

Supplemental information

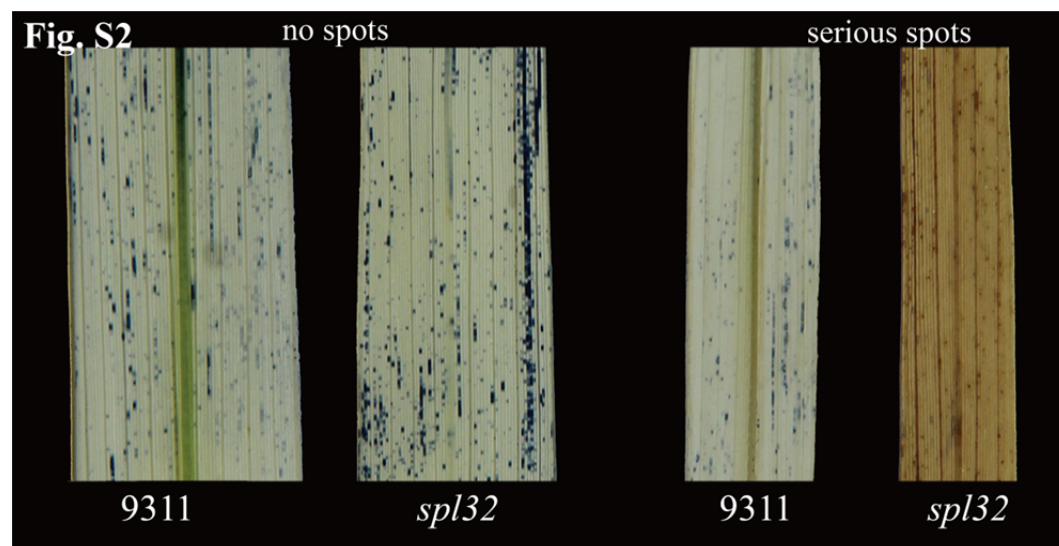
Title: *SPL32*, encoding a Fd-GOGAT, plays an important role in leaf senescence and defense response in rice (*Oryza sativa* L)

Liting Sun^{1,*}, Yihua Wang^{1,*}, Ling-long Liu¹, Chunming Wang¹, Ting Gan¹, Zhengyao Zhang¹, Yunlong Wang¹, Di Wang¹, Mei Niu¹, Wuhua Long¹, Xiaohui Li¹, Ming Zheng¹, Ling Jiang¹ & Jianmin Wan^{1,2}

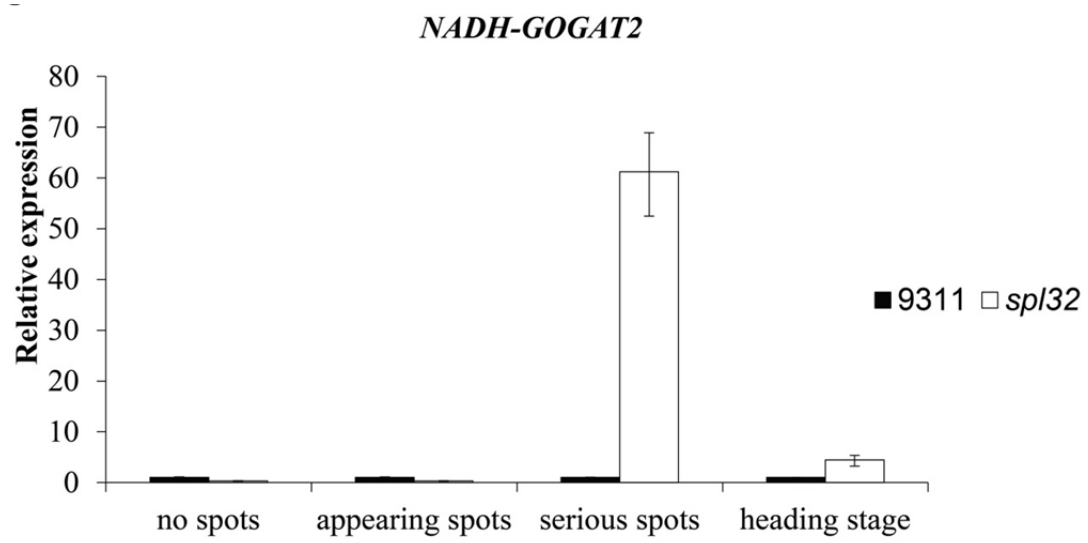
¹State Key Laboratory for Crop Genetics and Germplasm Enhancement, Jiangsu Plant Gene Engineering Research Center, Nanjing Agricultural University, Nanjing 210095, China. ²National Key Facility for Crop Gene Resources and Genetic Improvement, Institute of Crop Science, Chinese Academy of Agricultural Sciences, Beijing 100081, China. *These author contribute equally to this work. Correspondence and requests for materials should be addressed to Jianmin Wan (E-mail: wanjm@njau.edu.cn or wanjianmin@caas.cn; Tel & Fax: +86-25-84396516).



Supplemental Figure 1. Light induced experiment. We chose the leaves in the tillering stage. Leaves of 9311 and *spl32* mutant on the right panel were covered by tinfoil for 10 days.



Supplemental Figure 2. Dying experiments. Leaves with no spots or serious spots from *spl32* mutant and the corresponding sections of leaves in the wild-type 9311 at tillering stage were stained by nitrotriazolium blue chloride



Supplemental Figure3. Expression of *NADH-GOGAT2* in leaves with or without lesion spots at different developmental stages.

Supplemental Table S1 Segregation of the F₂ population from the wild-type 9311 and *spl32* mutant

Cross	WT	Mutant	The actual separation ratio	$\chi^2_{(3:1)}(\chi^2_{0.05}<3.84)$
<i>spl32</i> × 9311	117	38	3.08:1	0.0064
9311 × <i>spl32</i>	101	35	2.89:1	0.0294

Supplemental Table S2 Polymorphic InDel markers between parents in target region

primer	Forward primer sequence	Reverse primer sequence	PAC number
zzy-2	TTAGCCAATTCCATCAAG	ATTCTTTCCAGCGACAA	OJ1458_B07
zzy-3	CATTTGACCGTTCATCTT	TGACATCTCAGCGTTGCA	P0470D12
zzy-18	ATTGGTGGGTTGCTTGGT	GTGGGCATAGCCTTTGGT	OJ1477_F01
zzy-20	GGATCAATGCCATCTGTT	GCAATAAGCACTTGTTTCG	OJ1477_F01
zzy-21	GACAGACAACTTGGAACA	GTTTGCAGTCTTGACTCTTG	OJ1477_F01
zzy-22	TATCCCTTTCCTGTTCGT	TTGGAAAGACACCAGAAA	OJ1477_F01
zzy-23	AGACAGAGCCTTATTCAA	AACCTCCCAACCTTTCTA	P0496C02
zzy-24	CGTTACTTTGTTCGTCCTA	CTTGTCACGCATACTGTTG	P0496C02
zzy-30	ATGGATGGTTAACGTTCT	CTTAGCACTGGCTCTTCA	P0047B07
zzy31	TTGCATGATATTCACCTG	CCTATAAGATTCCCTAAAGT	OJ1477_F01

Supplemental Table S3 Gene-specific primers used in this study

Primers	Oligos	Function
Fd-GOGAT-GFP-F	AGCCCAGATCAACTAGTATGGCCACGCTCCACGTG	Subcellular
Fd-GOGAT-GFP-R	CACCATGGATCCCCCGGGCTTCGCCGATTGTACTGTTGT	Localization
CWW-11-1-F	TGGGTGGTGGTAGACGAG	Cloning of
CWW-11-1-R	CACTTGGAATTGCAGTAGA	<i>Fd-GOGAT</i>
zf-1-F	TGACTGGGCCAACAAGCAA	Verification of
zf-1-R	CATAGCGACCTACCACTGATACATT	57-bp deletion in <i>sp132</i>
GT-2 F	TGCAGGCCAGCTCAACATTACG	qRT-PCR for
GT-2 R	TCTCCACCAGCCATACCCTTTC	expression pattern analyses
Flag-F	TTCATTTTCATTTGGAGAGAA	Complemented
Flag-R	GGTACCCCGGGTTCGAAATC	verification
NADH-2 F	GGAAGATACGGACCAACT	qRT-PCR
NADH-2 R	AACATACAGAAGCAGCATT	
<i>PR1a</i> F	TTCATCACCTGCAACTACTCG	qRT-PCR
<i>PR1a</i> R	TGCATAAACACGTAGCATAGCAT	
<i>PR1b</i> F	GTGTGCGGGCACTACACG	qRT-PCR
<i>PR1b</i> R	CGGCTTATAGTTGCATGTGA	
<i>PR5</i> F	ATCGACGGCTACAACGTC	qRT-PCR
<i>PR5</i> R	GTGTCTTGGTGTGTCTTCG	
<i>PR10</i> F	CACCATCTACACCATGAAGC	qRT-PCR
<i>PR10</i> R	AGCACATCCGACTTTAGGAC	