

SUPPLEMENTARY INFORMATION

Grapevine Priming Under Adverse Temperature Stresses

**E. O. Lutskiy^a * (<https://orcid.org/0000-0003-0521-0827>), M. A. Sundyрева^a
(<https://orcid.org/0000-0002-1338-1725>), and M. O. Baranov^a (<https://orcid.org/0009-0009-5973-3886>)**

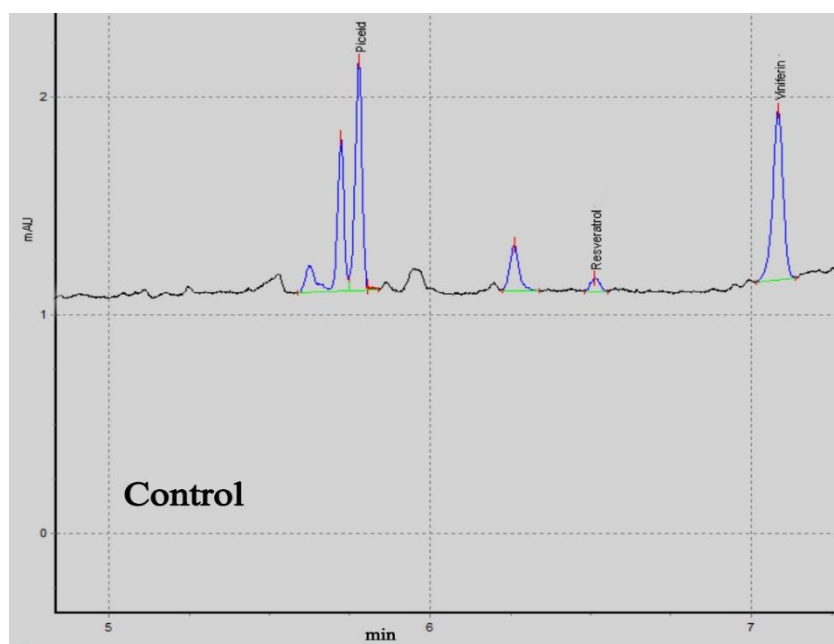
^a North Caucasian Federal Scientific Center of Horticulture, Viticulture, Winemaking, Krasnodar
350072, Russia

*e-mail: arcuswengart@mail.ru

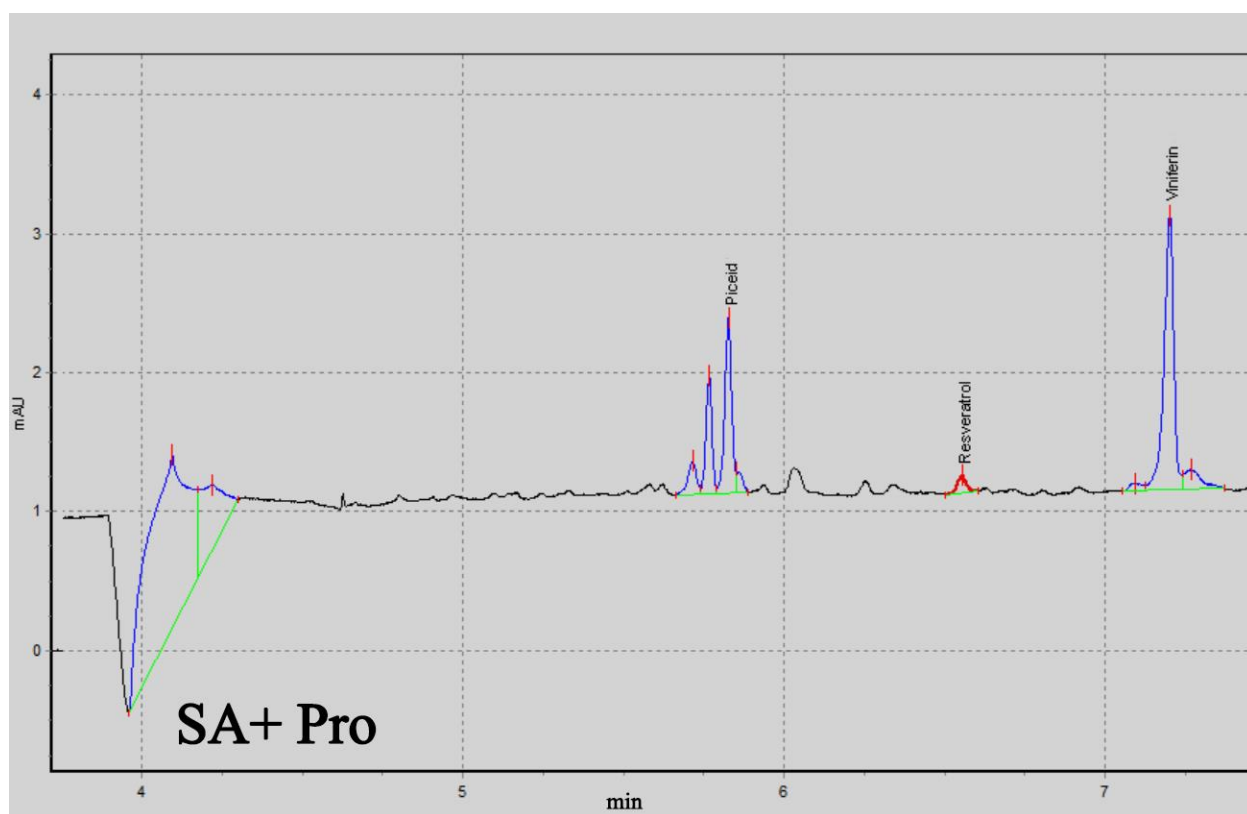
Supplementary Table S1. List of primers used for RT-PCR

Primer	Name	Sequence	Source
<i>GPX</i>	Glutathione peroxidase	F: GAGCACAGGAACCTGGGAGTAA R: AGCACTATCGCCATTCACATCA	https://doi.org/10.1007/s10658-015-0725-5
<i>NCED</i>	9-cis-epoxycarotenoid dioxygenase	F: CCTCTGTCTCACCGCAATGG R: AGAGAGTGATGGACGGGCTG	https://doi.org/10.5586/asb.p.3546
<i>RafS1</i>	RAF proto-oncogene serine/threonine-protein kinase	F: CCATCTTGAGGTGCCAGTACTACG R: ATGGTGTTGCCGTCGTGAAGAG	https://doi.org/10.3389/fpls.2021.754537
<i>GolS3</i>	Galactinol synthase 3	F: TGCCGAGCAGGACTATTTGA R: CAAGGTTGTAGGTGGGTGGA	https://doi.org/10.3389/fpls.2021.754537
<i>SS4</i>	Starch synthase 4	F: TGGCTACTTTGCTCAGGACA R: GTCCTTGCTGCTTGATACGG	https://doi.org/10.3389/fpls.2020.00321
<i>TPPB</i>	Trehalose-6-phosphate phosphatase B	F: GATCCAAAATCAGCTATTGGTATGG R: GTACCCACCCGGTACAGTCG	https://doi.org/10.1007/s00425-012-1611-4
<i>TRE</i>	Trehalase	F: TGAGGTGGATCAGAACCAACTATG R: TTCACCGCCACCTCCAATT	https://doi.org/10.1007/s00425-012-1611-4
<i>ICE1</i>	Inducer of CBF Expression 1	F: GCCAAGGGTTGAAGTGA R: CCGCCTGTTGAACGTCTAG	https://doi.org/10.3389/fpls.2022.1021483
<i>CBF1</i>	C-repeat/DRE Binding Factor 1	F: GGG GCG AAG GTC GTG TT R: GTC CCA TCG TTT CCA TTT T	https://doi.org/10.3389/fpls.2022.1021483
<i>CBF3</i>	C-repeat/DRE Binding Factor 3	F: GCA TAA GCG GAA AGC AG R: CCG CAC TTC GCA TAC CC	https://doi.org/10.3389/fpls.2022.1021483
<i>CBF4</i>	C-repeat/DRE Binding Factor 4	F: ACCCTCACCCGCTCGTATG R: CCGCGTCTCCCGAAACTT	https://doi.org/10.3389/fpls.2018.00970

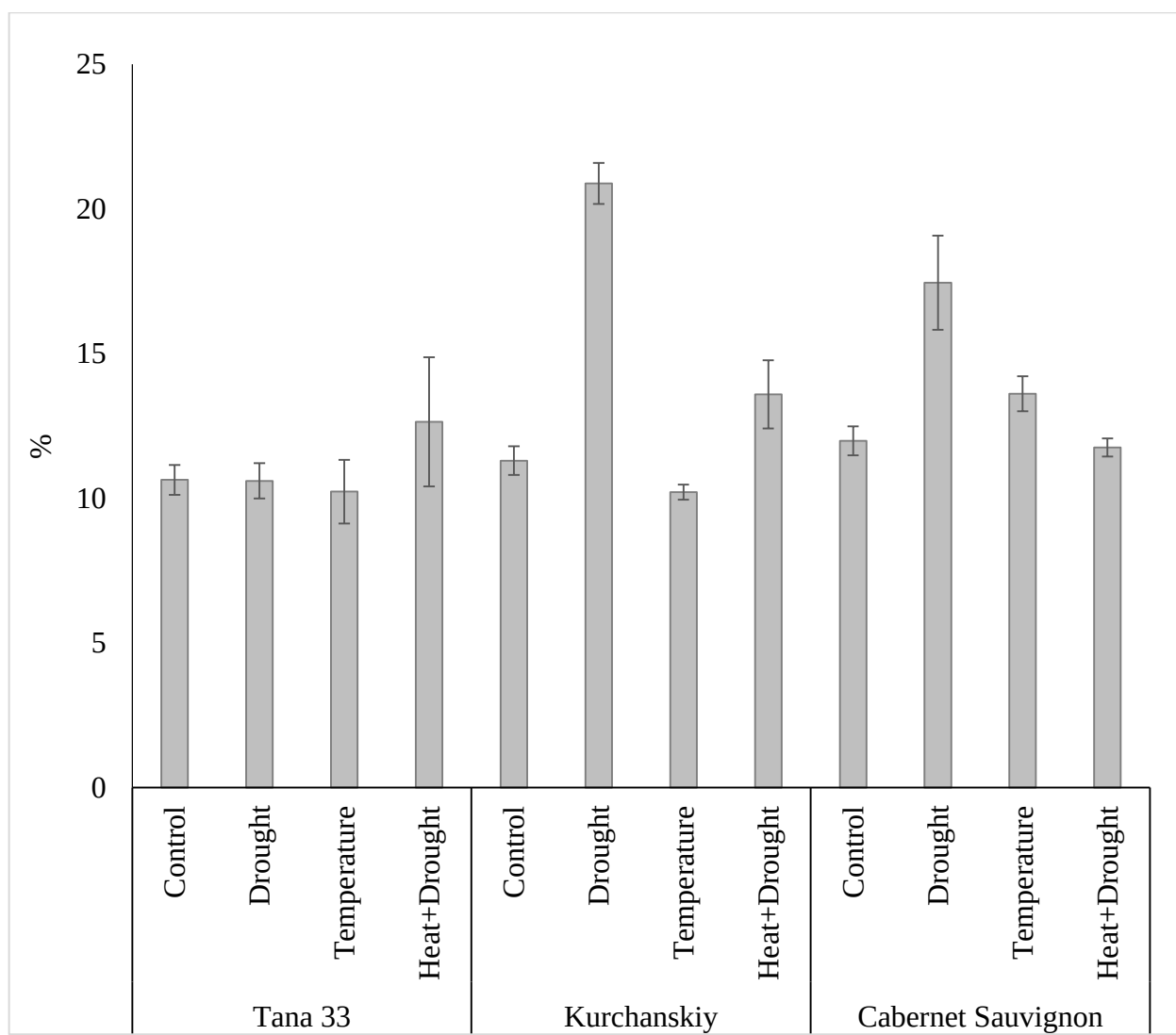
<i>O6fdes</i>	Delta-6 fatty acid desaturase	F: CAATTCGGCCTTGGATGTCT R: GCCAAACTTATCTTCACCCTCTT	https://doi.org/10.1371/journal.pone.0186952
<i>HSP17</i>	17.7 kDa class I heat shock protein	F: GAAGAAGAGCCAGAAGAGAAG G: ACACACGAAGCGACCAAG	https://doi.org/10.1186/1756-0500-5-220
<i>APX</i>	Ascorbate peroxidase	F: GGTCCGTTTGGGACAATGAA R: CGGAAATTGCTCCTTGATCG	https://doi.org/10.1007/s10658-015-0725-5
<i>O3fdes</i>	Omega-3 fatty acid desaturase	F: GGTATCGTGGAAAGGAATGGAG R: GTGATAGTGCGGGATTTGAGG	https://doi.org/10.1371/journal.pone.0186952
<i>OSM</i>	Osmotin	F: ACTGCAACTTCGATGCGTCA R: TGCGAATTGCGCTAAGGTGT	https://doi.org/10.1007/s10658-015-0725-5
<i>STS</i>	Strictosidine synthase	F: ATC GAA GAT CAC CCA CCT TG R: CTTAGCGGTTCTGAAG GACAG	https://doi.org/10.1186/s12864-016-3304-z
<i>NPRI.1</i>	Nonexpresser of PR genes 1.1	F: GCG GAA AGA GCC CAA GAT TA R: CTG GTG AGC CTC TTT GCT ATT	https://doi.org/10.1007/s11240-017-1260-1
<i>PR1</i>	Pathogenesis-related protein 1	F: CCC AGA ACT CTC CAC AGG A R: GCA GCT ACA GTG TCG TTC CA	https://doi.org/10.1111/j.1365-3059.2012.02628.x
<i>OPR3</i>	Oxophytodienoate-reductase 3	F: GACGAGGTGCAGGGCGATG R: CGGTCCTTTCCAGGGTCTTT	https://doi.org/10.1016/j.plaphy.2019.09.024
<i>ABA2</i>	Abscisic acid biosynthesis 2	F: TGGCTGGTTTCCGTGCTTTC R: ATGTACCTCGCCTCGTCGC	https://doi.org/10.5586/asbp.3546
<i>PAL</i>	Phenylalanine ammonia-lyase	F: ACA ACA ATG GAC TGC CAT CA R: CAC TTT CGA CAT GGT TGG TG	https://doi.org/10.1111/j.1365-3059.2012.02628.x
<i>MYC2</i>	Transcription factor MYC2	F: GCC AGC CTT ACC AGT GTT G R: GCC TCT CTT CTG CCT TTC G	https://doi.org/10.1016/j.jcrep.2024.113825
<i>SNRK</i>	SNF1-related serine/threonine-protein kinase	F: CCCATCCCAAAGTGTAGAAGAG R: TCATCAGCGTCAATGTCATCAAG	https://doi.org/10.1007/s11240-020-01872-9
<i>PYL4</i>	Abscisic acid receptor PYL4	F: CCGTCGTCCAGCAAATCG R: ACATCTCCATCGCCAACAAC	https://doi.org/10.1007/s11240-020-01872-9
<i>JAZ</i>	Jasmonate ZIM-domain protein	F: AGAAACAGAGGAACCAAAACCG R: CCGCCAGACAGCGACAAC	https://doi.org/10.1007/s11240-020-01872-9
<i>AOC</i>	Allene oxide cyclase	F: CTACACGGGAGACCTGGAGAAG R: CTCCTTCTTCCCTGGAACATT	https://doi.org/10.1007/s10658-015-0725-5
<i>AAO3</i>	Abscisic aldehyde oxidase 3	F: ATTGCTTCAGGGTTCATCCG R: AGCTCGATACGTTTCAAACA	https://doi.org/10.5586/asbp.3546
<i>AOS</i>	Allene oxide synthase	F: CTTGCGAGGGATGAAAATCA R: GGCTAATTGGGTGTGCAGTT	https://doi.org/10.1007/s11033-018-4159-y
<i>JAR1</i>	Jasmonic acid-amido synthetase	F: TGGGATCATGACTGGCTC R: TGTCCCTCCTCAGCACTCCCC	https://doi.org/10.1007/s11240-020-01872-9
<i>VSP2</i>	Vegetative storage protein 2	F: AATGGGTGAATTTGGCTGA R: GACATTCCTCTGGGGTTCAA	https://doi.org/10.1016/j.plaphy.2019.09.024
<i>GAPDH</i>	Glyceraldehyde-3-phosphate dehydrogenase	F: TTCTCGTTGAGGGCTATTCCA R: CCACAGACTTCATCGGTGACA	https://doi.org/10.1007/s11033-018-4159-y



Supplementary Fig. S1. Electropherogram for stilbene count (control).



Supplementary Fig. S2. Electropherogram for stilbene count (treatment by salicylic acid (SA) and proline).



Supplementary Fig. S3. The comparative analysis of EL in different grape cultivars, tested in combined drought and high temperature stress.