

Additional file 4 - Genomic organisation and transcription of two copies of *F3'Hs* present in the grapevine genome

In **section a**, exon/intron structure of *F3'Hs* is shown as blue boxes (exons) connected by blue lines (introns); TEs are shown as coloured boxes.

In **section b** and **c**, selective amplification of exon junctions astride the terminal intron and expression of each *F3'H* copy. Two primer pairs (orange and green triangles) were designed in the internal and terminal exons. The terminal intron varied in size between 249 bp and 96 bp in *F3'Ha* and *F3'Hb*, respectively. Each primer pair anneals perfectly to the target *F3'H*, but has a mismatch at the 3' terminal nucleotide with the paralogous *F3'H*. Selectivity of primer pairs for either *F3'Ha* or *F3'Hb* was validated by amplifying PN40024 genomic DNA and by Sanger sequencing of the PCR amplicons. Selectivity for either *F3'Ha* or *F3'Hb* was also confirmed by assessing the size of the amplified genomic DNA (vs. the size prediction of 523 bp and 370 bp astride the second intron in *F3'Ha* and *F3'Hb*, respectively) and, for the expressed *F3'Ha*, by inferring intron size from the comparison between amplicons from genomic DNA and cDNA. Expression of *F3'Ha* was assessed by semi-quantitative PCR using cDNA from leaf, petiole, tendril, flower, shoot, and berry skin and flesh, in two grapevine cultivars ('Merlot' and 'Aglanico'). Expression of *F3'Ha* was also assessed in berry skin of four cultivars ('Aglanico', 'Marzemino', 'Grignolino', and 'Nebbiolo') at four stages of fruit development. cDNA was normalised using the constitutive gene *VvUbiquitin*. Transcripts of *F3'Hb* were never detected under the same experimental conditions.

In **section d**, expression of *F3'Ha* was assessed by quantitative PCR in berry skin at 8 developmental stages in the cultivars 'Aglanico', 'Marzemino', 'Grignolino', and 'Nebbiolo'. Transcript levels of *F3'Ha* increased at full-veraison (stage of 100% coloured berries) by approximately 2-fold in all cultivars, with substantial differences among cultivars only at harvest. Transcript levels are expressed as arbitrary units, normalised using the constitutive gene coding for *VvUbiquitin*. Bars represent standard deviation of three biological replicates. Differences among cultivars at each sampling date were tested for significance by one-way ANOVA. Means were separated by a Student–Newman–Keuls test and significant differences at $P < 0.05$ are indicated by different letters.

