

Structural basis for PPAR partial or full activation revealed by a novel ligand binding mode

Davide Capelli^{1,5}, Carmen Cerchia^{2,5}, Roberta Montanari¹, Fulvio Loiodice³, Paolo Tortorella³, Antonio Laghezza³, Laura Cervoni⁴, Giorgio Pochetti^{1,*} and Antonio Lavecchia^{2,*}

¹Istituto di Cristallografia, Consiglio Nazionale delle Ricerche, Via Salaria Km. 29,300, 00015 Monterotondo Stazione, Roma, Italy

²Dipartimento di Farmacia, Università degli Studi di Napoli, Via Montesano 49, 80131 Napoli, Italy

³Dipartimento di Farmacia-Scienze del Farmaco, Università degli Studi di Bari “Aldo Moro”, Via E.Orabona 4, 70126 Bari, Italy

⁴Dipartimento di Scienze Biochimiche “A. Rossi Fanelli”, Università di Roma “La Sapienza”, Piazzale A. Moro 5, 00185 Roma, Italy

⁵Co-first author

*Correspondence: giorgio.pochetti@ic.cnr.it (G.P.), antonio.lavecchia@unina.it (A.L.)

Supporting Information

Table S1. List of crystal structures of PPAR γ in complex with partial agonists used for this study.

PDB code	Ligand code	Ligand name	Resolution (Å)	Reference
2FVJ	RO0	1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-4- {[4-(2-methoxyphenyl)piperidin-1-yl]methyl} isoquinoline	1.99	¹
2G0G	SP0	3-fluoro-n-[1-(4-fluorophenyl)-3-(2-thienyl)-1h-pyrazol-5-yl]benzenesulfonamide	2.54	²
2G0H	SP3	N-[1-(4-fluorophenyl)-3-(2-thienyl)-1h-pyrazol-5-yl]-3,5-bis(trifluoromethyl)benzenesulfonamide	2.3	²
2I4P	DRH	(2S)-2-(4-{2-[1,3-benzoxazol-2-yl(heptyl)amino]ethyl}phenoxy)-2-methylbutanoic acid	2.1	³
2I4Z	DRH	(2S)-2-(4-{2-[1,3-benzoxazol-2-yl(heptyl)amino]ethyl}phenoxy)-2-methylbutanoic acid	2.25	³
2P4Y	C03	(2R)-2-(4-chloro-3-{[3-(6-methoxy-1,2-benzisoxazol-3-yl)-2-methyl-6-(trifluoromethoxy)-1h-indol-1-yl]methyl}phenoxy)propanoic acid	2.25	⁴
2Q5P	241	(2S)-2-(3-{[1-(4-methoxybenzoyl)-2-methyl-5-	2.3	⁵

		(trifluoromethoxy)-1h-indol-3-yl)methyl}phenoxy)propanoic acid		
2Q5S	NZA	5-chloro-1-(4-chlorobenzyl)-3-(phenylthio)-1H-indole-2-carboxylic acid	2.05	5
2Q61	SF1	1-benzyl-5-chloro-3-(phenylthio)-1H-indole-2-carboxylic acid	2.2	5
2Q6R	SF2	5-chloro-1-(3-methoxybenzyl)-3-(phenylthio)-1H-indole-2-carboxylic acid	2.41	5
2Q6S	PLB	2-[(2,4-dichlorobenzoyl)amino]-5-(pyrimidin-2-yloxy)benzoic acid	2.4	5
2YFE	YFE	Amorfrutin 1	2	6
3B1M	KRC	(9AS)-8-acetyl-N-[(2-ethylnaphthalen-1-yl)methyl]-1,7-dihydroxy-3-methoxy-9Amethyl- 9-oxo-9,9A -dihydrodibenzo[B,D]furan-4-carboxamide	1.6	7
3CDP	YRG	(2S)-2-(4-chlorophenoxy)-3-phenylpropanoic acid	2.8	8
3D6D	LRG	(2S)-2-(biphenyl-4-yloxy)-3-phenylpropanoic acid	2.4	9
3FUR	Z12	2,4-dichloro-N-[3,5-dichloro-4-(quinolin-3-yloxy)phenyl]benzenesulfonamide	2.3	10
3H0A	D30	[(4-{[2-(pent-2-yn-1-yloxy)-4-{[4-(trifluoromethyl)phenoxy]methyl}phenyl]sulfanyl}-5,6,7,8-tetrahydronaphthalen-1-yl)oxy]acetic acid	2.1	11
3K8S	Z27	2-chloro-N-{3-chloro-4-[(5-chloro-1,3-benzothiazol-2-yl)sulfanyl]phenyl}-4-(trifluoromethyl)benzenesulfonamide	2.55	12
3LMP	CEK	(9AS)-8-acetyl-1,7-dihydroxy-3-methoxy-9A-methyl-N-(1-naphthylmethyl)-9-oxo-9,9A-dihydrodibenzo[B,D]furan-4-carboxamide	1.9	13
3OSI	XDH	4,4'-propane-2,2-diylbis(2,6-dichlorophenol)	2.7	14
3OSW	XDI	4,4'-propane-2,2-diylbis(2,6-dibromophenol)	2.55	14
3PBA	ZXG	2,6-dibromo-4-[2-(3,5-dibromo-4-hydroxyphenyl)propan-2-yl]phenyl hydrogen sulfate	2.3	15
4A4V	YFD	Amorfrutin 2	2	16
4A4W	YFB	Amorfrutin B	2	16
4PRG	072	(±)(2S,5S)-3-(4-(4-carboxyphenyl)butyl)-2-heptyl-4-	2.9	17

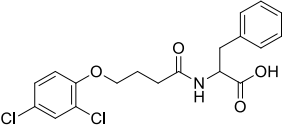
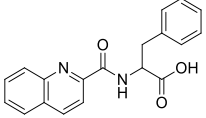
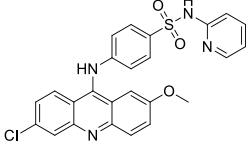
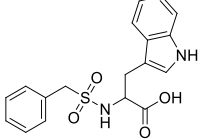
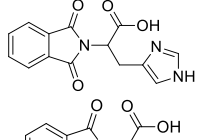
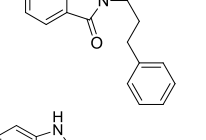
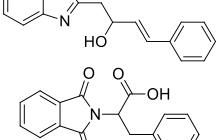
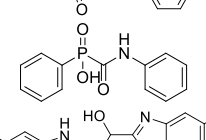
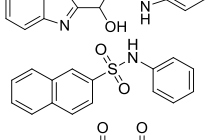
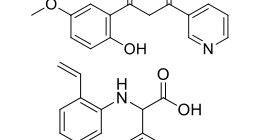
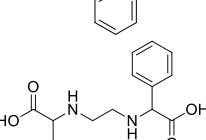
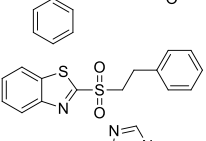
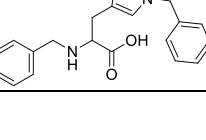
References

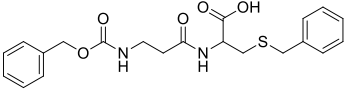
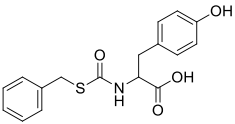
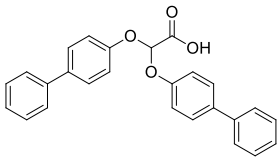
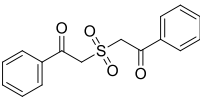
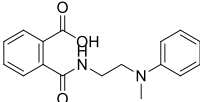
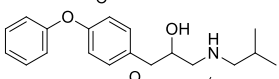
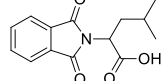
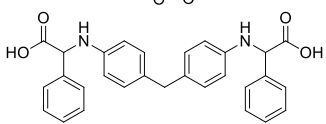
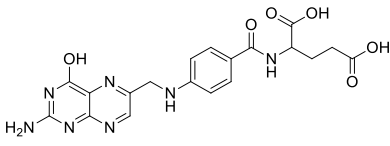
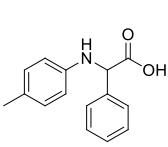
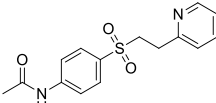
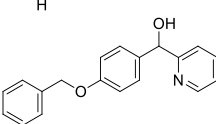
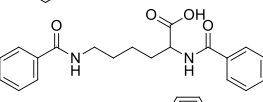
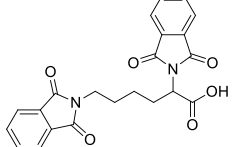
1. Burgermeister, E.; Schnobelen, A.; Flament, A.; Benz, J.; Stihle, M.; Gsell, B.; Rufer, A.; Ruf, A.; Kuhn, B.; Marki, H. P.; Mizrahi, J.; Sebokova, E.; Niesor, E.; Meyer, M. A novel partial agonist of peroxisome proliferator-activated receptor-gamma (PPARgamma) recruits PPARgamma-coactivator-1alpha, prevents triglyceride accumulation, and potentiates insulin signaling in vitro. *Mol Endocrinol* **2006**, 20, 809-30.
2. Lu, I. L.; Huang, C. F.; Peng, Y. H.; Lin, Y. T.; Hsieh, H. P.; Chen, C. T.; Lien, T. W.; Lee, H. J.; Mahindroo, N.; Prakash, E.; Yueh, A.; Chen, H. Y.; Goparaju, C. M.; Chen, X.; Liao, C. C.; Chao, Y. S.; Hsu, J. T.; Wu, S. Y. Structure-based drug design of a novel family of PPARgamma partial agonists: virtual screening, X-ray crystallography, and in vitro/in vivo biological activities. *J Med Chem* **2006**, 49, 2703-12.
3. Pochetti, G.; Godio, C.; Mitro, N.; Caruso, D.; Galmozzi, A.; Scurati, S.; Loiodice, F.; Fracchiolla, G.; Tortorella, P.; Laghezza, A.; Lavecchia, A.; Novellino, E.; Mazza, F.; Crestani, M. Insights into the Mechanism of Partial Agonism: CRYSTAL STRUCTURES OF THE PEROXISOME PROLIFERATOR-ACTIVATED RECEPTOR γ LIGAND-BINDING DOMAIN IN THE COMPLEX WITH TWO ENANTIOMERIC LIGANDS. *Journal of Biological Chemistry* **2007**, 282, 17314-17324.
4. Einstein, M.; Akiyama, T. E.; Castriota, G. A.; Wang, C. F.; McKeever, B.; Mosley, R. T.; Becker, J. W.; Moller, D. E.; Meinke, P. T.; Wood, H. B.; Berger, J. P. The Differential Interactions of Peroxisome Proliferator-Activated Receptor γ Ligands with Tyr473 Is a Physical Basis for Their Unique Biological Activities. *Molecular Pharmacology* **2008**, 73, 62-74.
5. Bruning, J. B.; Chalmers, M. J.; Prasad, S.; Busby, S. A.; Kamenecka, T. M.; He, Y.; Nettles, K. W.; Griffin, P. R. Partial Agonists Activate PPAR γ Using a Helix 12 Independent Mechanism. *Structure* **2007**, 15, 1258-1271.
6. Weidner, C.; de Groot, J. C.; Prasad, A.; Freiwald, A.; Quedenau, C.; Kliem, M.; Witzke, A.; Kodelja, V.; Han, C. T.; Giegold, S.; Baumann, M.; Klebl, B.; Siems, K.; Muller-Kuhrt, L.; Schurmann, A.; Schuler, R.; Pfeiffer, A. F.; Schroeder, F. C.; Bussow, K.; Sauer, S. Amorfrutins are potent antidiabetic dietary natural products. *Proc Natl Acad Sci U S A* **2012**, 109, 7257-62.
7. Wakabayashi, K.; Hayashi, S.; Matsui, Y.; Matsumoto, T.; Furukawa, A.; Kuroha, M.; Tanaka, N.; Inaba, T.; Kanda, S.; Tanaka, J.; Okuyama, R.; Wakimoto, S.; Ogata, T.; Araki, K.; Ohsumi, J. Pharmacology and in vitro profiling of a novel peroxisome proliferator-activated receptor gamma ligand, Cerco-A. *Biol Pharm Bull* **2011**, 34, 1094-104.
8. Fracchiolla, G.; Laghezza, A.; Piemontese, L.; Parente, M.; Lavecchia, A.; Pochetti, G.; Montanari, R.; Di Giovanni, C.; Carbonara, G.; Tortorella, P.; Novellino, E.; Loiodice, F. Synthesis, biological evaluation and molecular investigation of fluorinated peroxisome proliferator-activated receptors alpha/gamma dual agonists. *Bioorganic & medicinal chemistry* **2012**, 20, 2141-51.

9. Montanari, R.; Saccoccia, F.; Scotti, E.; Crestani, M.; Godio, C.; Gilardi, F.; Loiodice, F.; Fracchiolla, G.; Laghezza, A.; Tortorella, P.; Lavecchia, A.; Novellino, E.; Mazza, F.; Aschi, M.; Pochetti, G. Crystal Structure of the Peroxisome Proliferator-Activated Receptor γ (PPAR γ) Ligand Binding Domain Complexed with a Novel Partial Agonist: A New Region of the Hydrophobic Pocket Could Be Exploited for Drug Design. *Journal of Medicinal Chemistry* **2008**, 51, 7768-7776.
10. Motani, A.; Wang, Z.; Weiszmann, J.; McGee, L. R.; Lee, G.; Liu, Q.; Staunton, J.; Fang, Z.; Fuentes, H.; Lindstrom, M.; Liu, J.; Biermann, D. H.; Jaen, J.; Walker, N. P.; Learned, R. M.; Chen, J. L.; Li, Y. INT131: a selective modulator of PPAR gamma. *J Mol Biol* **2009**, 386, 1301-11.
11. Connors, R. V.; Wang, Z.; Harrison, M.; Zhang, A.; Wanska, M.; Hiscock, S.; Fox, B.; Dore, M.; Labelle, M.; Sudom, A.; Johnstone, S.; Liu, J.; Walker, N. P.; Chai, A.; Siegler, K.; Li, Y.; Coward, P. Identification of a PPARdelta agonist with partial agonistic activity on PPARgamma. *Bioorg Med Chem Lett* **2009**, 19, 3550-4.
12. Li, Y.; Wang, Z.; Furukawa, N.; Escaron, P.; Weiszmann, J.; Lee, G.; Lindstrom, M.; Liu, J.; Liu, X.; Xu, H.; Plotnikova, O.; Prasad, V.; Walker, N.; Learned, R. M.; Chen, J. L. T2384, a novel antidiabetic agent with unique peroxisome proliferator-activated receptor gamma binding properties. *J Biol Chem* **2008**, 283, 9168-76.
13. Furukawa, A.; Arita, T.; Satoh, S.; Wakabayashi, K.; Hayashi, S.; Matsui, Y.; Araki, K.; Kuroha, M.; Ohsumi, J. Discovery of a novel selective PPARgamma modulator from (-)-Cercosporamide derivatives. *Bioorg Med Chem Lett* **2010**, 20, 2095-8.
14. Riu, A.; Grimaldi, M.; le Maire, A.; Bey, G.; Phillips, K.; Boulahtouf, A.; Perdu, E.; Zalko, D.; Bourguet, W.; Balaguer, P. Peroxisome Proliferator-Activated Receptor γ Is a Target for Halogenated Analogs of Bisphenol A. *Environ Health Persp* **2011**, 119, 1227-1232.
15. Riu, A.; le Maire, A.; Grimaldi, M.; Audebert, M.; Hillenweck, A.; Bourguet, W.; Balaguer, P.; Zalko, D. Characterization of Novel Ligands of ER α , ER β , and PPAR γ : The Case of Halogenated Bisphenol A and Their Conjugated Metabolites. *Toxicological Sciences* **2011**, 122, 372-382.
16. de Groot, J. C.; Weidner, C.; Krausze, J.; Kawamoto, K.; Schroeder, F. C.; Sauer, S.; Bussow, K. Structural characterization of amorfrutins bound to the peroxisome proliferator-activated receptor gamma. *J Med Chem* **2013**, 56, 1535-43.
17. Oberfield, J. L.; Collins, J. L.; Holmes, C. P.; Goreham, D. M.; Cooper, J. P.; Cobb, J. E.; Lenhard, J. M.; Hull-Ryde, E. A.; Mohr, C. P.; Blanchard, S. G.; Parks, D. J.; Moore, L. B.; Lehmann, J. M.; Plunket, K.; Miller, A. B.; Milburn, M. V.; Kliewer, S. A.; Willson, T. M. A peroxisome proliferator-activated receptor gamma ligand inhibits adipocyte differentiation. *Proc Natl Acad Sci U S A* **1999**, 96, 6102-6.

Table S2. Compounds identified as hits against PPAR γ from the SBVS and their activity in cell-based transactivation assays.

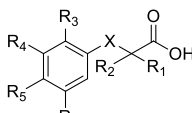
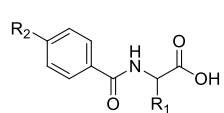
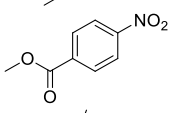
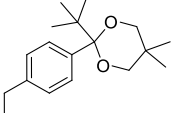
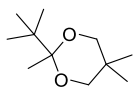
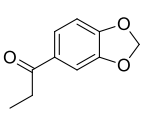
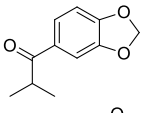
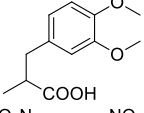
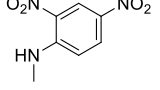
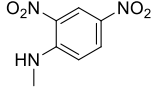
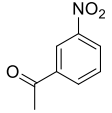
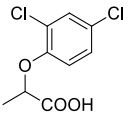
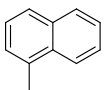
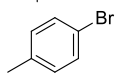
Cpd	2D Structure	PPAR α		PPAR γ		PPAR δ		XP score
		EC ₅₀ (μ M)	E _{max} ^a	EC ₅₀ (μ M)	E _{max} ^a	EC ₅₀ (μ M)	E _{max} ^a	
1		i	i	i	i	i	i	-6.47
2		i	i	i	i	i	i	-6.29
3		i	i	i	i	i	i	-6.04
4		i	i	i	i	i	i	-7.28
5		i	i	i	i	i	i	-6.41
6		4.4 $\pm$ 2.4	124 $\pm$ 12	14.8 $\pm$ 0.9	10 $\pm$ 6	i	i	-8.74
7		i	i	i	i	i	i	-7.68
8		i	i	i	i	i	i	-8.54
9		i	i	i	i	i	i	-8.98
10		i	i	i	i	i	i	-8.70
11		i	i	i	i	i	i	-6.76
12		i	i	i	i	i	i	-5.33
13		2.55 $\pm$ 0.35	148 $\pm$ 22	10.9 $\pm$ 2.4	16 $\pm$ 7	i	i	-8.47
14		i	i	i	i	i	i	-6.95

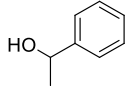
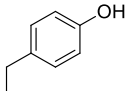
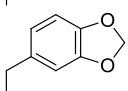
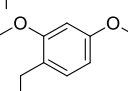
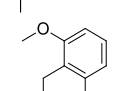
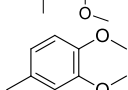
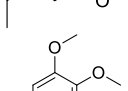
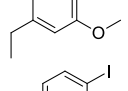
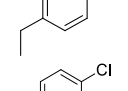
15		i	i	i	i	i	i	-9.36
16		i	i	i	i	i	i	-8.70
17		i	i	i	i	i	i	-6.83
18		i	i	i	i	i	i	-5.88
19		i	i	i	i	i	i	-8.53
20		i	i	i	i	i	i	-8.25
21		i	i	i	i	i	i	-7.43
22		i	i	i	i	i	i	-8.24
23		i	i	i	i	i	i	-6.87
24		i	i	i	i	i	i	-8.01
25		i	i	i	i	i	i	-5.96
26		i	i	i	i	i	i	-7.11
27		i	i	i	i	i	i	-7.36
28		i	i	i	i	i	i	-6.66
29		i	i	i	i	i	i	-6.47
30		i	i	i	i	i	i	-6.71

31		i	i	i	i	i	i	-10.16
32		i	i	i	i	i	i	-8.54
33		1.74 ± 0.05	42 ± 2	0.91 ± 0.34	53 ± 5	i	i	-7.16
34		i	i	i	i	i	i	-6.17
35		i	i	i	i	i	i	-7.68
36		i	i	i	i	i	i	-7.31
37		i	i	i	i	i	i	-7.27
38		2.55 ± 0.21	32 ± 1	11 ± 1	11 ± 4	8.4 ± 2	16 ± 1	-3.11
39		i	i	i	i	i	i	-8.28
40		18.85 ± 2.90	37 ± 12	i	i	i	i	-5.19
41		i	i	i	i	i	i	-5.97
42		i	i	i	i	i	i	-6.95
43		i	i	i	i	i	i	-7.17
44		i	i	i	i	i	i	-7.95
Wy-14,643		1.6 ± 0.3	100 ± 10	i	i	i	i	
Rosiglitazone		i	i	0.04 ± 0.02	100 ± 9	i	i	
L-165,041		i	i	i	i	1.6 ± 0.3	100 ± 10	

^a Efficacy values were calculated as the percentage of the maximum obtained fold induction with the reference compounds. i = inactive at tested concentrations.

Table S3. Structures of hits selected by substructure search.

 45-59  60-69							
Cpd	X	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆
45	CH ₂		H	H	H	H	H
46	CH ₂		H	COOH	H	H	H
47	CH ₂		H	H	H	H	H
48	CH ₂		H	H	H		H
49	CH ₂		H	H	OCH ₃	OCH ₃	OCH ₃
50	CH ₂		H	H	OCH ₃	OCH ₃	OCH ₃
51	CH ₂		H	H	H	OCH ₃	OCH ₃
52	CH ₂		H	H	H	H	H
53	CHOH		H	H	H	H	H
54	CO		H	H	NO ₂	H	H
55	O		H	Cl	H	Cl	H
56	O	CH ₃	CH ₃	H	H		H
57	O	H	H	H	H		H
58	O	H	H	H	H		H
59	O	CH ₃	H	H	H		H
60	-		H	-	-	-	-

61	-		H	-	-	-	-
62	-		H	-	-	-	-
63	-		H	-	-	-	-
64	-		H	-	-	-	-
65	-		H	-	-	-	-
66	-		H	-	-	-	-
67	-		H	-	-	-	-
68	-		I	-	-	-	-
69	-		I	-	-	-	-

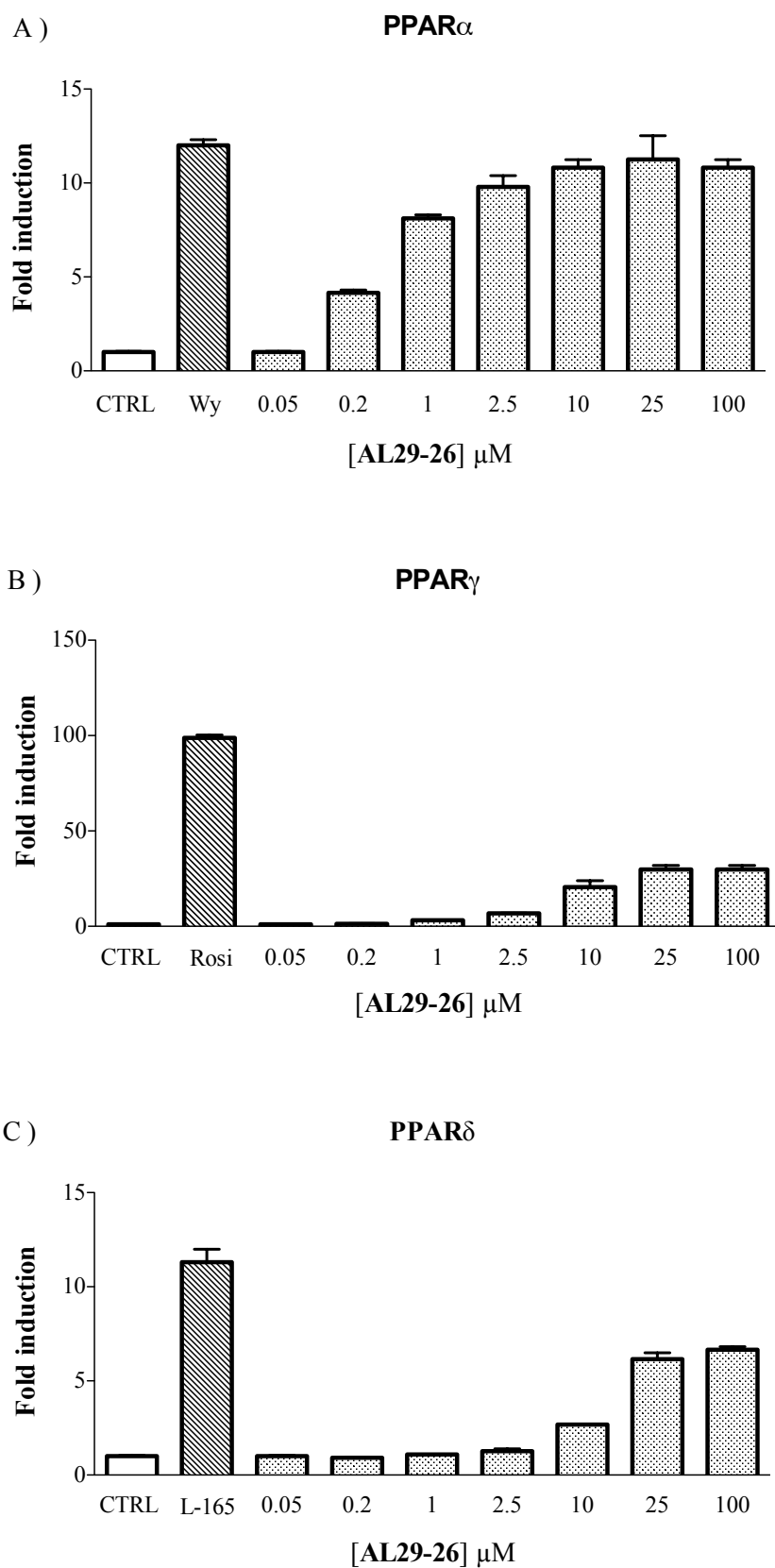


Figure S1. Transactivation assay on PPAR α (A), PPAR γ (B) and PPAR δ (C) of AL29-26 in HepG2 cells. Data are expressed as fold change compared with vehicle-treated cells (DMSO, 0.1%) and represent the mean of assays performed in triplicate

± SEM. Reference compounds: Wy-14,643 (10 µM) rosiglitazone (2 µM) and L-165,041 (2 µM).

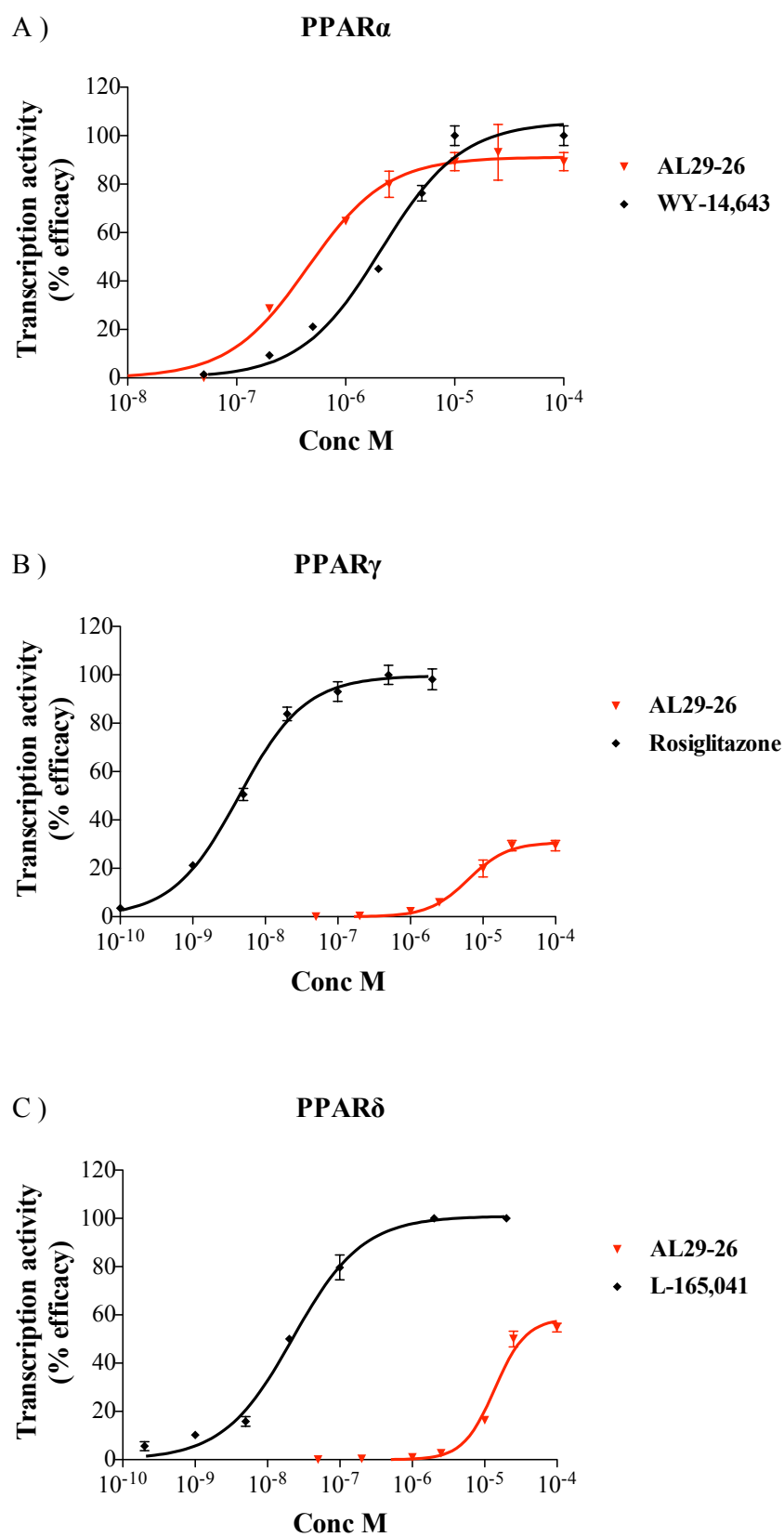


Fig. S2. Activation of GAL4-PPAR ligand binding domain chimeras in transiently transfected HepG2 cells by AL29-26. Dose–response curves are shown for human

PPAR α (A), PPAR γ (B) and PPAR δ (C). The curves are expressed as percentage of maximum effect obtained by reference compounds and represent the mean of assays performed in triplicate $\pm$ SEM.

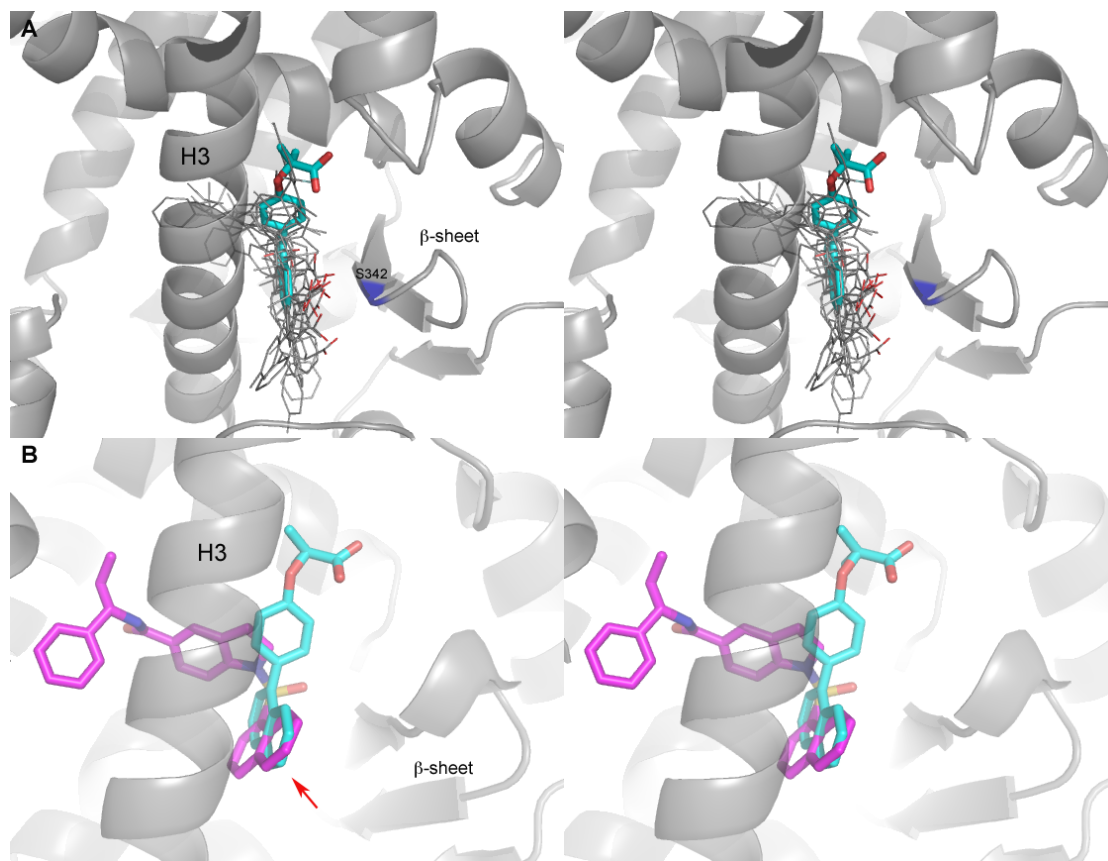


Figure S3. Comparison of PPAR γ complexes (stereo view)

(A) Superposition of PPAR γ complexes (gray) with known partial agonists (pdb codes: 3D6D, 4PVU, 4PWL, 4JL4, 4JAZ, 4E4K, 2Q5P, 2Q6S, 2Q5S, 4E4Q, 5F9B) onto the PPAR γ complex with AL29-26 (cyan). The carboxylate groups are depicted in red, the residue S342 in blue.

(B) Superposition of SR2067 (magenta) (pdb code 4R06) onto the PPAR γ complex with AL29-26 (cyan). The red arrow indicates the naphthalene groups of the two ligands.

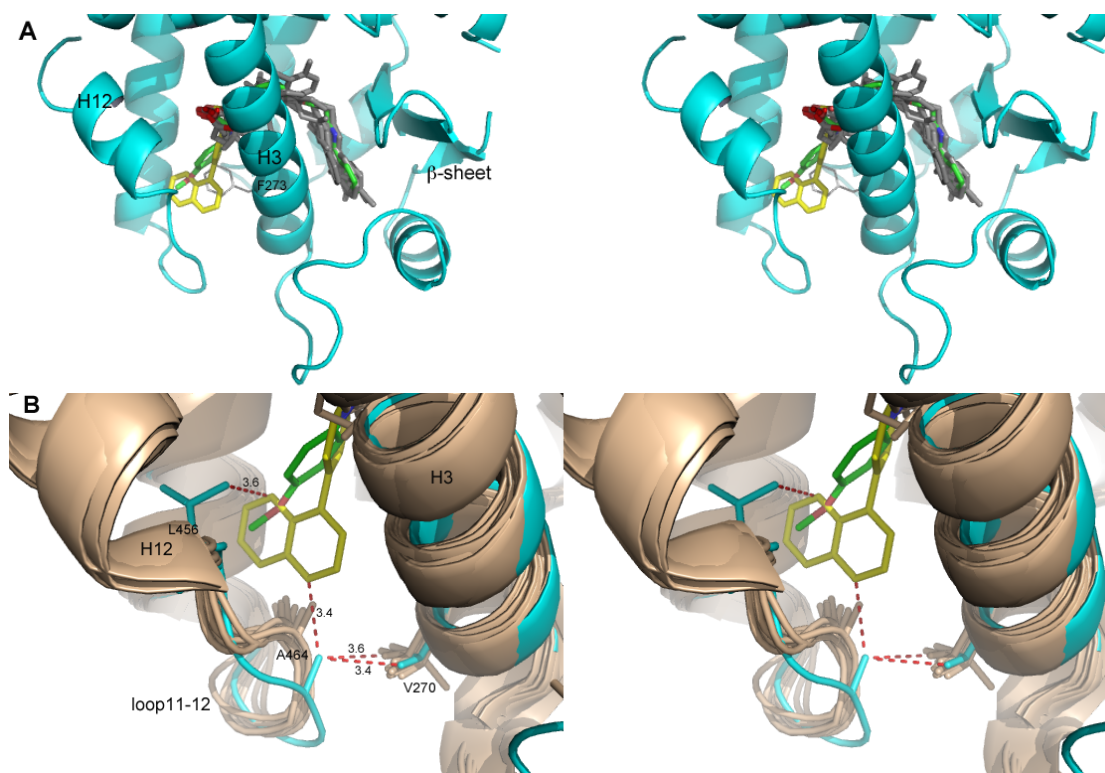


Figure S4. Comparison of PPARα complexes

(A) Superposition of PPARα complexes (gray) with known partial agonists (pdb codes: 2REW, 4BCR, 1K7L, 3SP6, 3FEI, 2GTK, 3G8I, 3ET1, 3KDT, 1I7G) onto the PPARα complex with AL29-26 (ligand yellow, protein cyan). The ligand BMS-631707 (PDB code 2REW) is shown in green. The "closed" (trans) conformation of F273 side-chain is also shown (gray).

(B) New conformation of the loop 11-12 in the PPARα/AL29-26 complex: superposition of the loops 11-12 of known PPARα structures (light-brown) (same pdb codes of Figure 5A) with that of PPARα/AL29-26 (ligand yellow, protein cyan). The ligand BMS-631707 (PDB code 2REW) is shown in green. Additional vdW interactions realized by AL29-26 are shown as red dashed lines.

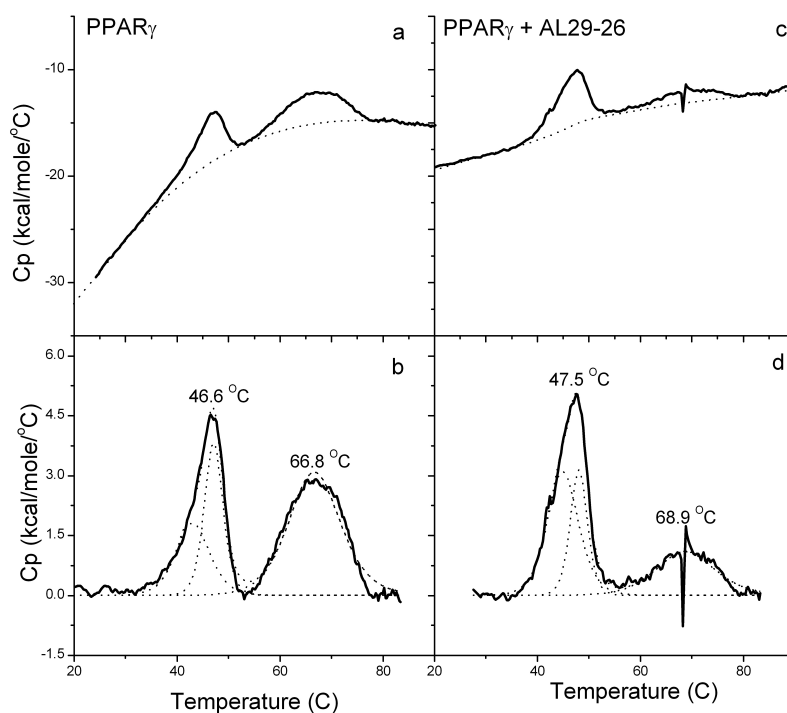


Figure S5. DSC thermograms of PPAR γ

(A) Experimental curve (solid line) and base line (dotted line) of PPAR γ 0.4 mg/ml in buffer Hepes 20 mM, pH8.0, TCEP 1 mM (scan speed 60 °C/h).

(B) Experimental curve after subtraction of the base line and deconvolution of the excess heat capacity function into three non-two-state transitions (dotted lines).

(C) Experimental curve (solid line) and base line (dotted line) of PPAR γ 0.4 mg/ml with AL29-26 20 μ M, in buffer Hepes 20 mM, pH8.0, TCEP 1 mM (scan speed 60 °C/h).

(D) Experimental curve after subtraction of the base line and deconvolution of the excess heat capacity function into three non-two-state transitions (dotted lines).

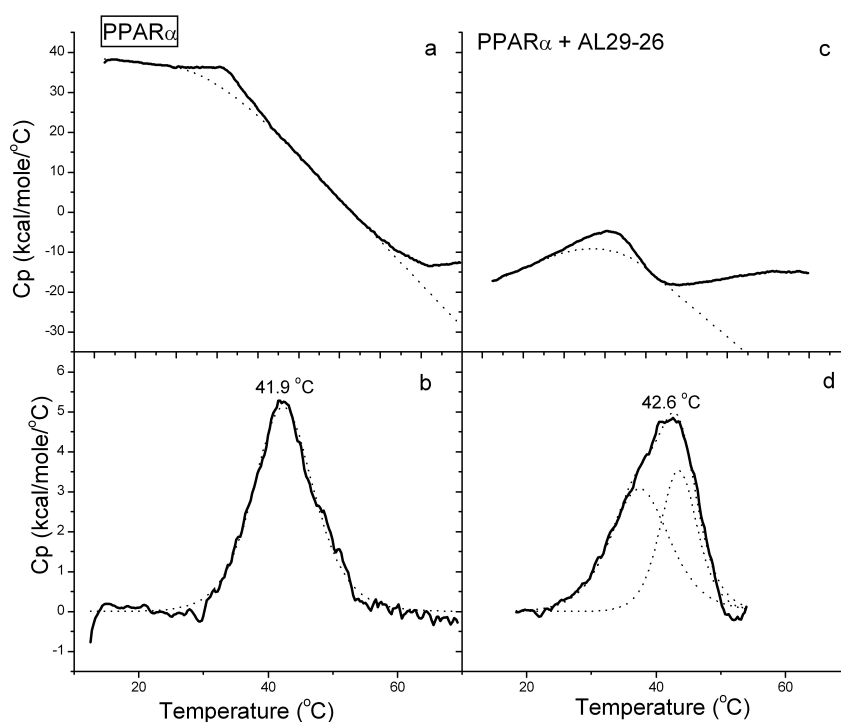


Figure S6. DSC thermograms of PPAR α

(A) Experimental curve (solid line) and base line (dotted line) of PPAR α 0.4 mg/ml in buffer Hepes 20 mM, pH8.0, TCEP 1 mM (scan speed 60 °C/h).

(B) Experimental curve after subtraction of the base line and deconvolution of the excess heat capacity function into one two-state transition (dotted lines).

(C) Experimental curve (solid line) and base line (dotted line) of PPAR α 0.4 mg/ml with AL29-26 20 μ M, in buffer Hepes 20 mM, pH8.0, TCEP 1 mM (scan speed 60 °C/h).

(D) Experimental curve after subtraction of the base line and deconvolution of the excess heat capacity function into two two-state transitions (dotted lines).

