

Table S1. Bacterial strains and plasmids used in the study.

Strain or plasmid	Description and/or genotype	Reference
<i>Escherichia coli</i>		
JM107	General laboratory strain used for all cloning experiments and DNA manipulations. <i>endA1</i> , <i>glnV44</i> , <i>thi</i> ⁻ , <i>relA1</i> , <i>gyrA96</i> , $\Delta(lac-proAB)$ [F', <i>traD36</i> , <i>proAB</i> ⁺ , <i>lacI</i> ^q ZΔM15], <i>hsdR17</i> (<i>R</i> _K ⁻ <i>m</i> _K ⁺), λ ⁻ .	[1]
DH5α	Strain used for inter-bacterial competitive growth assays as a control. <i>F</i> ⁻ , $\Delta(argF-lac)169$, $\phi80dlacZ58(M15)$, $\Delta phoA8$, <i>glnX44</i> (AS), λ ⁻ , <i>deoR481</i> , <i>rfbC1</i> , <i>gyrA96</i> (<i>NalR</i>), <i>recA1</i> , <i>endA1</i> , <i>thiE1</i> , <i>hsdR17</i> .	[2]
MC4100	Strain used for inter-bacterial competitive growth assays as prey <i>F</i> ⁻ , [<i>araD139</i>] _{B/r} , $\Delta(argF-lac)169$, λ ⁻ , <i>e14</i> ⁻ , <i>flhD5301</i> , $\Delta(fruK-yeiR)725(fruA25)$, <i>relA1</i> , <i>rpsL150(strR)</i> , <i>rbsR22</i> , $\Delta(fimB-fimE)632(::IS1)$, <i>deoC1</i> .	[3]
BL21(DE3)	Genomic DNA used as a template for the amplification of <i>lacI</i> ^q <i>F</i> ⁻ , <i>lon-11</i> , $\Delta(ompT-nfrA)885$, $\Delta(galM-ybhJ)884$, $\lambda DE3$ [<i>lacI</i> , <i>lacUV5-T7 gene 1</i> , <i>ind1</i> , <i>sam7</i> , <i>nin5</i>], $\Delta46$, [<i>mal</i> ⁺] _{K-12} (λ ^S), <i>hsdS10</i> .	[4]
<i>Bacillus subtilis</i>	Used for the amplification of the <i>sacB</i> gene.	Gift from Audrius Gegeckas
<i>Acinetobacter sp.</i>		
V15	Clinical isolate, IC_other, unique pulsotype U18.	[5]
V15 $\Delta bfmRS$	V15 derivative with deleted <i>bfmRS</i> operon.	This study
V15 $\Delta bfmRS$ <i>pbfmRS</i>	V15 $\Delta bfmRS$ complemented with pUC_AcORI_Ptac_ <i>bfmRS</i> _TER_ <i>lacI</i> ^q 2.	This study
V15 $\Delta bfmRS$ <i>pbfmR</i>	V15 $\Delta bfmRS$ complemented with pUC_AcORI_Ptac_ <i>bfmR</i> _TER_ <i>lacI</i> ^q 2.	This study
V15 $\Delta bfmRS$ <i>pbfmS</i>	V15 $\Delta bfmRS$ complemented with pUC_AcORI_Ptac_ <i>bfmS</i> _TER_ <i>lacI</i> ^q 2.	This study
V15 Δhcp	V15 derivative with deleted <i>hcp</i> gene.	This study
V15 Δhcp <i>phcp</i>	V15 Δhcp complemented with pUC_AcORI_Ptac_ <i>hcp</i> _TER_ <i>lacI</i> ^q 2.	This study
V15 $\Delta bfmRS\Delta hcp$	V15 $\Delta bfmRS$ derivative with deleted <i>hcp</i> gene.	This study

V15 $\Delta bfmRS\Delta hcp$ $pbfmRS$	V15 $\Delta bfmRS\Delta hcp$ complemented with pUC_AcORI_Ptac_bfmRS_TER_lacI ^q 2.	This study
V15 $\Delta bfmRS\Delta hcp$ $pbfmR$	V15 $\Delta bfmRS\Delta hcp$ complemented with pUC_AcORI_Ptac_bfmR_TER_lacI ^q 2.	This study
V15 Δcdi^{V15}	V15 derivative with partial deletion of <i>cdiBAI</i> operon.	This study
V15 $\Delta hcp\Delta cdi^{V15}$	V15 Δhcp derivative with partial deletion of <i>cdiBAI</i> operon.	This study
V15 $\Delta bfmRS\Delta cdi^{V15}$	V15 $\Delta bfmRS$ derivative with partial deletion of <i>cdiBAI</i> operon.	This study
V15 $\Delta bfmRS\Delta hcp\Delta cdi^{V15}$	V15 $\Delta bfmRS\Delta hcp$ derivative with partial deletion of <i>cdiBAI</i> operon.	This study
V15 $\Delta bfmRS\Delta cdi^{V15}$ $pbfmRS$	V15 $\Delta bfmRS\Delta cdi^{V15}$ complemented with pUC_AcORI_Ptac_bfmRS_TER_lacI ^q 2.	This study
V15 $\Delta bfmRS\Delta cdi^{V15}$ $pbfmR$	V15 $\Delta bfmRS\Delta cdi^{V15}$ complemented with pUC_AcORI_Ptac_bfmR_TER_lacI ^q 2.	This study
V15 $\Delta bfmRS\Delta hcp\Delta cdi^{V15}$ $pbfmRS$	V15 $\Delta bfmRS\Delta hcp\Delta cdi^{V15}$ complemented with pUC_AcORI_Ptac_bfmRS_TER_lacI ^q 2.	This study
V15 $\Delta bfmRS\Delta hcp\Delta cdi^{V15}$ $pbfmR$	V15 $\Delta bfmRS\Delta hcp\Delta cdi^{V15}$ complemented with pUC_AcORI_Ptac_bfmR_TER_lacI ^q 2.	This study
<i>A. baylyi</i> ADP1		ATCC 33305
<i>A. baylyi</i> ADP1 $pcdiI^{V15}$	<i>A. baylyi</i> ADP1 strain complemented with pUC_AcORI_Ptac_cdiI ^{V15} _TER_lacI ^q 2_gm.	This study
ABV36/II-a1	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-a1	[5,6]
AB52/II-a	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-a	
ABV63/II-a2	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-a2	
ABS11/II-a1	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-a1	
ABS08/II-a3	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-a3	
AB15/II-b	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-b	
AB60/II-c	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-c	
AB330/II-d	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-d	
ABV22/II-d	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-d	
AB323/II-e	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-e	
AB141/II-f	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-f	
AB106/II-g	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-g	
AB109/II-h	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-h	

AB288/II-j2	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-j2	
AB282/II-j4	Clinical <i>A. baumannii</i> isolate, IC II, pulsotype II-j4	
Other		
<i>Pseudomonas aeruginosa</i> P16	Clinical isolate	[7]
<i>Klebsiella pneumoniae</i> K39	Clinical isolate	[8]

Plasmids

pUC19	Used as a scaffold for the construction of suicide pUC19_ <i>sacB</i> ; Amp ^R .	[9]
pWH1266	<i>Acinetobacter calcoaceticus</i> plasmid fragment containing <i>ori</i> cloned to pBR322	[10]
pUC19_ <i>sacB</i>	<i>A. baumannii</i> suicide vector containing <i>sacB</i> gene from <i>Bacillus</i> spp. cloned via <i>Xba</i> I and <i>Pae</i> I; Amp ^R .	This study
pUC19_ <i>sacB</i> _ <i>bfmRS</i> UPDwn_ gmR	pUC19_ <i>sacB</i> derivative with $\Delta bfmRS::aac3I$; Amp ^R ; Gm ^R .	This study
pUC19_ <i>sacB</i> _ <i>hcp</i> UP Dwn_ gmR	pUC19_ <i>sacB</i> derivative with $\Delta hcp::aac3I$; Amp ^R ; Gm ^R .	This study
pUC_ gm	<i>A. baumannii</i> <i>aac3I</i> gene cloned into pUC19; Amp ^R ; Gm ^R .	This study
pUC_ gm_ AcORI	<i>Acinetobacter</i> sp. <i>ori</i> cloned to pUC19_ gm; Amp ^R ; Gm ^R .	This study
pUC_ gm_ AcORI_ <i>gfp</i>	pUC_ gm_ AcORI derivative with <i>gfp</i> gene from pAcGFP1-C3 cloned downstream to <i>Acinetobacter</i> sp. <i>ori</i> ; Amp ^R , Gm ^R .	This study
pUC_ gm_ AcORI_ <i>Pta</i> c_ <i>gfp</i>	pUC_ gm_ AcORI_ <i>gfp</i> derivative with <i>Ptac</i> from pKK223-3 cloned downstream to <i>Acinetobacter</i> sp. <i>ori</i> ; Amp ^R ; Gm ^R .	This study
pUC_ gm_ AcORI_ <i>Pta</i> c_ <i>gfp</i> _ <i>TER</i>	pUC_ gm_ AcORI_ <i>Ptac</i> _ <i>gfp</i> derivative with terminator (TER) sequence from pKK223-3 cloned downstream <i>gfp</i> gene; Amp ^R ; Gm ^R .	This study
pUC_ AcORI_ <i>Ptac</i> _ <i>gfp</i> _ <i>TER</i> _ <i>lacI</i> ^{q2}	pUC_ gm_ AcORI_ <i>Ptac</i> _ <i>gfp</i> _ <i>TER</i> derivative with <i>lacI</i> ^q downstream the terminator sequence, replacing <i>aac3I</i> ; Amp ^R .	This study
pUC_ AcORI_ <i>Ptac</i> _ <i>TE</i> R_ <i>lacI</i> ^{q2}	pUC_ AcORI_ <i>Ptac</i> _ <i>gfp</i> _ <i>TER</i> _ <i>lacI</i> ^{q2} derivative where <i>gfp</i> gene is removed; Amp ^R .	This study
pUC_ AcORI_ <i>Ptac</i> _ <i>bfmRS</i> _ <i>TER</i> _ <i>lacI</i> ^{q2}	pUC_ AcORI_ <i>Ptac</i> _ <i>gfp</i> _ <i>TER</i> _ <i>lacI</i> ^{q2} derivative where <i>gfp</i> gene is replaced with <i>bfmRS</i> operon; Amp ^R .	This study

pUC_AcORI_Ptac_bf mR_TER_lacI ^q 2	pUC_AcORI_Ptac_gfp_TER_lacI ^q 2 derivative where <i>gfp</i> gene is replaced with wild-type <i>bfmR</i> allele; Amp ^R .	This study
pUC_AcORI_Ptac_bf mS_TER_lacI ^q 2	pUC_AcORI_Ptac_gfp_TER_lacI ^q 2 derivative where <i>gfp</i> gene is replaced with wild-type <i>bfmS</i> allele; Amp ^R .	This study
pUC_AcORI_Ptac_hc p_TER_lacI ^q 2	pUC_AcORI_Ptac_gfp_TER_lacI ^q 2 derivative where <i>gfp</i> gene is replaced with <i>hcp</i> gene; Amp ^R .	This study
pUC_AcORI_Ptac_im m ^{V15} _TER_lacI ^q 2	pUC_AcORI_Ptac_gfp_TER_lacI ^q 2 derivative where <i>gfp</i> gene is replaced with <i>cdiI</i> ^{V15} gene; Amp ^R .	This study
pUC_AcORI_Ptac_cdi I ^{V15} _TER_lacI ^q 2_gm	pUC_AcORI_Ptac_imm ^{V15} _TER_lacI ^q 2 derivative where <i>bla</i> is replaced with <i>aac3I</i> ; Gm ^R .	This study
pAcGFP1-C3	Used as a source for <i>gfp</i> gene; Kan ^R .	Clonetech laboratories Inc.
pKK223-3	Used as a source for <i>Ptac</i> promoter and termination sites; Amp ^R .	Cloning Vector from PL- Pharmacia. GenBank:M77 749.1

Amp^R, ampicillin resistant; Gm^R, gentamicin resistant; Kan^R, kanamycin resistant

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