

Details of the parameter estimation method

1. Experimental data used for parameter estimation:

Batch culture time-course data for WT, $\Delta pykF$ and Δpgi from [1]

- Cell concentration and extracellular glucose and acetate: Figure 2a and 2b of [1]
- Intracellular metabolite concentrations: Figure 5a of [1]
- Flux distribution data: Supplementary Table S-IV to S-VII of [1]

Batch culture time-course data for Δppc from [2]

- Cell concentration and extracellular glucose and acetate: Figure 4A of [2]

2. Methods of parameter estimation

2.1 Constrained optimization

Parameter estimation problem can be formulated as a constrained optimization problem:

$$\begin{aligned} & \text{Minimize } f(\mathbf{p}) \\ & \text{Subject to } \mathbf{g}(\mathbf{p}) \leq \mathbf{0} \end{aligned} \quad (\text{S1})$$

where $\mathbf{p} = (p_1, p_2, \dots, p_n)$ is the model parameter vector. f is the objective function which evaluates the deviation between estimated and reference parameters. $\mathbf{g} = (g_1, g_2, \dots, g_m)$ is the constraint function vector. f is given by

$$f(\mathbf{p}) = \lambda_1 \sum_{p_i \in \text{Class I}} \left| \log_{10} \frac{p_i}{p_i^*} \right| + \lambda_2 \sum_{p_i \in \text{Class II}} \left| \log_{10} \frac{p_i}{p_i^*} \right| + \lambda_3 \sum_{p_i \in \text{Class III}} \left| \log_{10} \frac{p_i}{p_i^*} \right|. \quad (\text{S2})$$

We categorized the model parameters into three classes according to reliability of their reference values. Class I includes the parameters for which measured values are available. Class II includes the parameters for which estimations are available from references. Class III includes the parameters for which no information is available. λ s are the penalty weights ($\lambda_1 = 100$, $\lambda_2 = 10$, $\lambda_3 = 0$). If parameter values are changed from reliable reference values, large penalties are imposed. $\mathbf{g}$ consists of nine constraint functions:

g_1 for fitting to experimental data (WT)

g_2 for fitting to experimental data ($\Delta pykF$)

g_3 for fitting to experimental data (Δpgi)

g_4 for fitting to experimental data (Δppc)

g_5 for reasonable molecular concentrations and fluxes (WT)

g_6 for reasonable molecular concentrations and fluxes ($\Delta pykF$)

g_7 for reasonable molecular concentrations and fluxes (Δpgi)

g_8 for reasonable molecular concentrations and fluxes (Δppc)

g_9 for reasonable activity changes of EIIA and transcription factors (WT). Detailed calculation procedures for g s are shown in the following sections. In order to solve the above constrained optimization problem, we employed a real-coded genetic algorithm (GA) with UNDX [3] as a crossover method and MGG [4] as a generation alternation method. We paralleled the GA using MPI. The GA was run on the super computer Shirokane3 provided by the Human Genome Center at The University of Tokyo.

2.2 Constraints for fitting to experimental data

g_1 - g_4 are the constraints for the deviation between model predictions and experimental data. g_1 is given by

$$g_1 = h_{1,1} + h_{1,2} + h_{1,3} - AE, \quad (S3)$$

where AE is the allowable error and set to 0.18 in this study. $h_{1,1}$ evaluates fitting to the cell concentration, extracellular glucose and acetate concentrations (Figure 2a and b of [1]):

$$h_{1,1} = \sum_{t=\{0,1,\dots,10\}} \left(\frac{X(t) - X^*(t)}{\max_{t=\{0,1,\dots,10\}} (X^*(t))} \right)^2 + \sum_{t=\{0,1,\dots,10\}} \left(\frac{GLC^{ex}(t) - GLC^{ex*}(t)}{\max_{t=\{0,1,\dots,10\}} (GLC^{ex*}(t))} \right)^2 + \sum_{t=\{0,1,\dots,10\}} \left(\frac{ACE^{ex}(t) - ACE^{ex*}(t)}{\max_{t=\{0,1,\dots,10\}} (ACE^{ex*}(t))} \right)^2, \quad (S4)$$

where t is the time (in hour). X , GLC^{ex} and ACE^{ex} show the predicted cell concentration, predicted extracellular glucose and acetate concentrations, respectively. The variables with an asterisk show experimental data.

$h_{1,2}$ evaluates fitting to the intracellular metabolite concentrations reported in Figure 5a of [1]:

$$h_{1,2}(x_i) = \sum_{t=\{4,5,6,7,8,8.5,9,10\}} \sum_{x_i \in LIST1} \left(\frac{x_i(t) - x_i^*(t)}{\max_{t=\{4,5,6,7,8,8.5,9,10\}} (x_i^*(t))} \right)^2, \quad (S5)$$

$$LIST1 = \{PYR, MAL, PEP, GAP, Ru5P, R5P, G6P, 6PG, S7P, FBP\}, \quad (S6)$$

where x_i shows the i th molecular concentration.

$h_{1,3}$ evaluates fitting to the flux distribution reported in Supplementary Table S-IV to S-VII of [1].

$$h_{1,3}(x_i) = \sum_{t=\{4,5,6,7\}} \sum_{x_i \in LIST2} \left(\frac{v_i(t)}{v_{Pis4}(t)} - \frac{v_i^*(t)}{v_{Pis4}^*(t)} \right)^2 + \sum_{t=\{8,8.5,9,10\}} \sum_{x_i \in LIST2} \left(\frac{v_i(t)}{v_{Ack}(t)} - \frac{v_i^*(t)}{v_{Ack}^*(t)} \right)^2, \quad (S7)$$

$$LIST2 = \{v_{Pis4}, v_{E,Pgi}, v_{E,Pfk}, v_{E,Fba}, v_{E,Gapdh}, v_{E,Pyk}, v_{E,Pdh}, v_{E,Ack}, v_{E,G6pdh}, v_{E,6gdh}, v_{E,Ru5p}, v_{E,R5pi}, v_{E,TkaA}, v_{E,Tal}, v_{E,TkaB}, v_{E,Cs}, v_{E,Icdh}, v_{E,akgdh}, v_{E,Sdh}, v_{E,Fum}, v_{E,Mdh}, v_{E,Ppc}, v_{E,Mez}, v_{E,Icl}, v_{E,Ms}, v_{E,Kdpg}\}, \quad (S8)$$

where v_i shows the i th flux. g_2 and g_3 are calculated in the same way as g_1 , where g_2 is for $\Delta pykF$ and g_3 for Δpgi . Since intracellular metabolite concentration and flux distribution data are lacking for Δppc , g_4 evaluates only fitting to data for cell concentration and extracellular glucose and acetate, i.e. $g_4 = h_{4,1} - AE$.

2.3 Constraints for concentrations and fluxes

All of molecular concentrations and fluxes should be within a biologically reasonable range. g_5 is employed for this propose and given by

$$g_5 = \sum_{t=\{0,0.1,\dots,12\}} \left(\sum_{x_i \in \text{Intracellular Metabolites}} h_{4,1}(x_i(t)) + \sum_{x_i \in \text{Proteins}} h_{4,2}(x_i(t)) + \sum_{\text{All } v_i} h_{4,3}(v_i(t)) \right), \quad (S9)$$

where x_i and v_i are the i th molecular concentration and the i th flux, respectively. $h_{5,1}$, $h_{5,2}$, and $h_{5,3}$ are given by

$$h_{5,1}(x_i(t)) = \begin{cases} x_i(t)^2 & (x_i(t) < 0) \\ 0 & (0 \leq x_i(t) \leq ub_{metabolite}) \\ \left(\frac{x_i(t) - ub_{metabolite}}{ub_{metabolite}} \right)^2 & (ub_{metabolite} < x_i(t)) \end{cases}, \quad (S10)$$

$$h_{5,2}(x_i(t)) = \begin{cases} x_i(t)^2 & (x_i(t) < 0) \\ 0 & (0 \leq x_i(t) \leq ub_{protein}) \\ \left(\frac{x_i(t) - ub_{protein}}{ub_{protein}} \right)^2 & (ub_{protein} < x_i(t)) \end{cases}, \quad (S11)$$

$$h_{5,3}(v_i(t)) = \begin{cases} 0 & (|v_i(t)| \leq ub_{flux}) \\ \left(\frac{|v_i(t)| - ub_{flux}}{ub_{flux}} \right)^2 & (ub_{flux} < |v_i(t)|) \end{cases}. \quad (S12)$$

$ub_{metabolite}$, $ub_{protein}$ and ub_{flux} are the upper bounds and set to 20 mM, 0.2 mM, 6000 mM/h, respectively. g_6 , g_7 ,

and g_8 are calculated in the same way as g_5 , where g_6 is for $\Delta pykF$, g_7 for Δpgi , and g_8 for Δppc .

2.4 Constraints for transcription factors

It is known that the activities of EIIA and transcription factors change in response to the carbon source shift from glucose to acetate. g_9 is employed to reproduce those experimental observations:

$$g_9 = \sum_{i \in \{EIIA, Crp, Cra, PdhR\}} h_{9,1}(Variation_i). \quad (S13)$$

$h_{9,1}$ is given by

$$h_{9,1}(Variation_i) = \begin{cases} \left(\frac{Variation_i - lb_{Variation}}{lb_{Variation}} \right)^2 & (Variation_i < lb_{Variation}) \\ 0 & (lb_{Variation} \leq Variation_i) \end{cases}, \quad (S14)$$

where $lb_{Variation}$ is the lower bound and set to 0.5. $Variation_i$ is given by

$$Variation_{EIIA} = \max_{t=\{0,0.1,\dots,12\}} \left(\frac{EIAP(t)}{EIAP(t) + EI(A)(t)} \right) - \min_{t=\{0,0.1,\dots,12\}} \left(\frac{EIAP(t)}{EIAP(t) + EI(A)(t)} \right), \quad (S15)$$

$$Variation_{Crp} = \max_{t=\{0,0.1,\dots,12\}} \left(\frac{CrpAMP(t)}{CrpAMP(t) + Crp(t)} \right) - \min_{t=\{0,0.1,\dots,12\}} \left(\frac{CrpAMP(t)}{CrpAMP(t) + Crp(t)} \right), \quad (S16)$$

$$\begin{aligned}
Variation_{Cra} = & \max_{t=\{0,0.1,\dots,12\}} \left(\frac{CraFBP(t)}{CraFBP(t) + Cra(t)} \right) \\
& - \min_{t=\{0,0.1,\dots,12\}} \left(\frac{CraFBP(t)}{CraFBP(t) + Cra(t)} \right),
\end{aligned} \tag{S17}$$

$$\begin{aligned}
Variation_{PdhR} = & \max_{t=\{0,0.1,\dots,12\}} \left(\frac{PdhRPYR(t)}{PdhRPYR(t) + PdhR(t)} \right) \\
& - \min_{t=\{0,0.1,\dots,12\}} \left(\frac{PdhRPYR(t)}{PdhRPYR(t) + PdhR(t)} \right).
\end{aligned} \tag{S18}$$

References

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