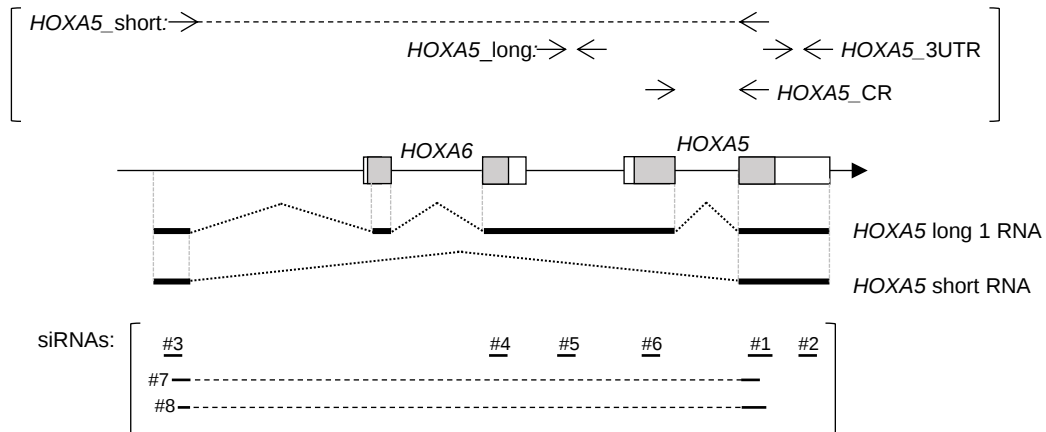


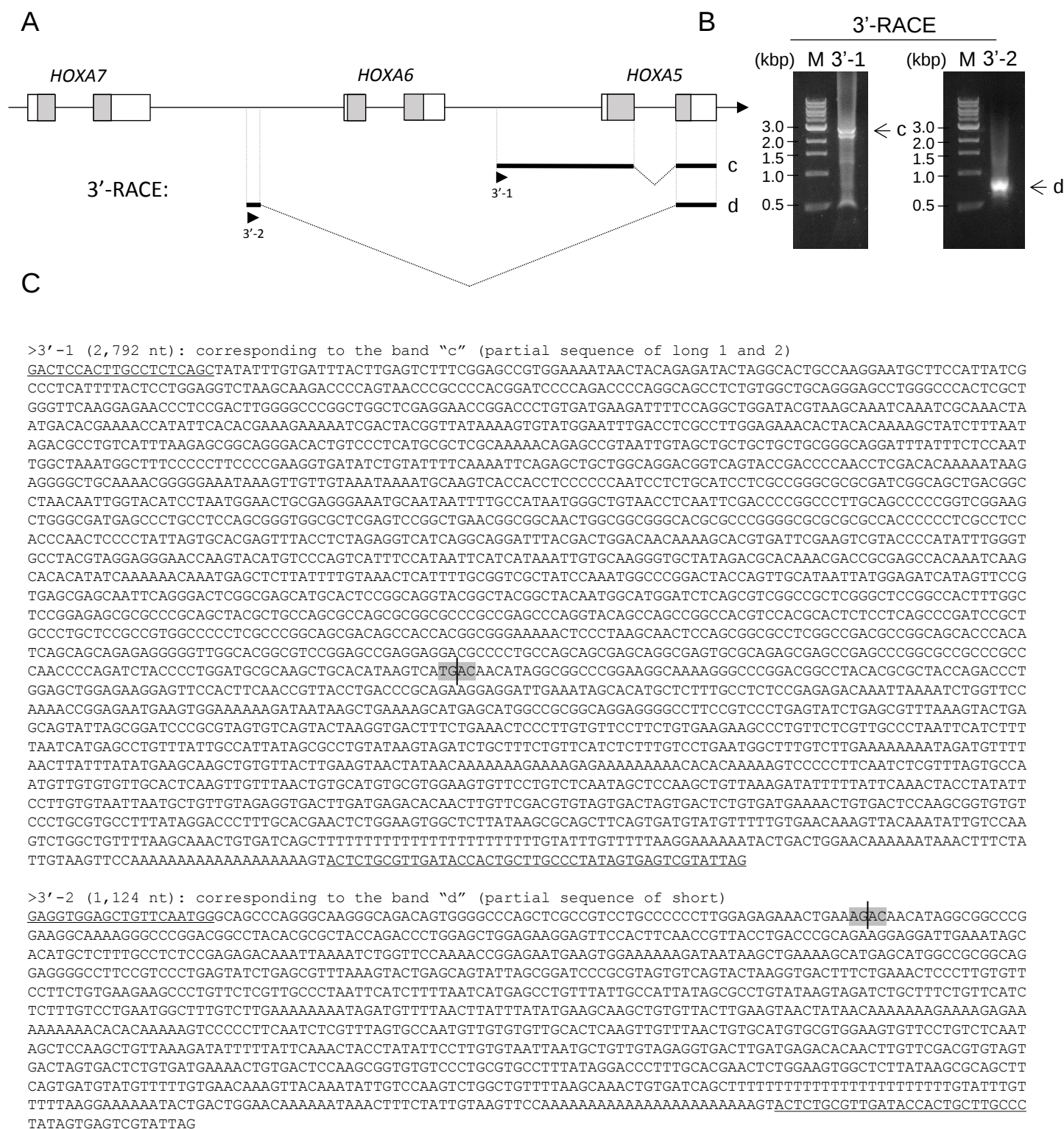
Primer sets for qPCR:



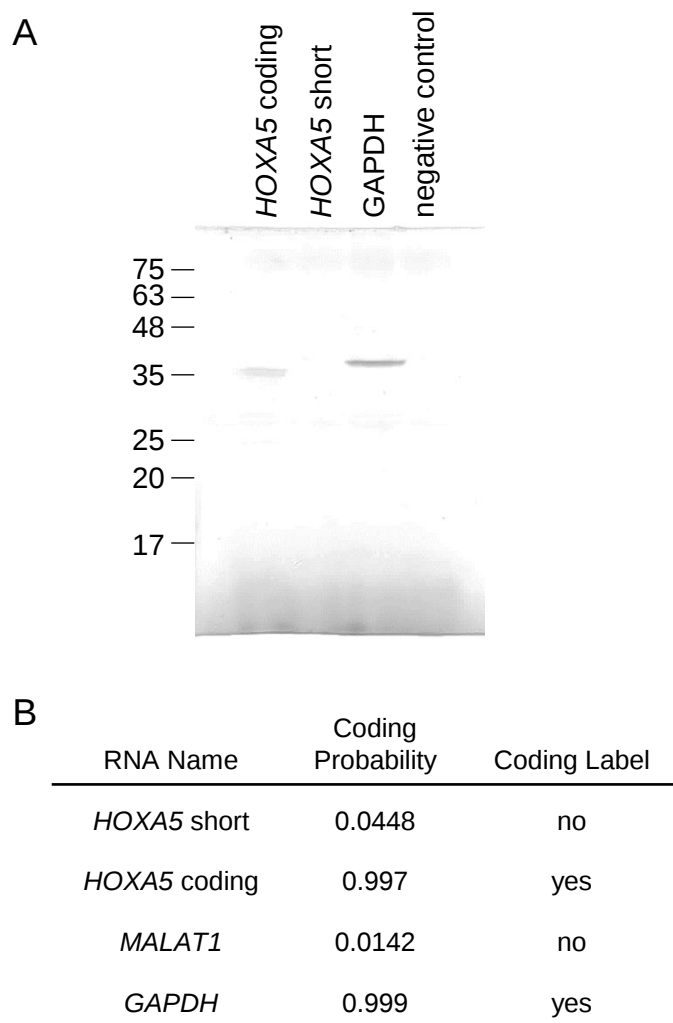
**Figure S1. Schematic representation of the primer sets used for the qPCR assay and siRNAs targeting of the *HOXA6-HOXA5* locus.** Primers for qPCR amplification of the indicated transcripts or genomic regions are indicated by the left/right arrows. Transcripts, *HOXA5* short and *HOXA5* long 1; regions, *HOXA5* coding region (CR) and 3' UTR. Targeted sequences of siRNAs #1 to #8 are indicated by the solid lines and the dashed lines for siRNAs #7 and #8 indicate the skipped sequences of the siRNAs.



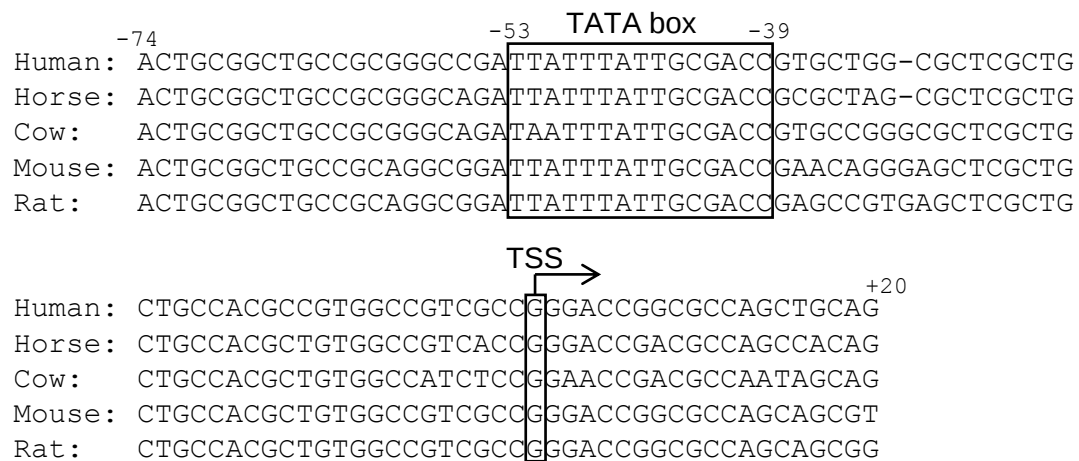
**Figure S2. 5'-rapid amplification of cDNA ends (RACE) experiments of the *HOXA6-HOXA5* locus. A.** Scheme diagram of the gene-specific primers used for 5'-RACE experiment. **B.** Electrophoretic analysis of PCR amplification products. **C.** Nucleotide sequences of the PCR products. Primers used are underlined. Grey boxes indicate the junctions between different exons. M, DNA ladder marker.



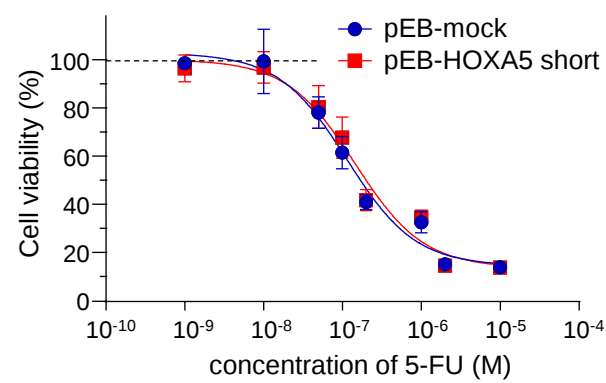
**Figure S3. 3'-rapid amplification of cDNA ends (RACE) experiments of the *HOXA6-HOXA5* locus.** **A.** Scheme diagram of the gene-specific primers used for 3'-RACE experiment. **B.** Electrophoretic analysis of PCR amplification products. **C.** Nucleotide sequences of the PCR products. Primers used are underlined. Grey boxes indicate the junctions between different exons. **M.** DNA ladder marker.



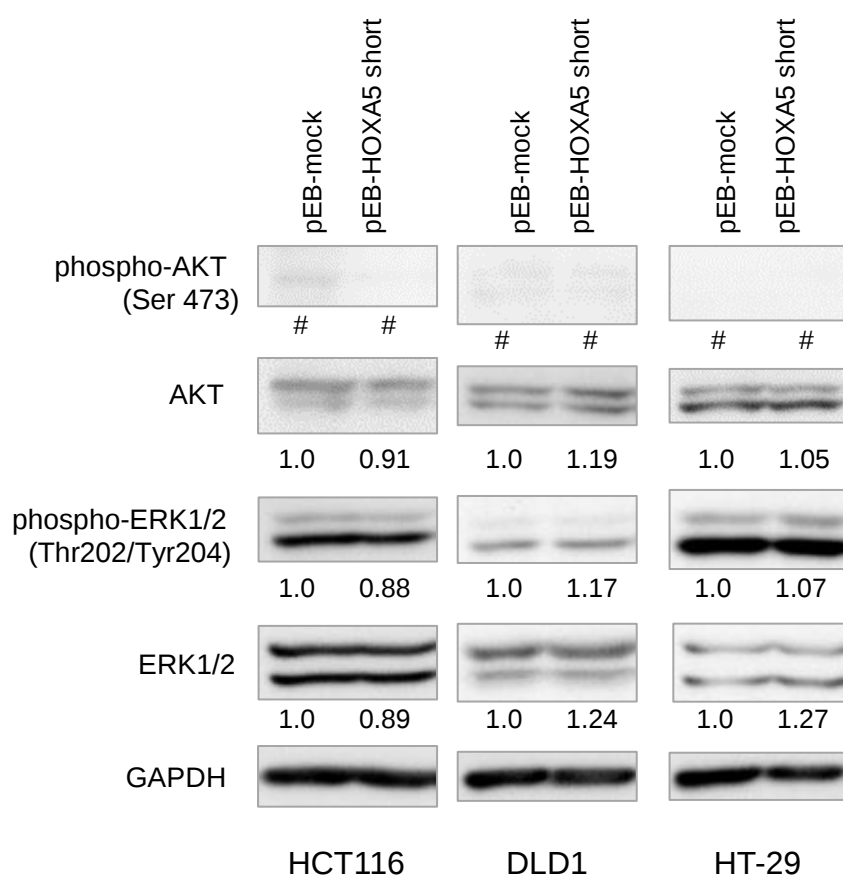
**Figure S4. Analysis of translation potency of the *HOXA5* short RNA.** **A.** A T7 promoter-containing DNA fragments encoding full-length *HOXA5* RNA, *HOXA5* short RNA, or *GAPDH* were generated by PCR amplification and the resultant PCR products were subjected to *in vitro* transcription and translation assays, which included the incorporation of fluorescent lysine. The synthesized proteins were analyzed by 15% SDS-PAGE and detected using a fluoro-imaging instrument. **B.** The translation potency of *HOXA5* short RNA was calculated using Coding-Potential Assessment Tool (CPAT) software. Sequences of the coding regions of *HOXA5* and *GAPDH* were used as translatable sequences and that of *MALAT1*, known as a functional long non-coding RNA, was used as an untranslatable sequence.



**Figure S5. Evolutionary conserved sequences of a transcriptional start site of the *HOXA5* short RNA.** Sequence alignment of the upstream sequences of a transcriptional start site (TSS) in *HOXA5* short RNA indicates the presence of a consensus TATA box and a TSS in most species.



**Figure S6. Intrinsic chemoresistance to 5-FU in HOXA5 short RNA expressing HCT116 cells.** The cell viability of pEB-HOXA5 short or pEB-mock HCT116 cells was determined by Cell Count Reagent *SF* after treatment with increasing doses of 5-FU for 48 h.



**Figure S7. Effects of *HOXA5* short RNA on AKT and ERK activation.** Protein levels of phosphorylated AKT (Ser473; #9271, Cell Signaling Tech.), total AKT (#9272, Cell Signaling Tech.), phosphorylated ERK1/2 (#9101, Cell Signaling Tech.) and total ERK1/2 (#9102, Cell Signaling Tech.) were measured by western blot analysis. GAPDH levels were used as an endogenous quantitative control. The level of phospho-AKT, phospho-ERK1/2, AKT or ERK1/2 band relative to that of GAPDH was quantitatively analyzed by densitometry. #: The band corresponding to phospho-AKT was not sufficiently detected for densitometry analyses.