

Synthesis of (-)-deoxypodophyllotoxin and (-)-epipodophyllotoxin via a multi-enzyme cascade in *E. coli*

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1. Supplementary Material

***omt1* (GenBank [KT390155.1](#)), native DNA-sequence**

ATGGATACTAGGGCTGATGCTGAGATTAAAGCAATGGAGCTGATTGGTATTGGAGTACTTCCACTGGCAATGA
AGGCGATAATTGAGCTCAATGTGTTAGAGATCCTATCAAAAGCAGGACCAGATACCCAACTCACTGCTGCTCA
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CCCTTTGAGTATTCTGTCTCCGATACAAGGTTTTAATAAGGTTTTTAATGCTGGCATGTTTTGACCATTCTACTC
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GTGGTTGCCGACGCGCCTTCTTTTCCCTGGTGTGAAGCACATTGGTGGTGATATGTTTGAGAGTGTTCCCTCTG
GTGATGCAATTTTTCATGAAGTGGATACTTCATGACTGGGACGATGGACGTTGCCTGACTTTGCTGAAGAATTG
TTGGAATGCATTGCCAGAGCATGGAAAGGTGATAATAGTGGAGTGGATTCTACCATCAGATGCAGCGACTGAC
CCAACATCTCGCCGTGTCTTTCACAGCTGATTTGATGATGTTGGCTTTCAGCGAAGGGGGTAAAGAGCGAACCT
TGGGTGACTACGGAGCACTTGCAAAGGAAGCTGGTTTTACCACTGTCAAAGATTTCCCTTGCGCAAATGGCAT
TTCAGTCATTGAGTTCCATAAGAAGTGA

***omt3* (GenBank [KT390157.1](#)), native DNA-sequence**

ATGGAAATGGCTCCAACAATGGATTTAGAGATAAGAAATGGAAATGGTTATGGTGATTCTGGAGAGGAGCTTC
TAGCAGCACAAGCTCACATATACAACCACATATTCAACTTCATAAGCTCGATGGCACTGAAATGTGCAGTGGA
GTTAAACATAACCAGAAATTCTCCACAACCATCAACCCAAAGCGGTTACTCTCTCTGAACTAGTACAGGCCCTT
CAAATCCCCCAAGCAAAATCCGCGTGTCTGTATCGCCTGTTGCGAATACTAGTCCATTCTGGCTTCTTTGCCA
TAACGAAAATACAAAGCGAGGGAGATGAAGAGGGTTATTTACCAACCCTTTCTCTAAACTACTACTGAAAAA
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TCTATCATTGAAGTGTTCCCTAA

***cyp71cu1* (GenBank [KT390172.1](#)), native DNA-sequence (*cyp71cu1nat_wt*)**

N-terminally truncated part of the DNA-sequence (*CYP71CU1Δ20nat*) is highlighted in bold and underlined

ATG**GAAACATTTTCAGTGCCTCACACTTTTCCTTCTCTTCATCTCCACTGTTTTTCATCCTCAAG**AGAAAATTCT
CTCATAAACCAAACCTACCACCATCACCACCAAAGCTACCAATCCTAGGCAACTTTCATCAGCTAGGCACACT
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CCCTAACAACTGCTGCCAGGGCACTACTTTACGACTGCACAGATATATCTTTTGCACCTTATGGTGAGTACTG
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GAGGAATTGGCTTCTGATATGATAAAGGAGATTACTCGTTTGTCCAAACTGGAGCACCTGTGGATGTAACATA
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GTTTCAGAAGCTGTCCAGGGATTTTTTGGATCTTGTAGGAGCTTTTTGTTTTAATGATTTCTTCCAGGGATG
GCATGGATGGATGCTCTCACTGGGCTAAACAGGAAGCTGAAGAAGGGTTCAAGAGAATTAGATGACTTTGTTG
ATGAACTCATAGAAGAACGCATTGCTATGGTAAAAGATGGCGTTGAGCCAAATGAATTTCTGGATCTTCTACT
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CAACTCCTGAAACACTTGATATGACTGAGGACTATTGTTTTGCCCTCTTTAAGAAAAAACCTTTCATTTTAT
TCCTATTTCTCGATCCTCTTGA

***cyp71cu1*, *E. coli* codon optimized DNA-sequence (*cyp71cu1oc_wt*)**

N-terminally truncated part of the DNA-sequence (*cyp71cu1Δ20oc*) is highlighted in bold and underlined

ATG**GAAACCTTCCAGTGCCTGACCCTGTTCCCTGCTGTTTCATCTCTACCGTTTTTCATCCTGAAA**CGTAAATTCT
CTCACAAACCGAACCTGCCGCCGTCTCCGCCGAAACTGCCGATCCTGGGTAACTTCCACCAGCTGGGTACCCT
GGTTCACCGTGCTGTTACCGTTCTGGCTGCTAAATACGGTCCGCTGATGCTGCTGCACTTCGGTAAACCCCG
GTTCTGATCGTTTCTTCTCAGGAAACCGCTAAAGAAATCATGAAAACCCACGACCTGGCTCTGGCTAACCGTC
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GCGTGAAATGAAAAAATGGCTGTTCTGAACCTGCTGTCTATCAAAAAAATCCAGTCCTTCCGTTCTGTTTCGT
GAAGAACTGGCTTCTGACATGATCAAAGAAATCACCCGTCTGTCTAAAACCGGTGCTCCGGTTGACGTTACCA
ACATGCTGTACCACCTTCTCTGAAGACCTGCTGTTCCGTTGCACCCCTGGGTTTCAAACCGAAAGGTCAGCACAA
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TGAATCTTACACCTCTATCAACGTTGAAAACCTACATCATCCCGCCGTACACCTCTGTTATGATCAACATCTGG
CACATCCAGCGTGACCCGAAACTGTGGGACAAAGCTGAAGAATTTATCCCGGAACGTTTCATGAACTCTGGTA
TCGACTACAAATCTCACGACTACGAATTTATCCCGTTCGGTTCTGGTCGTCGTGGTTGCCCGGGTATGTCTTT
CGGTGTTGCTGCTGTTGAATTTGCTGTTGCTAACCTGCTGTACTGGTTGCACTGGAAATTCGTTGGTGACACC
ACCCCGGAAACCTGGACATGACCGAAGACTACTGCTTCGCTCTGTTCAAAAAAAACCGCTGCACCTTCATCC
CGATCTCTCGTTCTTCTTAA

2-odd (GenBank KT390173.12), native DNA-sequence

ATGGGTTCTACAGCACCCCTAAGGCTTCCAGTTATAGATTTATCCATGAAGAACTTGAAGCCTGGAACAACTT
CTTGGAACTCGGTACGCACCCAGGTACGGGAGGCACTGGAAGAATACGGTTGCTTTGAAGCTGTGATCGATGC
TGTGTCTCCAGAGCTGCAGAAGGCAGTATGTAACAAAGGACACGAGCTGCTTAATCTTCCATTAGAAACCAAG
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GCAGAATAACAGATTTGGAAAAAGTTGAGAGGTTCACTAATCTTATGTGGCCCGAGGGGAATAAGGATTTCTG
TGAAACTGTGTATTCTTATGGCAAACGAATGGCGGAGGTGGACCACATATTGAAAATGATGGTTTTCGAGAGT
TTTGGAATGGAGAAGCACTTCGACTCGTTCTGCGAATCAACAAATTACCTTCTCCATTTTCATGAGATACCAAC
AACCAGGGAAGGATGGACGTTACCTGCTCTTTCGTTGCATAAGGACAAGAGCATCTTGACCATAGTAAACCA
AAATGATGTCAAGGGATTGGAATTTGAAACCAAGGATGGAGAATGGATTTTACCTACAGCTGACAACCATATT
GTTCTTCTAGGAGACTGCTTCATGGCATGGAGCAATGGTAGATTACATAGTCCTCTTCACCGGGTCACGTTGG
TCGCGAACCAGGCGAGGTTATCTACATCATCGTTTTTCGTTTCCAAAGGACATAATAGAGACCCCTGCAGAGCT
GGTGGATGAAGAGCATCCTTTGCTATTTAATCCCTTTGAGATAACGGAGTTGCTTGCTTACTGTTTCACAAAA
GAGGGTGCAAAGGCGGTGTGTGACCTCAAGCAATACAAGGCGTACACAGGTGCATGA

***cyp82d61*, *E. coli* codon optimized DNA-sequence (*cyp82d61oc_wt*)**

N-terminally truncated part of the DNA-sequence (*cyp82d61*Δ23oc) is highlighted in bold and underlined; for native DNA-sequence of *cyp82d61* gene see GenBank KC110995.1

ATG**GACTCTCTGCACTGCCTGGAAACCCCTGCTGCTGGGTTTCTTCGTTCTGCTGCCGTGCTTCTTCTACTTCG**
TTTGGAACCAACGAACAACAAATCAAAGAACCGCCGAGCCGGCTGGTGCTTGCCGATCATCGGTCACCT
GCACCTGCTGGCTCGTGGTGACCTGCCGCACAAATCCTGTCTTCTTTCGCTGACAAAACGGTCCGGTTTTTC
AAAATCCAGCTGGGTGTTTACCAGGCTCTGGTTGTTAACAACCTCTGAAATCGCTAAAGAATGCTTACCACCA
ACGACCGTTTCTTCTGAACCGTCCGTCTGGTGTGCTGCTAAAATCATGGGTTACAACCTACGTTATGCTGGG
TGTTGCTCCGTACGGTCCGTACTGGCGTGACATGCGTAAAATCATCATGCTGGAATTCCTGTCTAACCGTCTG
CTGCAGTCTCTGAAACACGTTTGGCACTCTGAAATCTCTATCTCTTCTAAAGAACTGTACAACTGTGGGAAA
CCCAGAACATCGACTTCTGCCTGGTTGACATGAAACAGTGGCTGGCTGACCTGACCCTGAACATGTCTGTAA
AATGGTTGTTGGTAAACGTTTCTTCGGTTCTGCTTCTGCTTCTGCTTGCGAAGAAACCGAATCTTCTAACTGC
CCGAAAACCCTGCGTAACATGTTCCGTCTGATGGGTTCTTTCGTTCTGTCTGACTACCTGCCGTACCTGCGTT
GGCTGGACCTGGGTGGTCACGAAAAAGAAATGAAACGTACCGTTAAAGAACTGGACATCCTGTTCAAAGGTTG
GCTGGACGAACACAAACGTAAACGTCTGTCTGGTGGTAAAGAAGACGACGACCAGGACTTCATGGACGTTATG
CTGTCTATCCTGGAAGAATCTAAACTGGGTAACGACGTTGACACCATCAACAAAACCGCTTGCTGGCTATCA
TCCTGGGTGGTGCTGACACCACCTGGGCTACCCTGACCTGGGCTCTGTCTCTGCTGCTGAACAACCCGAACGC
TCTGAAAAAAGCTCAGGACGAACCTGGACCTGCACGTTGGTCGTGACCGTAACGTTGACGAATCTGACCTGGTT
AAACTGACCTACATCGACGCTATCATCAAAGAAACCCCTGCGTCTGTACCCGCCGGGTCCGCTGCTGGGTCCGC
GTGTTGTTACCGAAGACTGCACCATCGCTGGTTACCACGTTTCGTGCTGGTACCCGTCTGATCGTTAACGCTTG
GAAAATCCAGCGTGACCCGCTGGTTTGGTCTCAGCCGCACGAATACCAGCCGGAACGTTTCTGGAACGTGAC
GTTGACATGAAAGGTCAGCACTTCGAACTGATCCCGTTTCGGTTCTGGTCGTGCTTGCCCGGCTATCTCTC
TGGCTCTGCAGGTTCTGCCGCTGACCTGGCTCACATCCTGCACGGTTTCGAACTGCGTACCCCGAACAGAA
CAAAGTTGACATGACCGAAACCCCGGGTATCGTTTACGCTAAAGCTACCCCGCTGGAAGTTCTGGTTGCTCCG
CGTATCTCTCCGAAATGCTTCGTTTAA

2. Supplementary Methods

Expressions of OMT3, OMT1 and 2-ODD

Expressions were evaluated in *E. coli* BL21 (DE3), C41(DE3) and C43 (DE3). Protein expression was induced at $OD_{600} = 0.6$ by the addition of 0.5 mM IPTG (additional supplementation with 0.1 mM $FeSO_4$ in the case of 2-ODD), followed by incubation at 25°C, 120 rpm for 20 h.

Expressions of wildtype and N-terminal truncated CYP71CU1 and CYP82D61 variants

Expressions were evaluated in *E. coli* C41(DE3). Protein expression was induced at $OD_{600} = 0.6$ by the addition of 0.5 mM IPTG, with additional supplementation with 0.5 mM 5-aminolevulinic acid (5-ALA) and 0.1 mM $FeSO_4$. Incubations were carried out at 120 rpm for 20 h or 48 h, with temperatures of 20°C, 25°C and 30°C. Cells were harvested by centrifugation (3,220 x g, 4°C, 20 min) and the cell pellet was resuspended in 50 mM potassium phosphate buffer, pH 7.5, 100 μ M PMSF. Cells were disrupted by sonication, the cell debris was sedimented by centrifugation (12,300 x g, 4°C, 30 min) and separated from the soluble protein fraction.

P450 concentrations within the soluble protein fractions were determined by recording CO difference spectra according to the method of Omura and Sato [1] to allow comparison between wildtype and truncated versions of CYP71CU1 and CYP82D61.

The results of the screening and optimized expression conditions are summarized in Tables S8, S9, and S10 within this appendix.

Table S1. Summary of generated *cyp71cu1* and *cyp82d61* gene variants.

Name of variant	DNA-sequence	Inserted modification (second amino acid)	No. of N-terminal truncated amino acids
<i>cyp71cu1Δ20nat</i>	Native	Ala	1 - 20 (Δ20)
<i>cyp71cu1Δ20oc</i>	<i>E. coli</i> codon optimized	Ala	1 - 20 (Δ20)
<i>cyp82d61Δ23oc</i>	<i>E. coli</i> codon optimized	Ala	1 - 23 (Δ23)

Table S2. Summary of genes used for establishment of multi-enzyme cascade reactions.

Gene	Complete name or purpose	Organism	Reference
<i>cyp719aΔ23oc</i>	(-)-pluviatolide synthase (truncated, codon optimized)	<i>Sinopodophyllum hexandrum</i>	Decembrino <i>et al.</i> , 2020 [2]
<i>atr2</i>	NADPH-cytochrome P450 reductase 2 (ATR2)	<i>Arabidopsis thaliana</i>	Kranz-Finger <i>et al.</i> , 2018 [3]
<i>omt3</i>	(-)-pluviatolide-O- methyltransferase	<i>Sinopodophyllum hexandrum</i>	This work
<i>cyp71cu1Δ20nat</i>	(-)-5'-desmetoxy-yatein hydroxylase (truncated, native DNA-sequence)	<i>Sinopodophyllum hexandrum</i>	This work
<i>omt1</i>	(-)-5'-desmethyl-yatein O- methyltransferase	<i>Sinopodophyllum hexandrum</i>	This work
<i>2-odd</i>	(-)-deoxypodophyllotoxin synthase	<i>Sinopodophyllum hexandrum</i>	This work
<i>cyp82d61Δ23oc</i>	Putative hydroxylation (truncated, codon optimized)	<i>Sinopodophyllum hexandrum</i>	This work

Table S3. Summary of plasmids used within this study.

Plasmid	Features	Copy number	Reference
pETDuet_atr2_cyp719aΔ23oc	ColE1 ori, P _{T7} , <i>lacI</i> , Amp ^R	~ 40	Decembrino <i>et al.</i> , 2020 [2]
pCDFDuet_omt3_cyp71cu1Δ20nat	CloDf13 ori, P _{T7} , <i>lac</i> , Sm ^R	20 - 40	This work
pCDFDuet_omt3	CloDf13 ori, P _{T7} , <i>lac</i> , Sm ^R	20 - 40	This work
pCDFDuet_cyp71cu1Δ20nat	CloDf13 ori, P _{T7} , <i>lac</i> , Sm ^R	20 - 40	This work
pCDFDuet_cyp71cu1Δ20oc	CloDf13 ori, P _{T7} , <i>lac</i> , Sm ^R	20 - 40	This work
pCDFDuet_cyp71cu1nat_wt	CloDf13 ori, P _{T7} , <i>lac</i> , Sm ^R	20 - 40	This work
pCDFDuet_cyp71cu1oc_wt	CloDf13 ori, P _{T7} , <i>lac</i> , Sm ^R	20 - 40	This work
pCOLADuet_2-odd_omt1	ColA ori, P _{T7} , <i>lac</i> , Kan ^R	20 - 40	This work
pCDFDuet_omt1	CloDf13 ori, P _{T7} , <i>lac</i> , Sm ^R	20 - 40	This work
pCDFDuet_2-odd	CloDf13 ori, P _{T7} , <i>lac</i> , Sm ^R	20 - 40	This work
pETDuet_atr2_cyp82d61Δ23oc	ColE1 ori, P _{T7} , <i>lacI</i> , Amp ^R	~ 40	This work
pETDuet_cyp82d61oc_wt	ColE1 ori, P _{T7} , <i>lacI</i> , Amp ^R	~ 40	This work
pETDuet_cyp82d61Δ23oc	ColE1 ori, P _{T7} , <i>lacI</i> , Amp ^R	~ 40	This work

Table S4. Oligonucleotides used for generation of *cyp71cu1* and *cyp82d61* gene variants.

Name of variant	Oligonucleotides 5' → 3'
<i>cyp71cu1Δ20nat</i>	forward: CAGAAAATTCTCTCATAAACCAAAC reverse: GCCATATGTATATCTCCTTCTTTATAC
<i>cyp71cu1Δ20oc</i>	forward: CCGTAAATTCTCTCACAAAC reverse: GCCATATGTATATCTCCTTCTTATAC
<i>cyp82d61Δ23oc</i>	forward: CGTTTGGAAAAAACCGAAC reverse: GCCATATGTATATCTCCTTCTTATAC

Table S5. LC/MS methods used within this study. Gradients were made of methanol (solvent A) and ddH₂O + 0.1% formic acid (solvent B). A flow rate of 0.5 mL/min was employed with methods 1, 2 and 3, whereas 0.8 mL/min was used in case of method 4.

	Time [min]	Solvent B [%]
Method 1	0.01	57
	8.00	28
	13.00	0
	14.01	57
	20.00	57
	Time [min]	Solvent B [%]
Method 2:	0.01	80
	5.00	65
	10.00	65
	20.00	38
	25.00	0
	26.01	80
	35.00	80
	Time [min]	Solvent B [%]
Method 3	0.01	65
	5.00	50
	8.00	45
	22.00	38
	25.00	0
	26.01	80
	35.00	80
	Time [min]	Solvent B [%]
Method 4	0.01	80
	2.00	60
	5.00	55
	25.00	52
	32.50	0
	35.01	80
	40.00	80

Qualitative analysis

Table S6. Authentic reference compounds used for LC/MS identification of reaction products. Commercially not available compounds were identified via their characteristic m/z fragments as described elsewhere [4].

Compound	Manufacturer or reference	Purity	Molecular weight	RT [min] (method 4)	Characteristic m/z fragments
(-)-matairesinol 1	Phytolab	≥99% (HPLC)	358.38	9.0	359 [M+H] ⁺ 341 [M+H-H ₂ O] ⁺
(-)-pluviatolide 2	<i>Decembrino et al. 2020</i> (isolated in-house) [2]	≥95% (HPLC)	360.61	15.0	357 [M+H] ⁺ 339 [M+H-H ₂ O] ⁺
(-)-yatein 5	BioBioPha	≥97% (HPLC)	400.40	20.6	401 [M+H] ⁺ 383 [M+H-H ₂ O] ⁺
(-)-deoxypodophyllotoxin 6	Toronto Research Chemical	≥98% (HPLC)	398.40	19.8	399 [M+H] ⁺ 421 [M+Na] ⁺
(-)-5'-desmethoxy-yatein 3	Commercially not available [4]	-	370.40	21.0	<u>371 [M+H]⁺</u> <u>353 [M+H-H₂O]⁺</u>
(-)-5'-desmethyl-yatein 4	Commercially not available [4]	-	386.40	14.8	<u>387 [M+H]⁺</u> <u>369 [M+H-H₂O]⁺</u>
(-)-epipodophyllotoxin 7	Commercially not available [4]	-	414.41	10.4	<u>415 [M+H]⁺</u> <u>437 [M+Na]⁺</u>
(+)-sesamin (internal standard; IS)	TCI	>98% (GC)	354.35	31.4	337 [M+H-H ₂ O] ⁺

Underlined m/z fragments correspond to those referred by Lau and Sattely [4].

Quantitative analysis

Table S7. For quantitative analysis, substrate conversion values were calculated by the product distribution resulting from the MS-peak areas of reaction intermediates/products and substrate. Estimation of the concentration of the final product (-)-deoxypodophyllotoxin **6** was done via internal calibration in the range of 10 - 300 μM utilizing 200 μM (+)-sesamin as IS.

Conversion [%] Normalized to IS	$= 1 - (S_{\text{sample}} / IS_{\text{sample}}) / (S_{\text{control}} / IS_{\text{control}})$
Conversion [%]	$= \Sigma(P_{\text{area}}) / \Sigma(S_{\text{area}} + P_{\text{area}}) * 100$
Product distribution [%]	$= P_{\text{area}} / \Sigma(S_{\text{area}} + P_{\text{area}}) * 100$

IS: internal standard; S: substrate; P = product.

3. Supplementary Results

Expressions of OMT3, OMT1, 2-ODD and CYP71CU1

OMT3 – 41 kDa

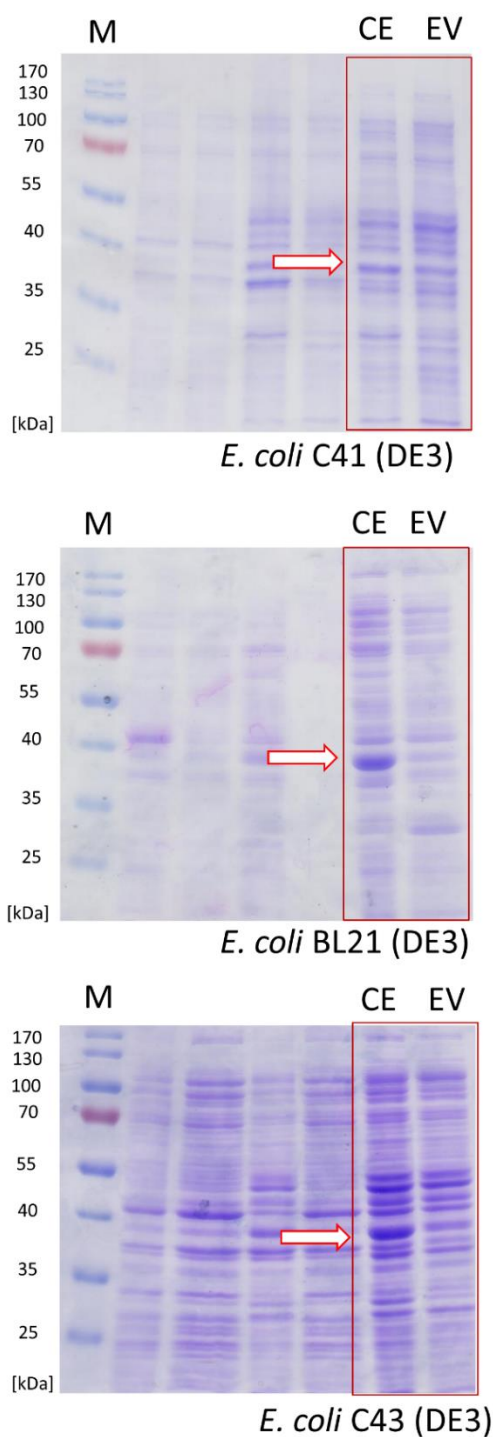


Figure S1. 12.5% SDS-gel of OMT3 (41 kDa; marked by red arrow) expression in various *E. coli* strains. M: Molecular weight size marker, EV: Vector control without *omt3* gene, CE: Soluble protein fraction after cell disruption.

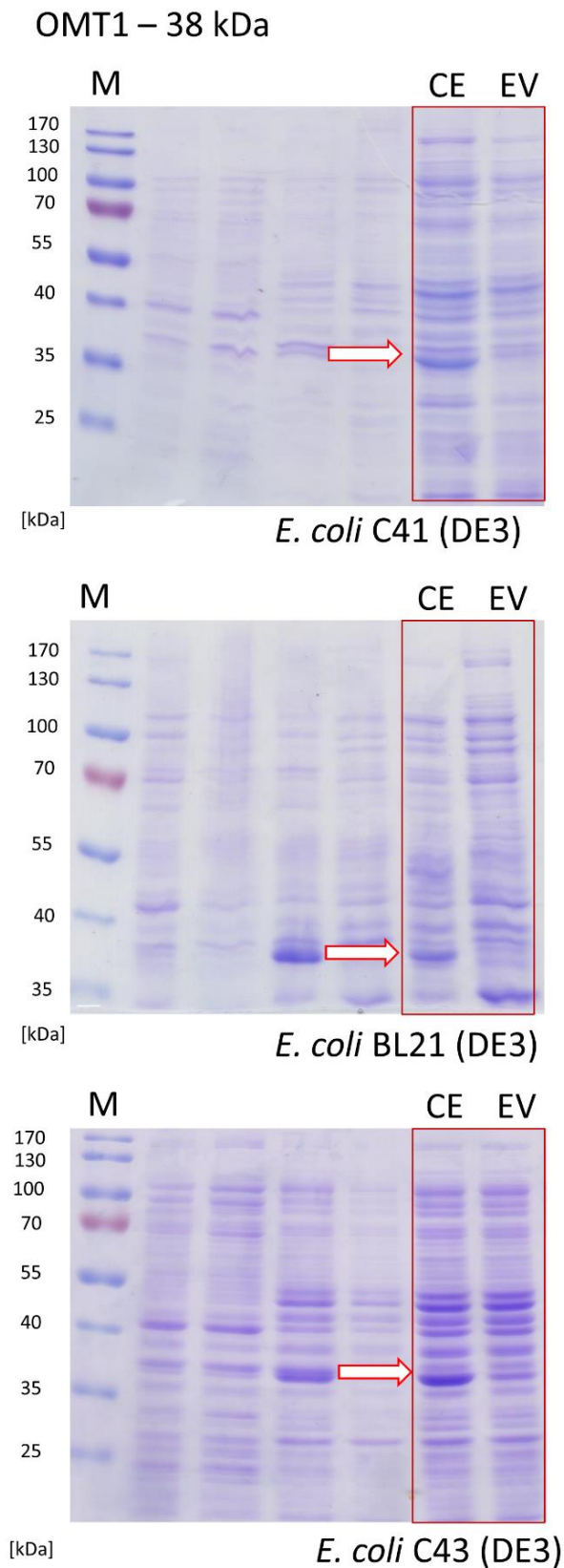


Figure S2. 12.5% SDS-gel of OMT1 (38 kDa; marked by red arrow) expression in various *E. coli* strains. M: Molecular weight size marker, EV: Vector control without *omt1* gene, CE: Soluble protein fraction after cell disruption.

2-ODD – 35 kDa

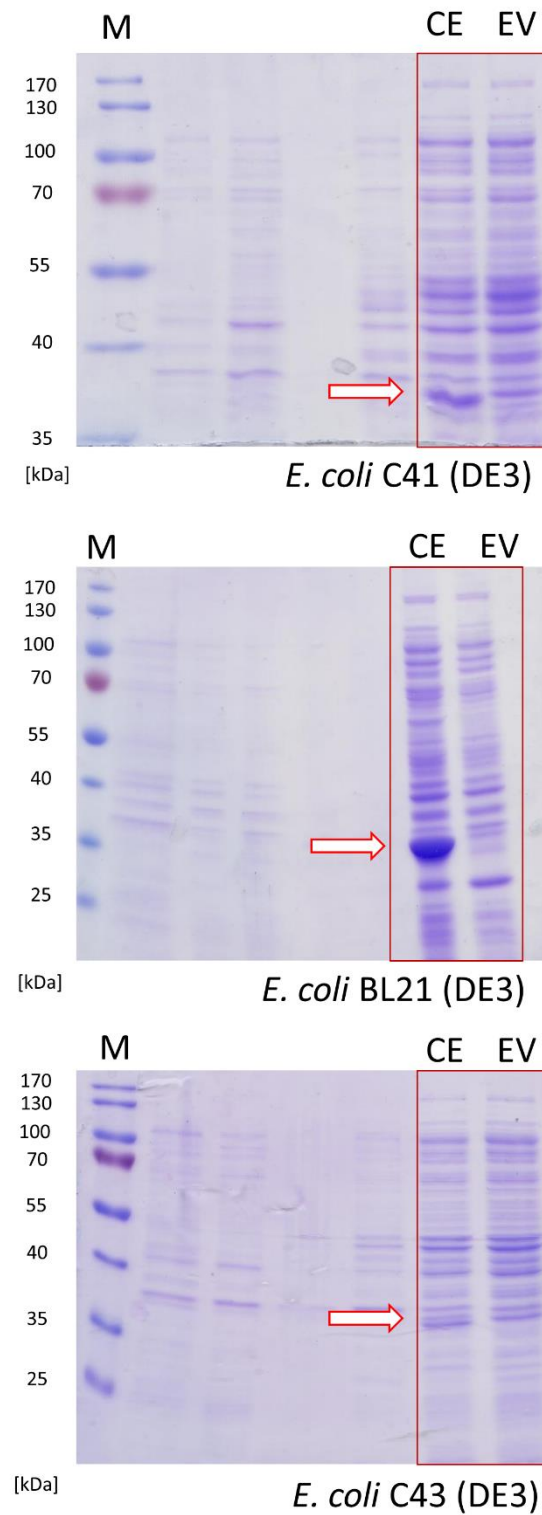


Figure S3. 12.5% SDS-gel of 2-ODD (35 kDa; marked by red arrow) expression in various *E. coli* strains. M: Molecular weight size marker, EV: Vector control without 2-odd gene, CE: Soluble protein fraction after cell disruption.

CYP71CU1 WT – 56 kDa

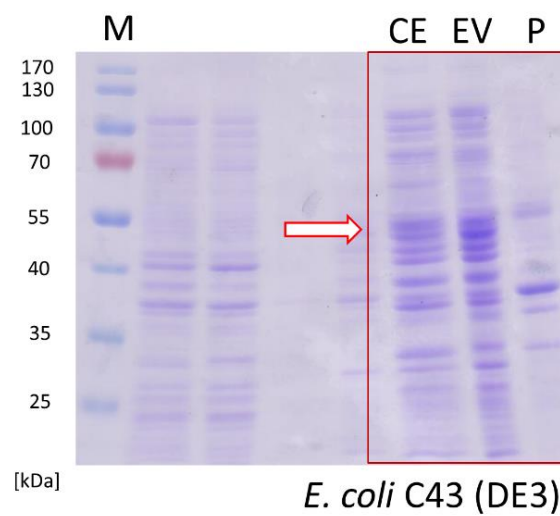
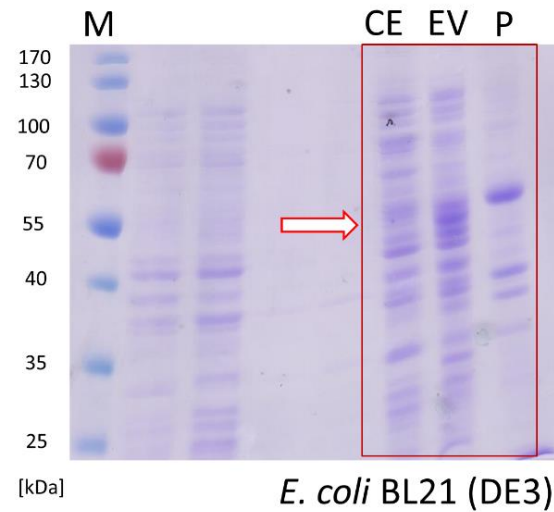
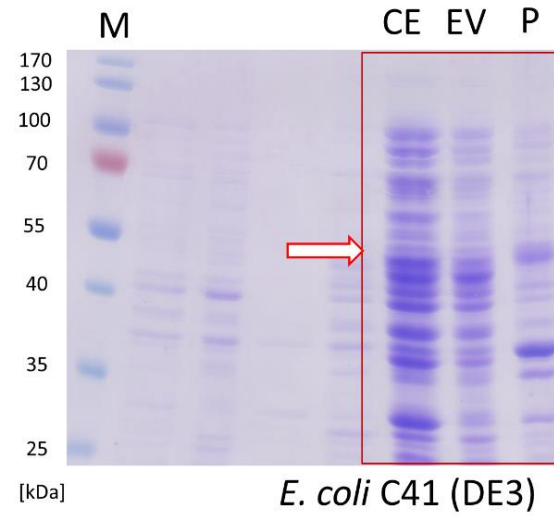


Figure S4. 12.5% SDS-gel of CYP71CU1 (56 kDa; marked by red arrow) expression in various *E. coli* strains. M: Molecular weight size marker, EV: Vector control without *cyp71cu1* gene, CE: Soluble protein fraction after cell disruption, P: Insoluble protein fraction after cell disruption.

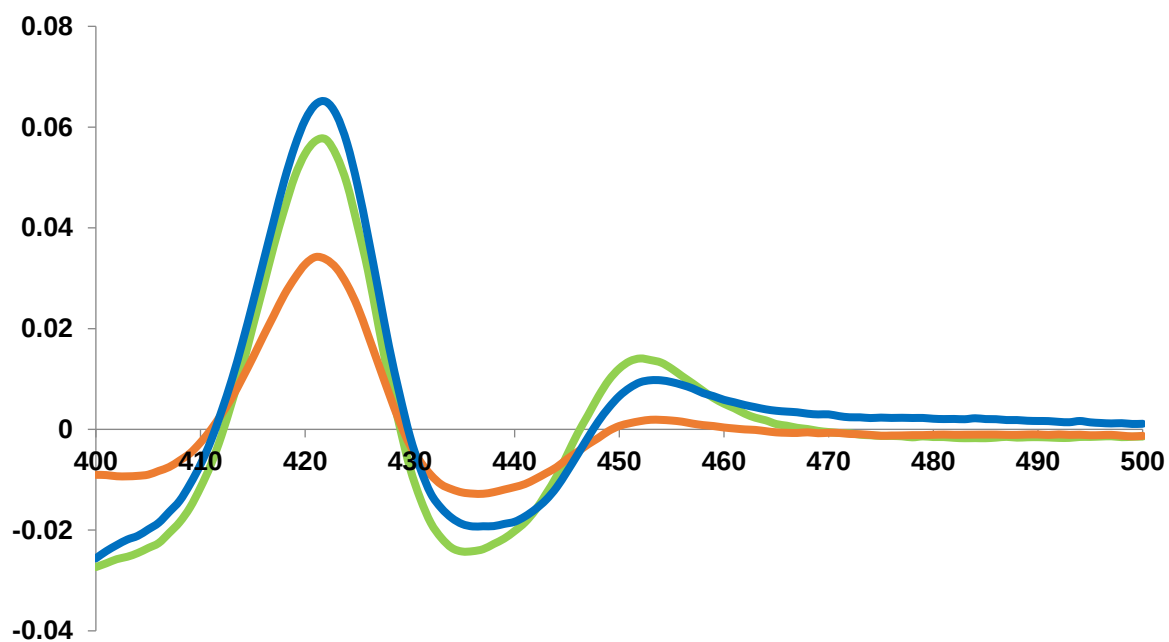


Figure S5. CO-difference spectra of soluble CYP71CU1OC_WT in *E. coli* BL21 (DE3) (orange line), C41 (DE3) (blue line), and C43 (DE3) (green line) after 24 h expression.

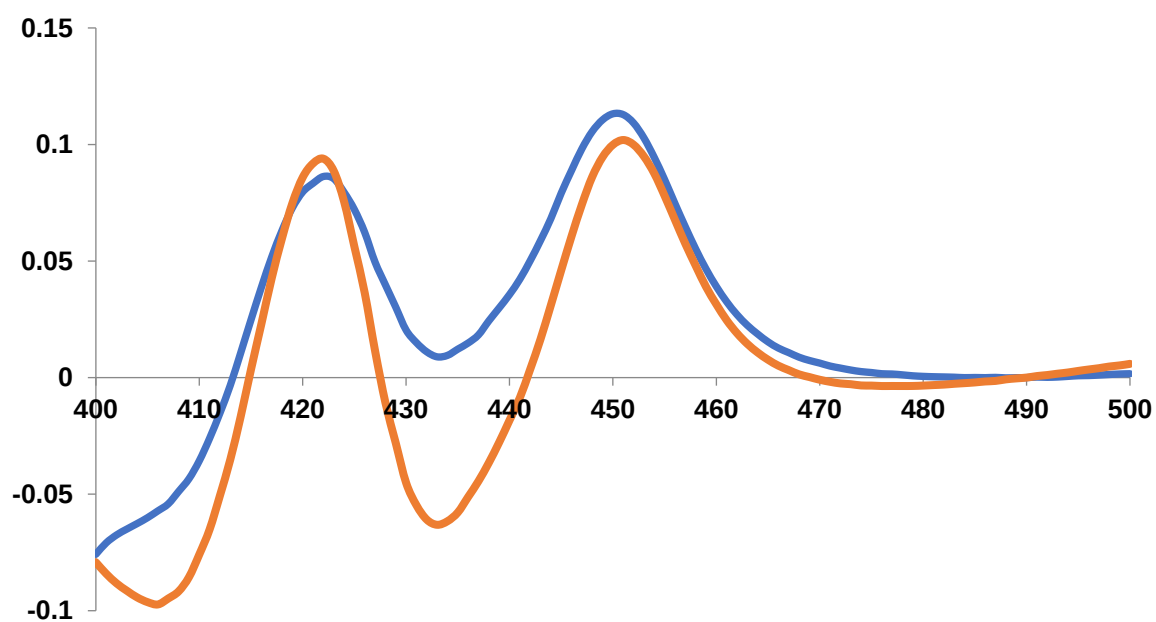


Figure S6. CO-difference spectra of soluble CYP71CU1nat_WT (blue line) and CYP71CU1OC_WT (orange line) after 48 h expression.

Table S8. Expression levels of CYP71CU1 variants in *E. coli* C41(DE3) at 25°C after 48 h.

Variant	Inserted modification	Truncated amino acids	P450 concentration	
			[$\mu\text{g/g}_{\text{cww}}$]	[$\text{mg/l}_{\text{culture volume}}$]
CYP71CU1nat_WT	-	-	263 $\pm$ 16	8 $\pm$ 5
CYP71CU1 Δ 20nat	Ala	1 - 20	813 $\pm$ 163	26 $\pm$ 6
CYP71CU1oc_WT	-	-	159 $\pm$ 16	6 $\pm$ 1
CYP71CU1 Δ 20oc	Ala	1 - 20	465 $\pm$ 138	15 $\pm$ 4

Table S9. Expression levels of N-terminally truncated CYP71CU1 variants in *E. coli* C41(DE3) at various incubation temperatures after 48 h.

Temperature	P450 concentration			
	[$\mu\text{g/g}_{\text{cww}}$]		[$\text{mg/l}_{\text{culture volume}}$]	
	CYP71CU1 Δ 20oc	CYP71CU1 Δ 20nat	CYP71CU1 Δ 20oc	CYP71CU1 Δ 20nat
20°C	282 $\pm$ 26	566 $\pm$ 39	11 $\pm$ 1	20 $\pm$ 3
25°C	465 $\pm$ 138	813 $\pm$ 163	15 $\pm$ 4	26 $\pm$ 6
30°C	112 $\pm$ 18	253 $\pm$ 108	6 $\pm$ 3	8 $\pm$ 3

Table S10. Expression levels of N-terminally truncated CYP82D61 variants in *E. coli* C41(DE3) at various incubation temperatures after 48 h.

Temperature	P450 concentration			
	[$\mu\text{g/g}_{\text{cww}}$]		[$\text{mg/l}_{\text{culture volume}}$]	
	CYP82D61oc_WT	CYP82D61 Δ 23oc	CYP82D61oc_WT	CYP82D61 Δ 23oc
20°C	n.d.	20 $\pm$ 17	n.d.	0.7 $\pm$ 0.6
25°C	n.d.	936 $\pm$ 34	n.d.	33 $\pm$ 2
30°C	250 $\pm$ 14	1395 $\pm$ 220	8 $\pm$ 1	42 $\pm$ 9

n.d.: Expression not detectable

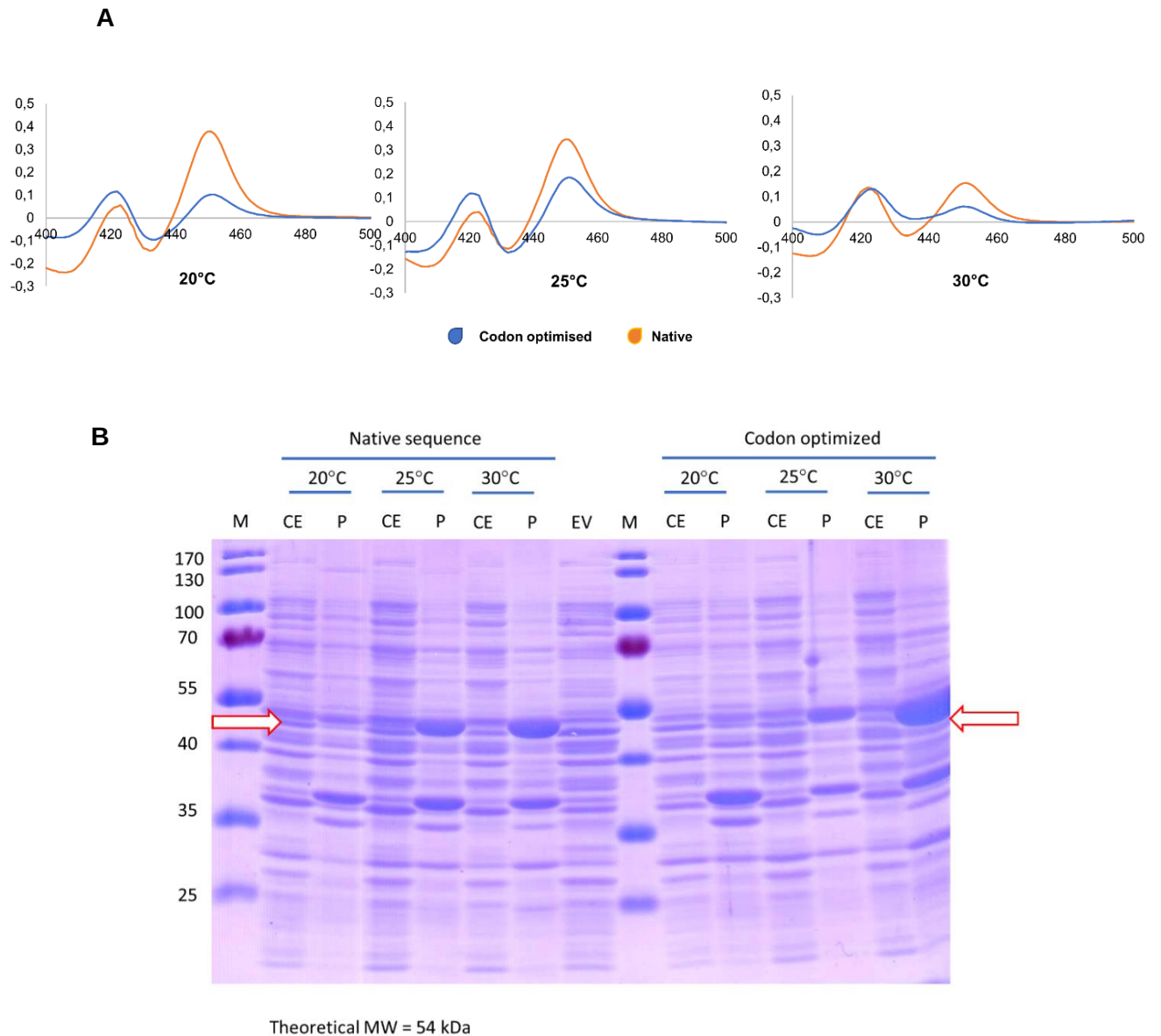


Figure S7. Expression analysis of CYP71CU1 variants incubated at different temperatures. **(A)** CO-difference spectra of CYP71CU1 Δ 20nat (orange line) and CYP71CU1 Δ 20oc (blue line). **(B)** 12.5% SDS-gel of CYP71CU1 Δ 20nat and CYP71CU1 Δ 20oc expressions (54 kDa; marked with a red arrow). M: Molecular weight size marker, EV: Vector control without *cyp71cu1* gene, CE: Soluble protein fraction after cell disruption, P: Insoluble protein fraction after cell disruption.

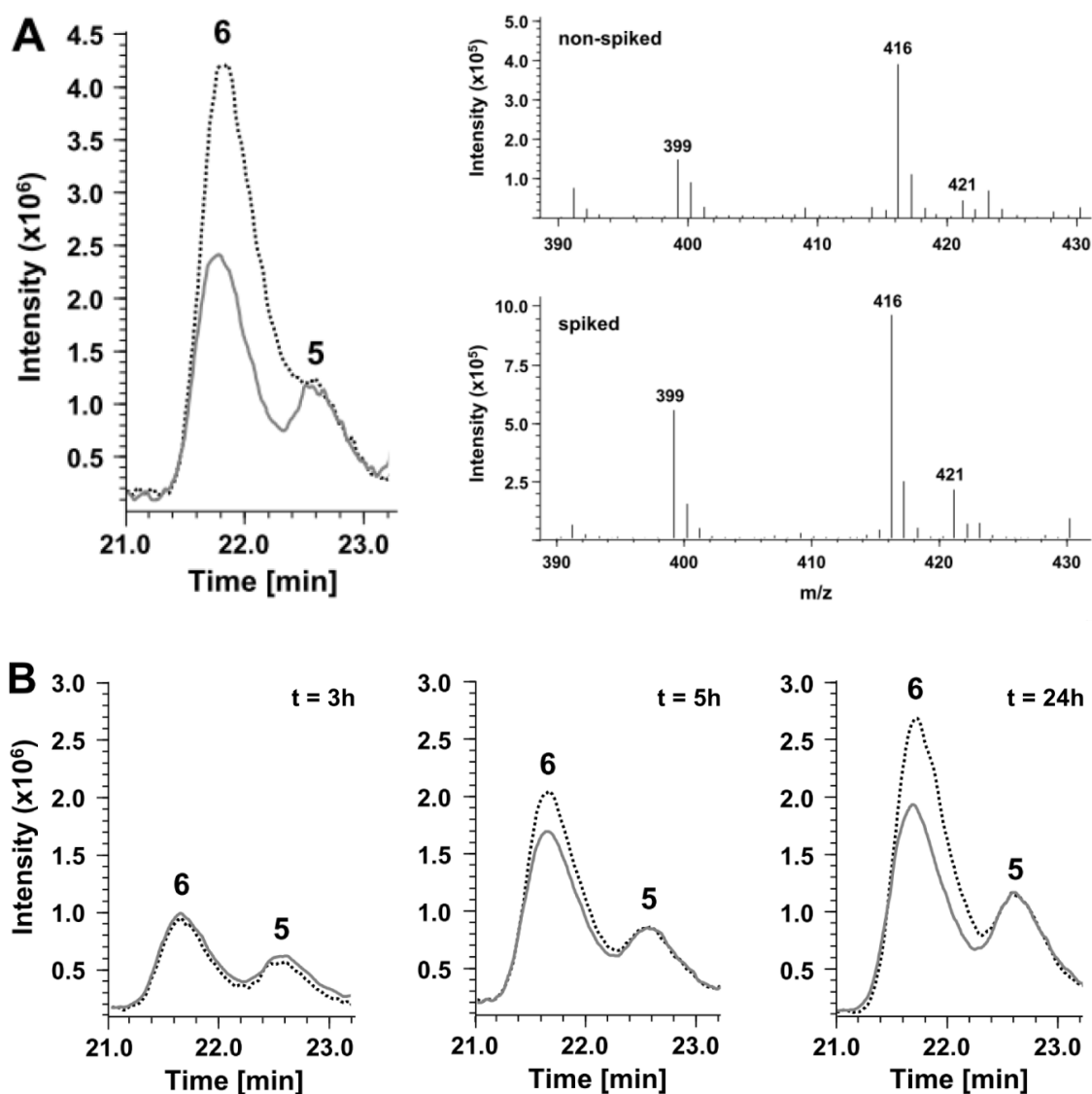


Figure S8. LC/MS analysis (method 4) of one-cell biotransformations of (-)-matairesinol **1** to (-)-deoxypodophyllotoxin **6**. **(A)** LC-chromatogram and m/z-fragmentation at RT = 21.8 min of a reaction sample spiked with 50 μ M (-)-deoxypodophyllotoxin **6** (dotted black line) in comparison to the respective non-spiked sample (solid grey line); 399 $[M+H]^+$; 416 $[M+H_2O]$; 421 $[M+Na]^+$. **(B)** (-)-Deoxypodophyllotoxin **6** and (-)-yatein **5** peaks of reaction samples at $t = 3$ h, 5 h and 24 h with addition of 2.5 mM 2-oxoglutarate at $t = 4$ h (dotted black lines) in comparison to the respective samples without addition of 2-oxoglutarate (solid grey lines).

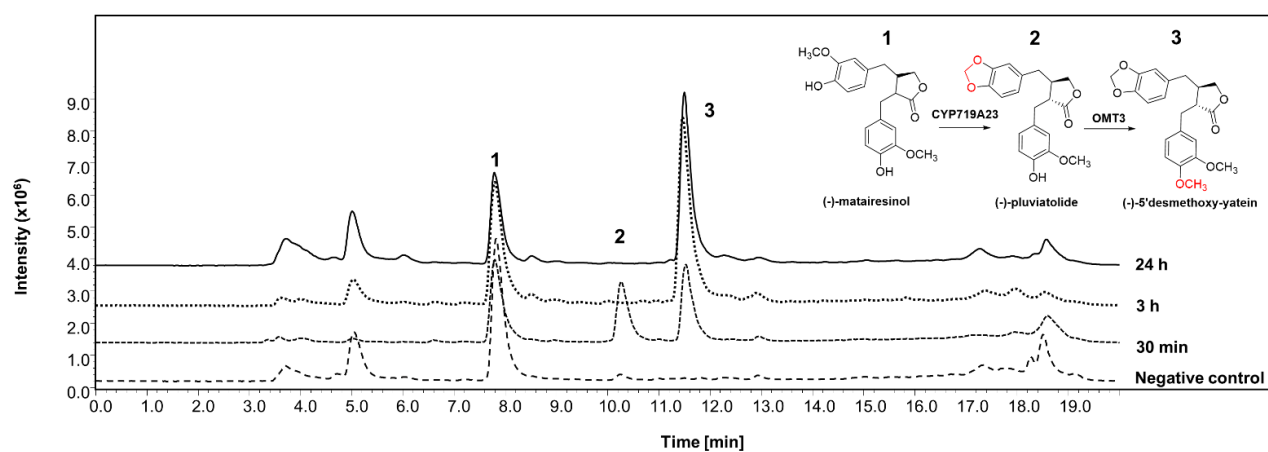


Figure S9. LC/MS analysis (method 1) of OMT3 activity in the cascade reaction with CYP719A23.

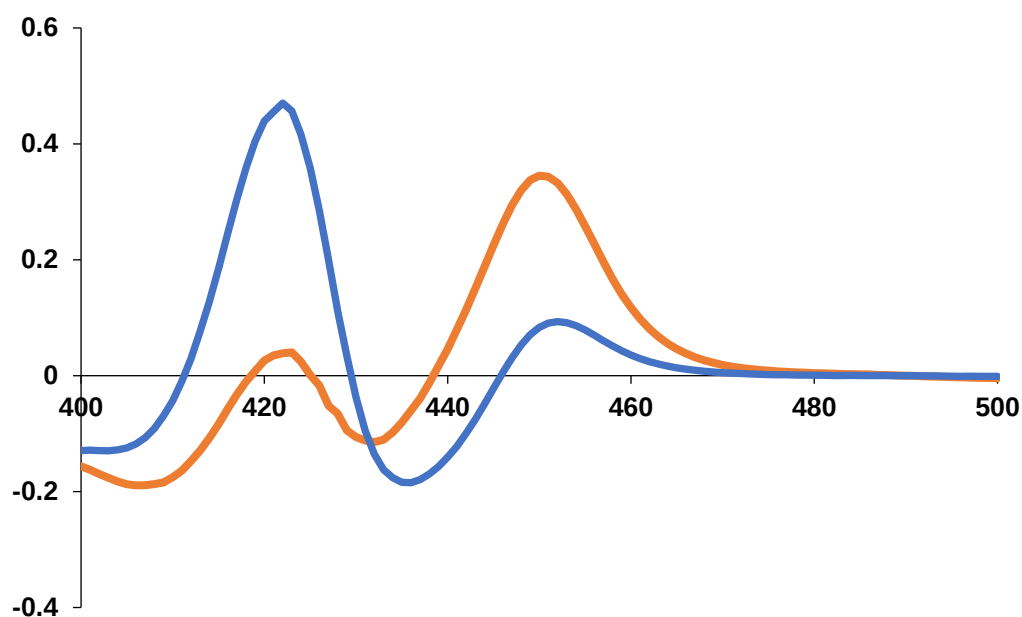
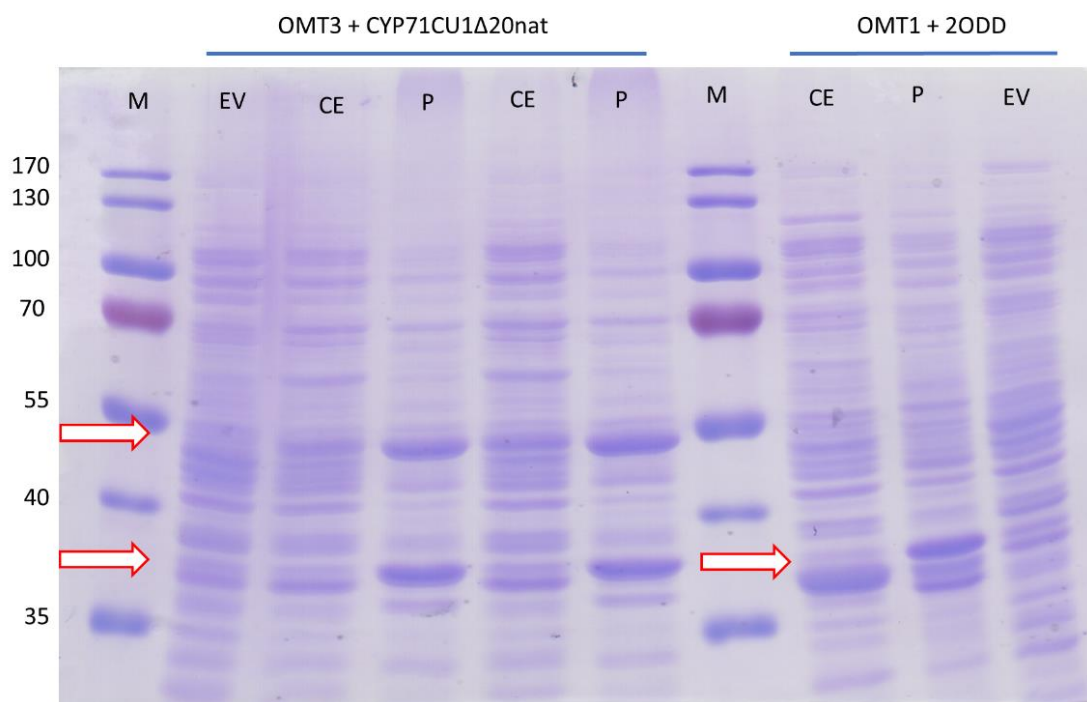
A**B**

Figure S10. (A) CO-difference spectra of single expressed CYP71CU1Δ20nat (orange line) and CYP71CU1Δ20nat co-expressed with OMT3 (blue line). **(B)** 12.5% SDS-gel of CYP71CU1Δ20nat (54 kDa) co-expression with OMT3 (41 kDa), and OMT1 (38 kDa) with 2-ODD (35 kDa). M: Molecular weight size marker, EV: Vector control without respective genes, CE: Soluble protein fraction after cell disruption, P: Insoluble protein fraction after cell disruption.

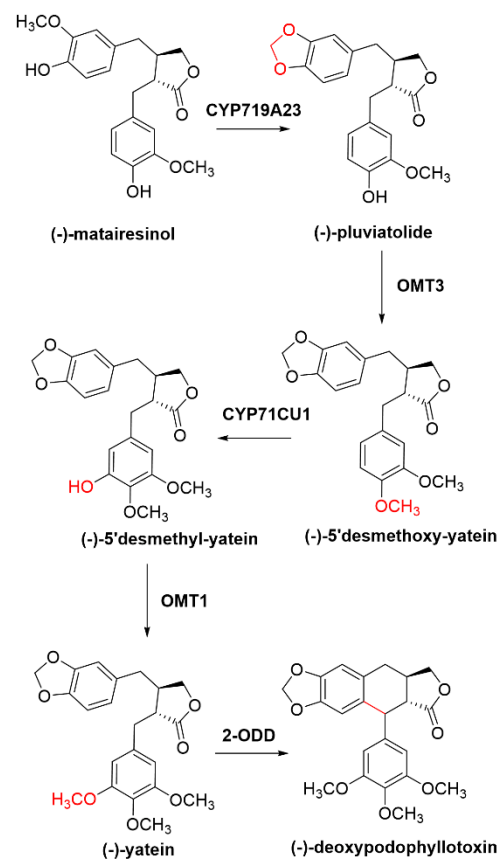
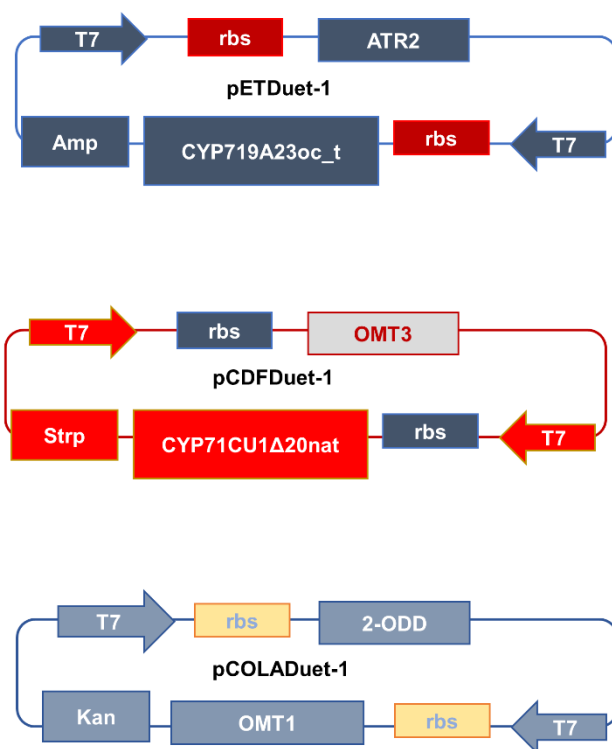
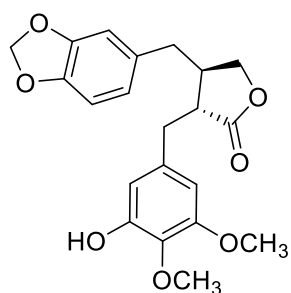


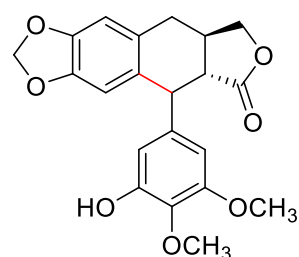
Figure S11. Schematic overview on the plasmid-based modular strategy developed to synthesize (-)-deoxypodophyllotoxin **6** from (-)-matairesinol **1** in *E. coli* via a one-cell 5-steps 6-enzyme reaction cascade.

Molecular Weight: 386,40



(-)-5'-desmethyl-yatein

Molecular Weight: 384,38



(-)-5'-desmethyl-deoxypodophyllotoxin?

2-ODD?

Figure S12. Hypothesized side reaction of 2-ODD occurring in the two-cell approach.

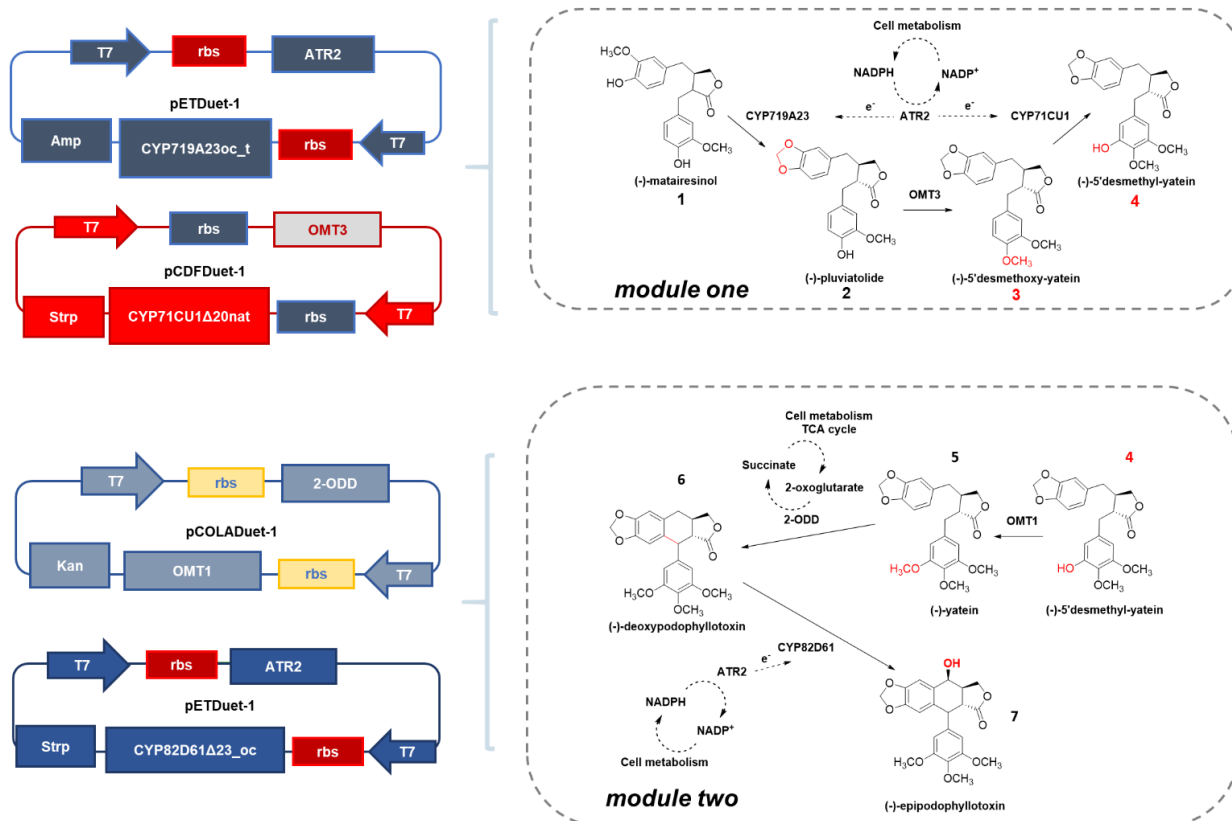


Figure S13 Schematic overview on the plasmid-based modular strategy developed to synthesize (-)-epipodophyllotoxin **7** from (-)-matairesinol **1** in *E. coli* via a two-cell 6-steps 7-enzyme reaction cascade.

4. Supplementary References

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