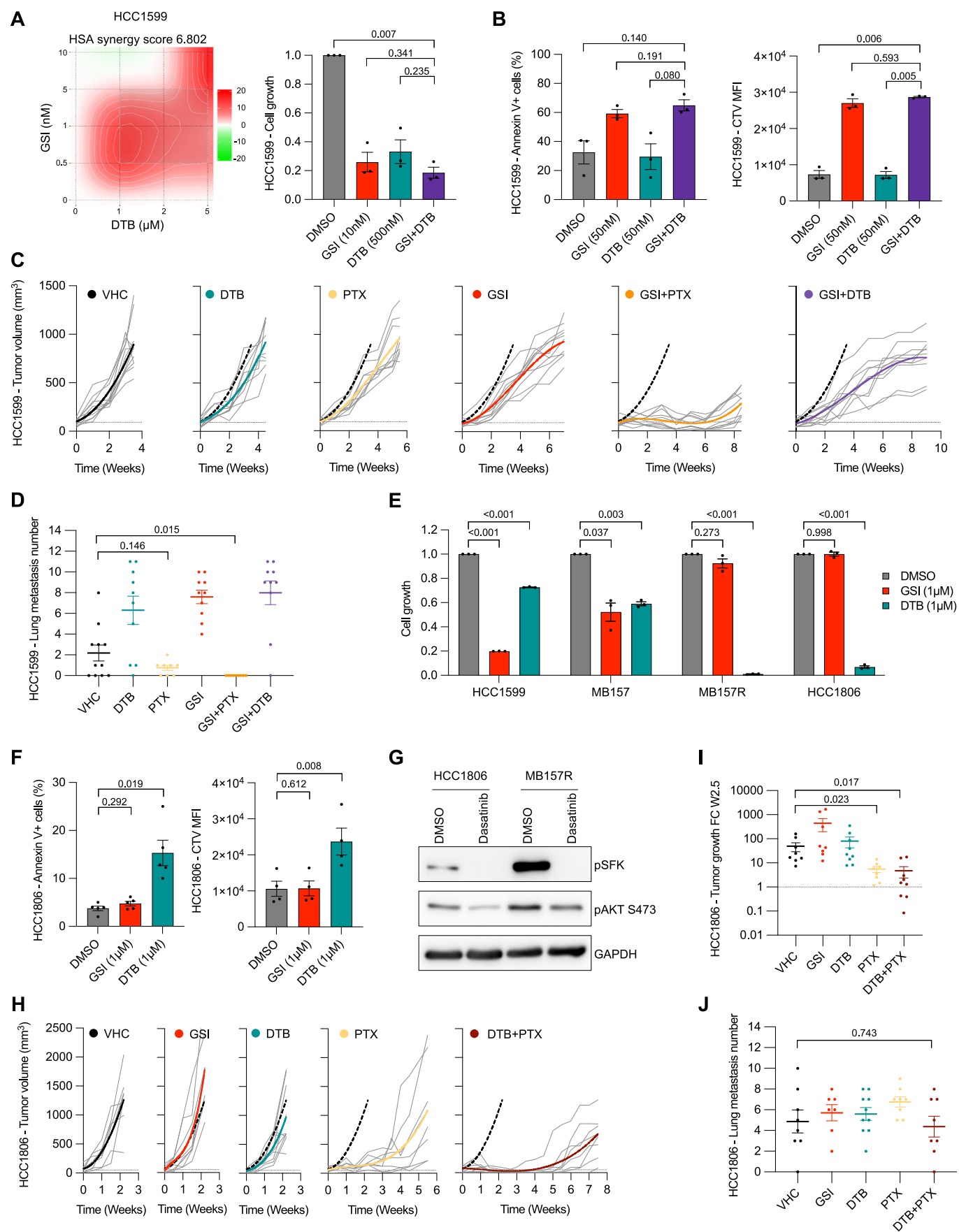
Figure EV3. Reciprocal SOX2 inhibition is mediated through Notch downstream transcriptional repressors of the HEY family.

(A) Representative immunoblotting of N1-ICD, SOX2 and EMT/Stemness markers derived from HCC1599, MB157, MB157R and HCC1806 cells. (B) Relative mRNA expression of SOX2 and EMT/Stemness markers in iN1-ICD or iEV control MB157R cells, 72 h after DOX induction $n = 3-4$. (C) Cell proliferation assay of iN1-ICD or iEV control MB157R cells. Cell growth was assessed 6 days after DOX induction and normalized to day 0, $n = 4$. (D) Relative mRNA expression of SOX2 in iN1-ICD or iEV control HCC1806 cells, 72 h after DOX induction, $n = 3$. (E) Cell proliferation assay of iN1-ICD or iEV control HCC1806 cells. Cell growth was assessed 6 days after DOX induction and normalized to day 0, $n = 3$. (F) Representative immunoblotting of N1-ICD, SOX2 and EMT/Stemness markers derived from iN1-ICD or iEV control HCC1806 cells, 72 h after DOX induction. (G) Number and size of tumorspheres with representative pictures derived from iN1-ICD compared to iEV control HCC1806 cells after 14 days, $n = 3$. Scale = 100 μm . (H) Representative immunoblotting of N1-ICD, SOX2 and EMT/Stemness markers and (I) Relative mRNA expression of SOX2 in iN1-ICD or iEV control BT-549 cells, 72 h after DOX induction $n = 3$. (J) Number and size of tumorspheres derived from iN1-ICD compared to iEV control BT-549 cells after 14 days, $n = 3$. (K) Relative mRNA expression of *HEY/HES* family genes in MB157 and MB157R cells, $n = 3$. (L) Relative mRNA expression of *HEY2*, *HEYL*, *HES5* and *SOX2* in iN1-ICD or iEV control HCC1806 cells 72 h after transfection with siRNA *HEY2*, *HEYL*, *HES5* or Ctrl $n = 3-4$. Data from biological replicates are represented as mean $\pm$ SEM. Student *t* test (B-E, G, I-L) or one-way ANOVA (L) were used to determine *P* value.



◀ **Figure EV4. Escape of GSI-mediated in vivo tumor growth control due to TNBC tumor cell plasticity.**

(A) Tumor growth of HCC1599, MB157, MB157R and HCC1806 MIND xenografts ($n = 9-11$). (B) Quantification of tumor cells in invasive or ductal areas from HE coloration in HCC1599, MB157, MB157R and HCC1806 MIND xenografts at endpoint ($\sim 1000 \text{ mm}^3$) ($n = 3$). (C) Lung metastasis number in HCC1599, MB157, MB157R and HCC1806 MIND xenografts at endpoint, $n = 9-11$. (D) Representative pictures of N1-ICD, SOX2, CD49f and SLUG IHC stainings for HCC1599, MB157, MB157R and HCC1806 MIND xenograft tumors at endpoint, scale = 100 μm . (E) Representative images of co-immunofluorescence staining of N1-ICD – SOX2 for MB157 and MB157R cell lines in vitro, scale = 50 μm . (F) RFS of TNBC patients from METABRIC dataset with NOTCH^{High/Low} signature and SOX2^{High/Low} expression. TNBC patients are divided in 4 groups: NOTCH^{High}/SOX2^{High} ($n = 40$), NOTCH^{High}/SOX2^{Low} ($n = 38$), NOTCH^{Low}/SOX2^{High} ($n = 35$) and NOTCH^{Low}/SOX2^{Low} ($n = 32$). (G) Quantification of tumor cells in invasive or ductal areas from HE coloration in MB157-iSOX2 xenografts, $n = 3$. (H) Lung metastasis number in SOX2-expressing or EV control MB157 MIND xenografts treated with GSI (8 mg/kg, 3 $\times$ /week) or VHC for 1 week, $n = 10-11$. (I) Representative pictures of HE coloration, N1-ICD, SOX2 and CD49f IHC stainings for iSOX2 or iEV control MB157 MIND xenografts treated with GSI (8 mg/kg, 3 $\times$ /week) or VHC for 1 week, scale = 100 μm . (J) Tumor growth of iSOX2 or iEV control HCC1599 MIND xenografts treated with GSI (8 mg/kg, 3 $\times$ /week) or VHC for 1 week, $n = 9-13$. (K) Lung metastasis number in iSOX2 or iEV control HCC1599 MIND xenografts treated with GSI (8 mg/kg, 3 $\times$ /week) or VHC for 1 week, $n = 9-13$. (L) Tumor growth of HCC1599 xenografts treated with GSI (8 mg/kg) or VHC for 8 weeks, $n = 9-13$. Data from biological replicates are represented as mean $\pm$ SEM. Log-rank test (F), Cochran-Mantel-Haenszel test (G), one-way ANOVA (H, K) or two-way ANOVA (J), were used to determine P value.



◀ **Figure EV5. Combination therapies to treat GSI-sensitive and -resistant TNBC xenografts.**

(A) HAS-synergy heatmap derived from HCC1599 cells treated with a concentration matrix of GSI-DTB for 6 days and cell growth inhibition of HCC1599 cells treated with GSI (10 nM) and/or DTB (500 nM) for 6 days, $n = 3$. (B) Proportion of Annexin V+ cells ($n = 4$) and Cell Trace Violet mean fluorescence intensity (MFI) ($n = 3$) in HCC1599 cells treated with GSI (50 nM) and/or DTB (50 nM) for 6 days, by flow cytometry analysis. (C) Tumor growth of HCC1599 xenografts treated with GSI (8 mg/kg, 3×/week), DTB (15 mg/kg, 5×/week), PTX (15 mg/kg, 1×/week) or VHC, $n = 9-11$. (D) Lung metastasis number in HCC1599 xenografts treated with GSI (8 mg/kg, 3×/week), DTB (15 mg/kg, 5×/week), PTX (15 mg/kg, 1×/week) or VHC, $n = 8-11$. (E) Cell growth inhibition of HCC1599, MB157, MB157R and HCC1806 cells treated with GSI and/or DTB for 6 days, $n = 3$. (F) Proportion of Annexin V+ cells ($n = 5$) and Cell Trace Violet MFI ($n = 4$) in HCC1806 cells treated with GSI (1 μ M) or DTB (1 μ M) for 3 days, by flow cytometry analysis. (G) Representative immunoblotting of pSFK and pAKT S473 derived from MB157R and HCC1806 cells treated with Dasatinib 1 μ M for 18 h. (H) Tumor growth of HCC1806 MIND xenografts treated with GSI (8 mg/kg, 3×/week), DTB (15 mg/kg, 5×/week), PTX (15 mg/kg, 1×/week) or VHC, $n = 8-10$. (I) Tumor growth fold change (2.5 weeks after treatment) of HCC1806 MIND xenografts treated with GSI (8 mg/kg, 3×/week), DTB (15 mg/kg, 5×/week), PTX (15 mg/kg, 1×/week) or VHC, $n = 6-9$. (J) Lung metastasis number in HCC1806 MIND xenografts treated with GSI (8 mg/kg, 3×/week), DTB (15 mg/kg, 5×/week), PTX (15 mg/kg, 1×/week) or VHC, $n = 8-10$. Data from biological replicates are represented as mean $\pm$ SEM. One-way ANOVA (A, B, D-F, I, J) was used to determine P value.