

Differential utilization of binding loop flexibility in T cell receptor ligand selection and cross-reactivity

Cory M. Ayres^{a,b}, Daniel R. Scott^{a,1}, Steven A. Corcelli^a, and Brian M. Baker^{a,b,*}

^aDepartment of Chemistry & Biochemistry, University of Notre Dame, Notre Dame, IN
46556 USA

^bHarper Cancer Research Institute, University of Notre Dame, Notre Dame, IN 46556 USA

*Corresponding author:

Phone: (574) 631-9810

Fax: (574) 631-6652

Email: brian-baker@nd.edu

¹Current address: National Institutes of Standards and Technology, Neutron Condensed
Matter Group, Gaithersburg, MD 20899 USA

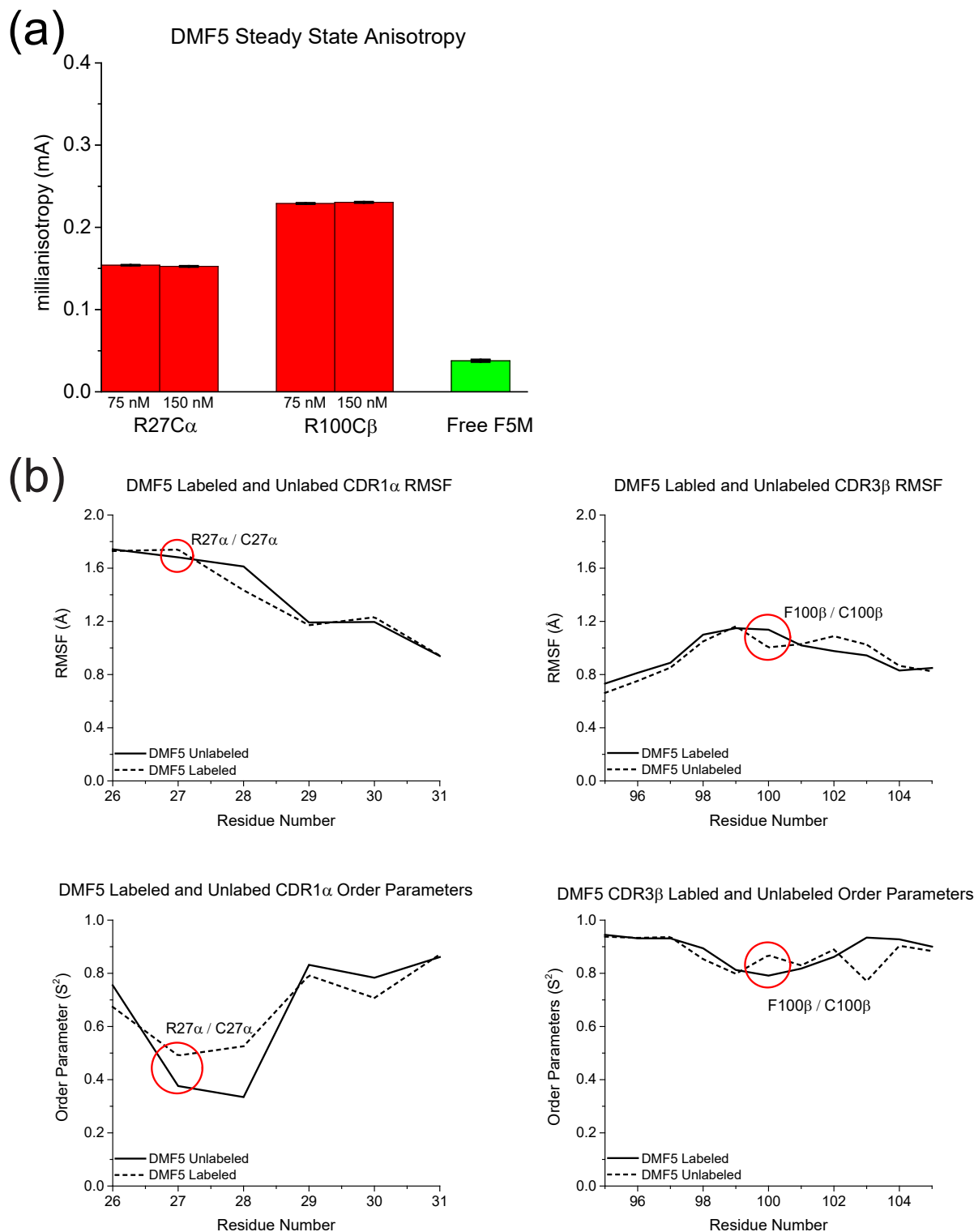


Fig. S1. Fluorescence anisotropy measurements validate the simulation methodology for DMF5. **A)** Steady state anisotropy measurements for DMF5 fluorescently labeled at position 27 in CDR1 α or position 100 in CDR3 β . Consistent with the simulation results, position 27 α is more mobile on the nanosecond timescale (lower anisotropy) than position 100 β . Measurements for two protein concentrations are shown. Error bars reflect the standard deviation of 50 measurements. Green shows the anisotropy of free fluorescein. **B)** Incorporation of a cysteine mutation and fluorescent label does not significantly perturb the simulation results for position 27 α and neighboring residues or position 100 β and neighboring residues. The two panels show α carbon RMSF values and the bottom two show $C\alpha$ - $C\beta$ order parameters, with solid lines the results from the simulations of unlabeled protein and the dashed lines the simulations of labeled protein. Values for positions 27 α or 100 β are circled.