

Supplementary Material

Novel loading protocol combines highly efficient encapsulation of exogenous therapeutic toxin with preservation of extracellular vesicles properties, uptake and cargo activity

Stefania Zuppone¹, Natasa Zarovni², Kosuke Noguchi³, Francesca Loria², Carlo Morasso⁴, Andres Lohmus², Ikuhiko Nakase³, Riccardo Vago^{1,5} (✉)

¹ Urological Research Institute, Division of Experimental Oncology, IRCCS San Raffaele Scientific Institute, Milano, 20132, Italy.

² HansaBiomed Life Sciences, Tallinn, 12618, Estonia

³ Department of Biological Science, Graduate School of Science, Osaka Prefecture University, Osaka, 599-8531, Japan

⁴ Istituti Clinici Scientifici Maugeri IRCCS, Pavia, 27100, Italy

⁵ Università Vita-Salute San Raffaele, Milano, 20132, Italy

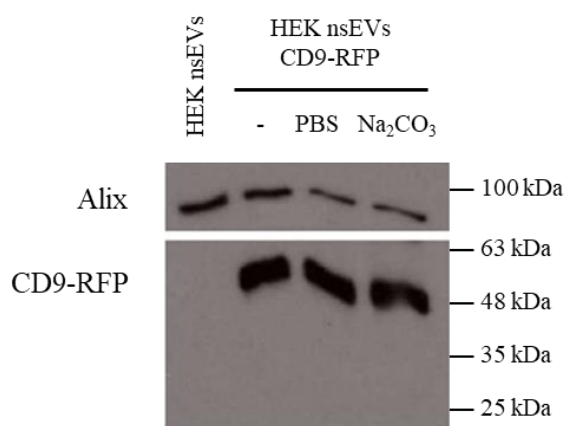


Fig. S1. HEK293 derived nsEVs structural stability upon sodium carbonate treatment. Western blot analysis Alix and RFP from untreated (-), PBS treated and sodium carbonate (Na₂CO₃) treated nsEVs derived from CD9-RFP expressing HEK293 cells. HEK293 WT derived nsEVs were used as negative control for CD9-RFP expression. CD9-RFP has been detected using anti-RFP antibody.

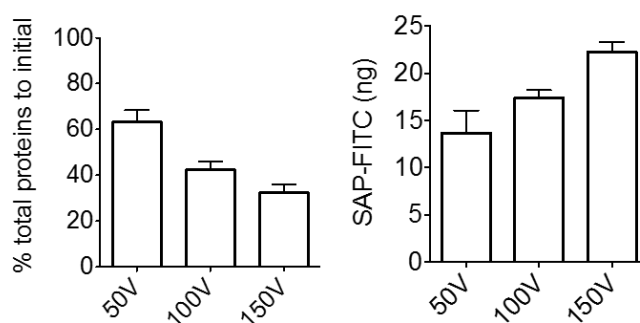


Fig. S2. Protein recovery and encapsulation efficiency evaluation after increasing voltages exposition and higher nsEVs:SAP-FITC ratios. A. BCA assay for quantification of total protein recovery after co-incubation of HEK293-derived nsEVs (25 ug/ml) with SAP-FITC (1:2 molecular ratio) and exposition to 50, 100, 150 V electroporation (see material and methods). Results are shown as percentage to initial. B. Quantification of SAP-FITC incorporation after incubation with HEK293-derived nsEVs (25 ug/ml) at a 1:2 molecular ratio and 50, 100, 150 V electroporation (see Material and Methods) by spectrofluorometer analysis.

Full blots

Fig. 1C

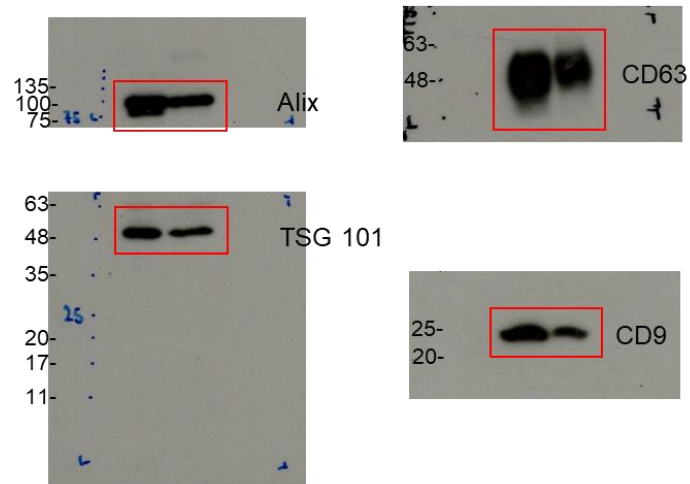


Fig. 6A



Sup. Fig. 1

