

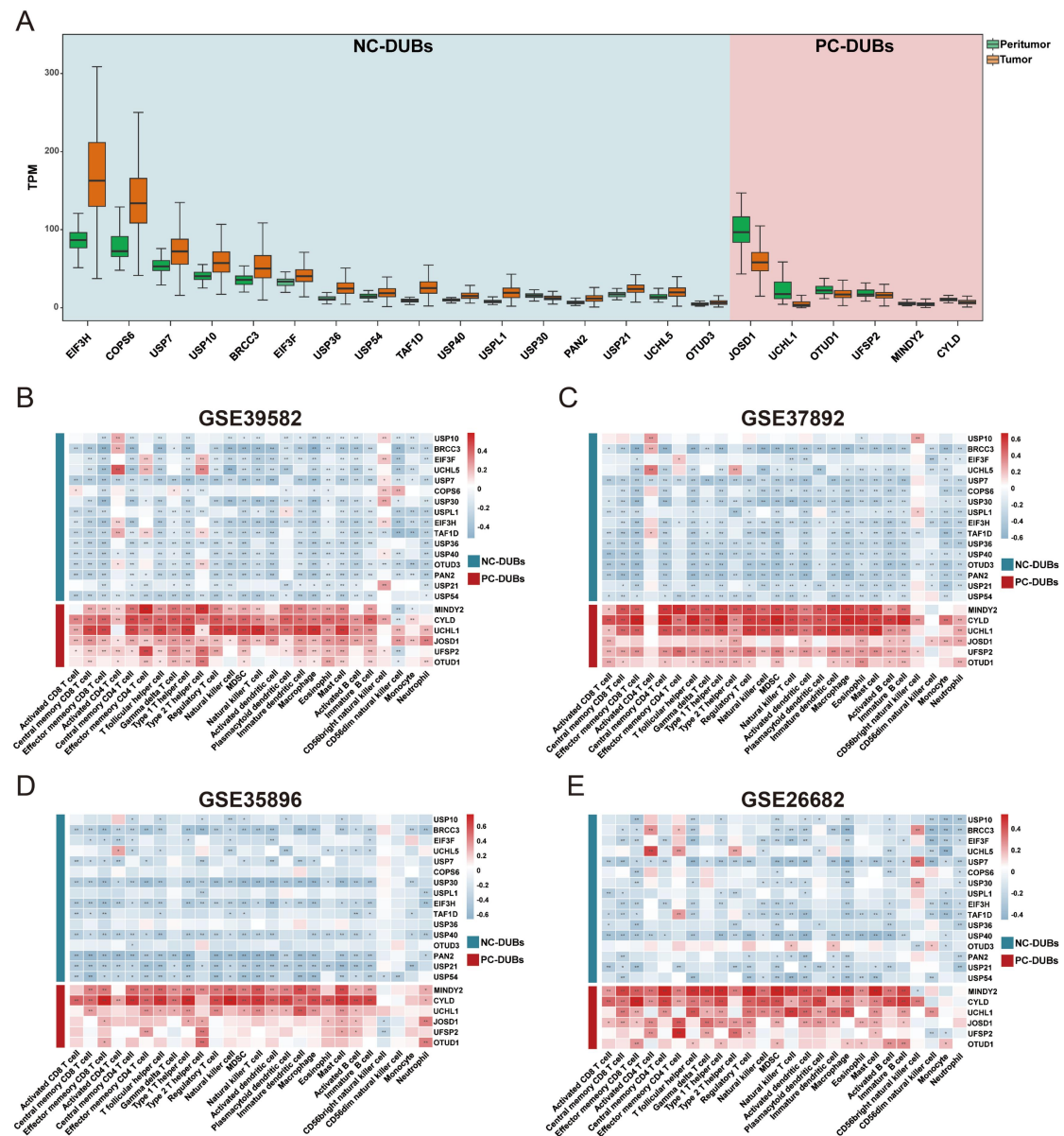


Figure S1. Related to Figure 1. (A). The expression of IR-DUBs in tumors and peritumor tissues in the TCGA cohort. Correlation of IR-DUBs with infiltrated immune cells in MSS CRC in (B) GSE39582, (C) GSE37892, (D) GSE35896, (E) GSE26682.

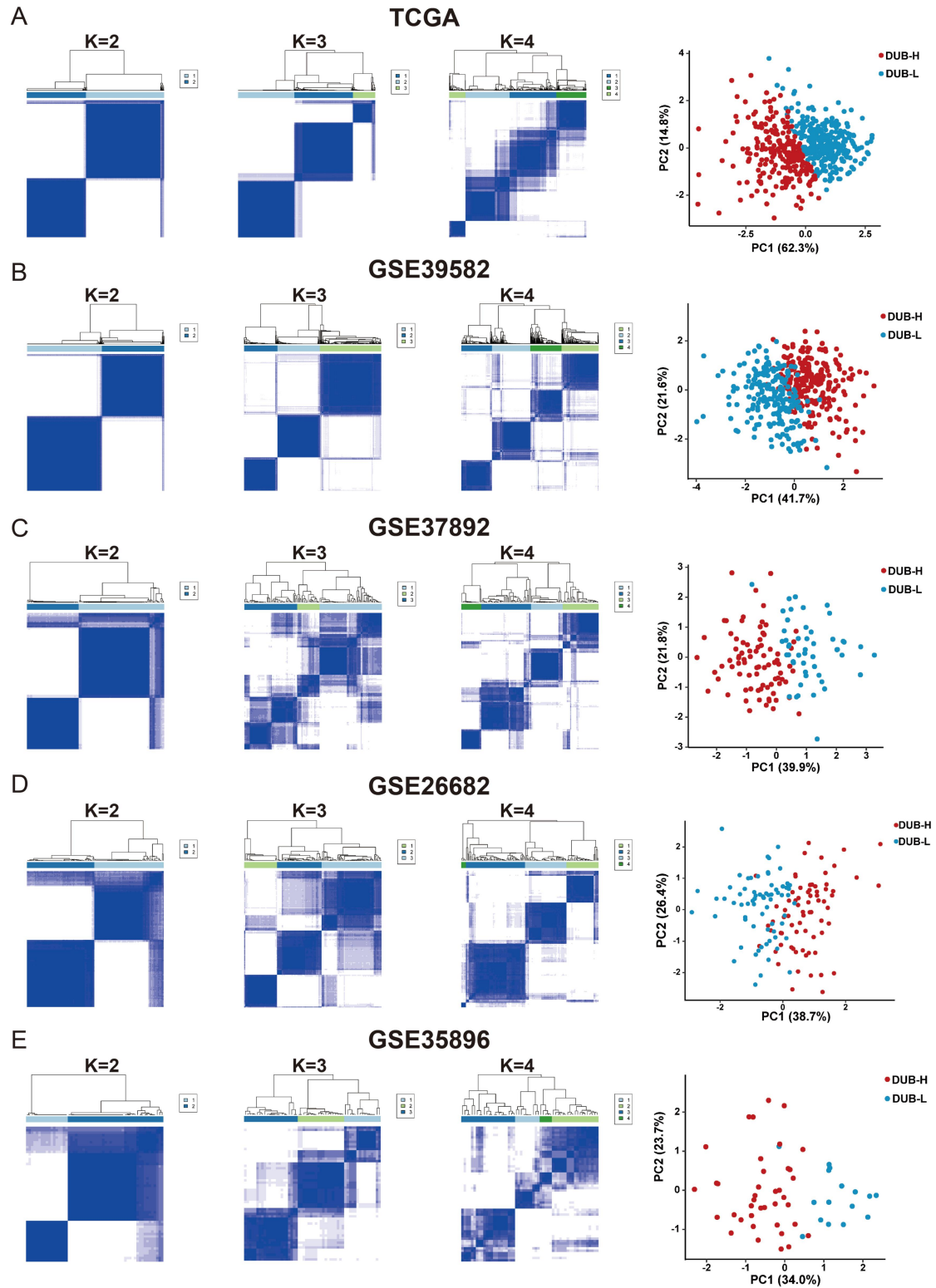


Figure S2. Related to Figure 2. (A-E). Consensus matrices of the TCGA cohort and four GEO cohorts for $k = 2 - 4$. The PCA results are displayed in the right panel between DUB-L and DUB-H classes

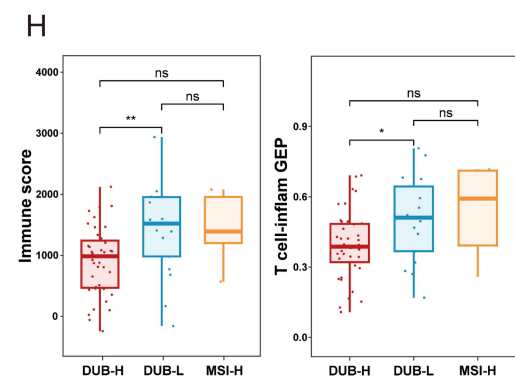
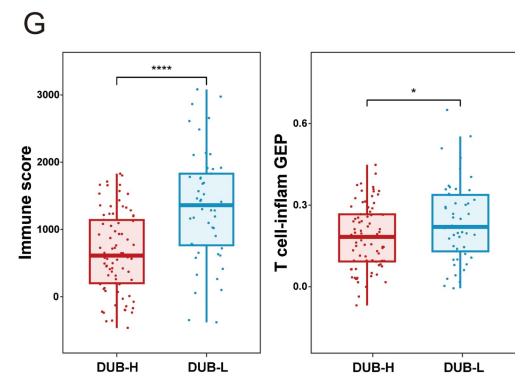
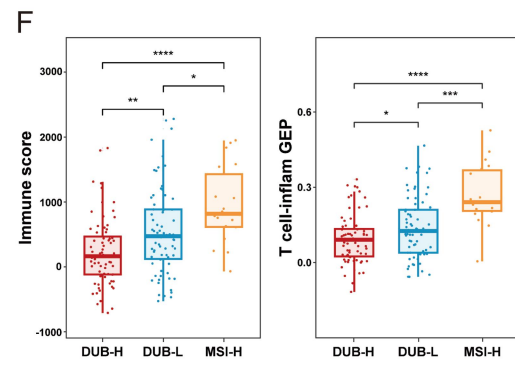
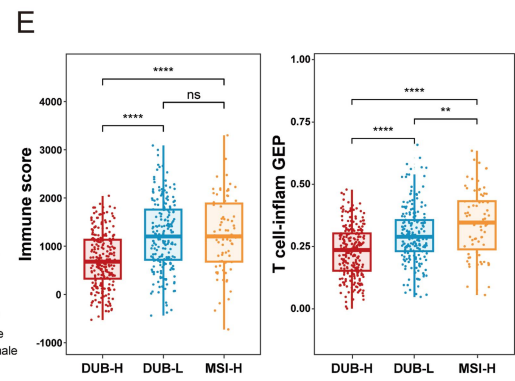


Figure S3. Related to Figure 2. (A-D) Expression spectrum of IR-DUBs within the consensus model and clinical information in DUB-L, DUB-H, and MSI-H CRC samples from (A) GSE39582, (B) GSE26682, (C) GSE37892, and (D) GSE35896 (E-H) Immune score and T cell inflamed GEP in DUB-L, DUB-H, and MSI-H CRC samples from (E) GSE39582, (F) GSE26682, (G) GSE37892, and (H) GSE35896.

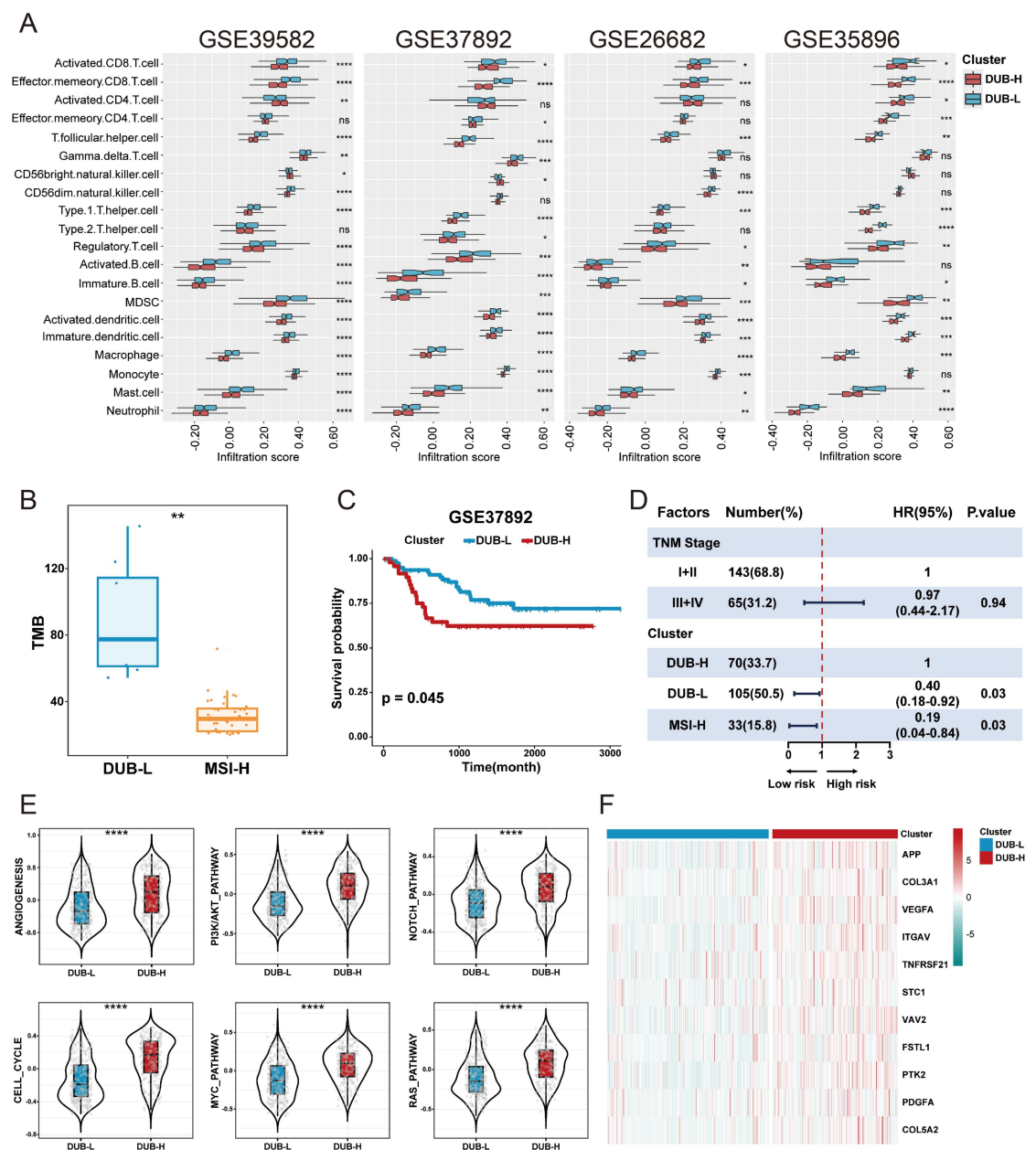


Figure S4. Related to Figure 2. (A). Infiltration score for different types

of immune cells in DUB-L and DUB-H CRC in four GEO cohorts (B). Tumor mutation burden of DUB-L and MSI-H CRC in tumor samples (with a cutoff TMB > 20 mut/Mb) (C). RFS survival plot of DUB-L and DUB-H samples in GSE37892 (D). Multivariate Cox regression of MSS subtypes and TMN stage in the TCGA cohort (E). GSVA enrichment results of DUB-L, DUB-H CRC (F). Heatmap of angiogenesis-related genes in DUB-L, DUB-H CRC

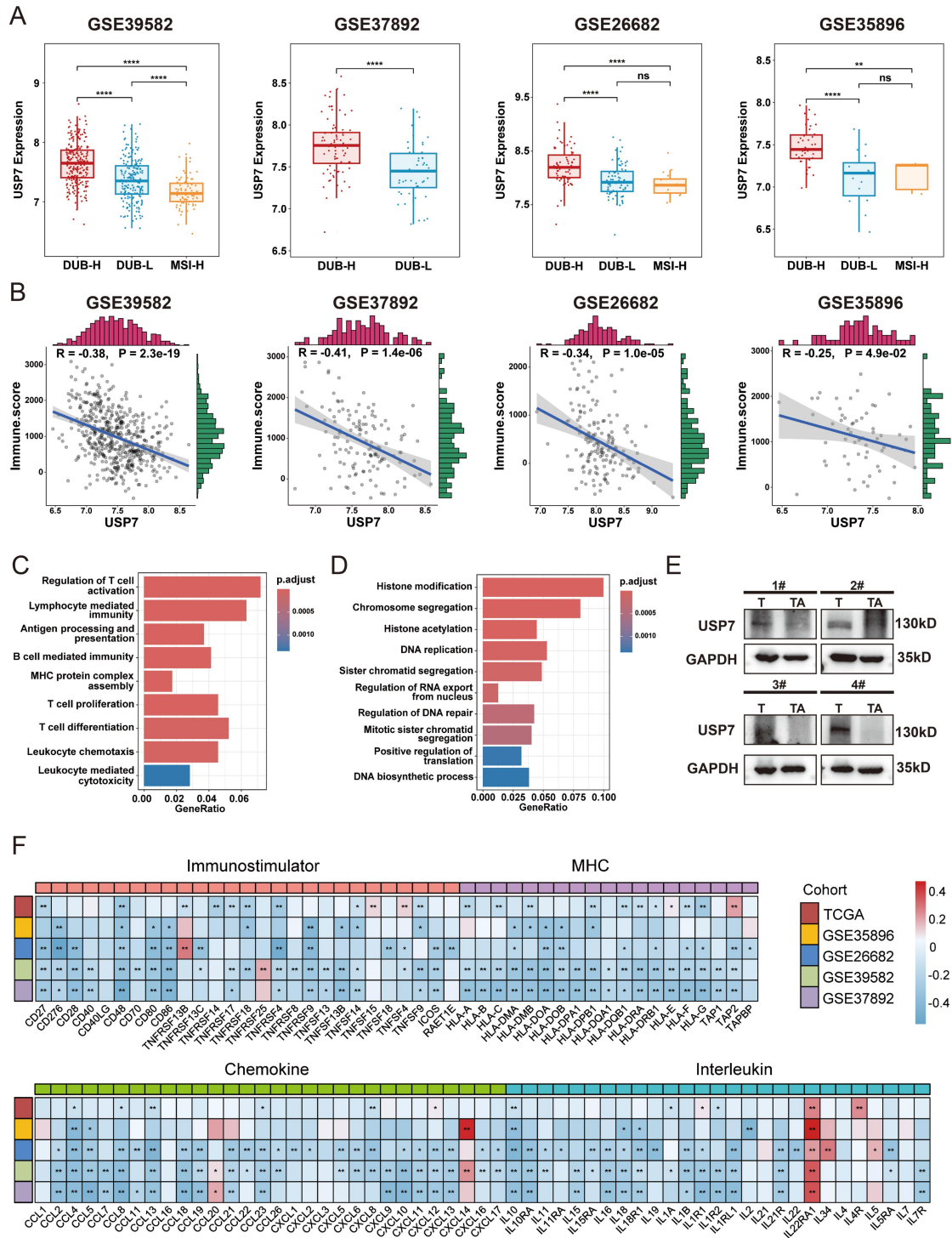


Figure S5. Related to Figure 3. (A). USP7 expression in DUB-L, DUB-H and MSI-H tumors in four GEO cohorts. (B). Correlation of USP7 expression with immune score in the MSS CRC samples across four GEO cohorts (C). Signaling pathways enriched in samples with low USP7 expression (D). Signaling pathways enriched in samples with high USP7 expression (E). USP7 protein level in tumor and peritumor tissues (F). Correlation of USP7 expression with immune-related factors in the TCGA and four GEO cohorts.

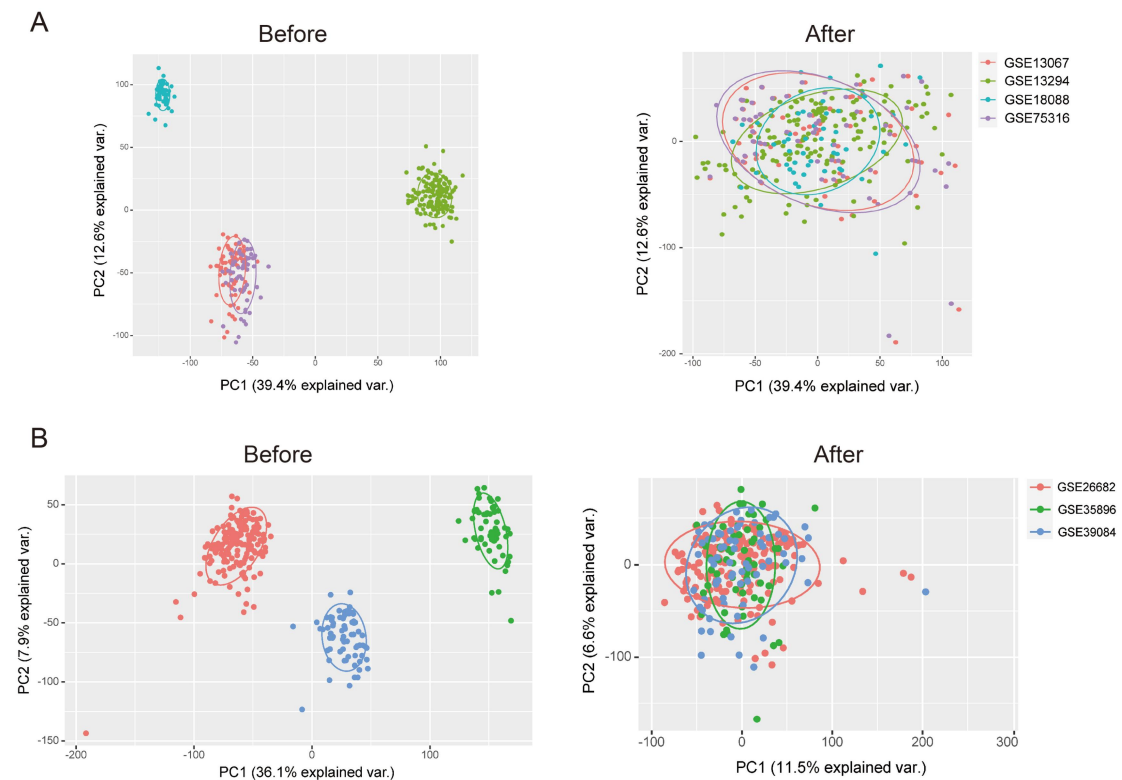


Figure S6. Related to Figure 4. (A-B) PCA analysis demonstrating the batch effects in pooled cohort 1 and cohort 2 before and after combination.

with low USP7 expression (C). Comparison of USP7 expression between tumorigenesis-related gene mutant and wild-type samples in the TCGA cohort (D). Comparison of USP7 expression between tumorigenesis-related gene mutant and wild-type samples in the GSE39582 cohort. (E). USP7 expression in proximal and distal CRC samples from the TCGA and GSE39582 cohorts (F). Correlation of USP7 expression and four mismatch repair genes (MSH2, MSH6, PMS2, MLH1) (G). USP7 expression across different TNM stages in the local cohort, TCGA and GSE39582 cohort.

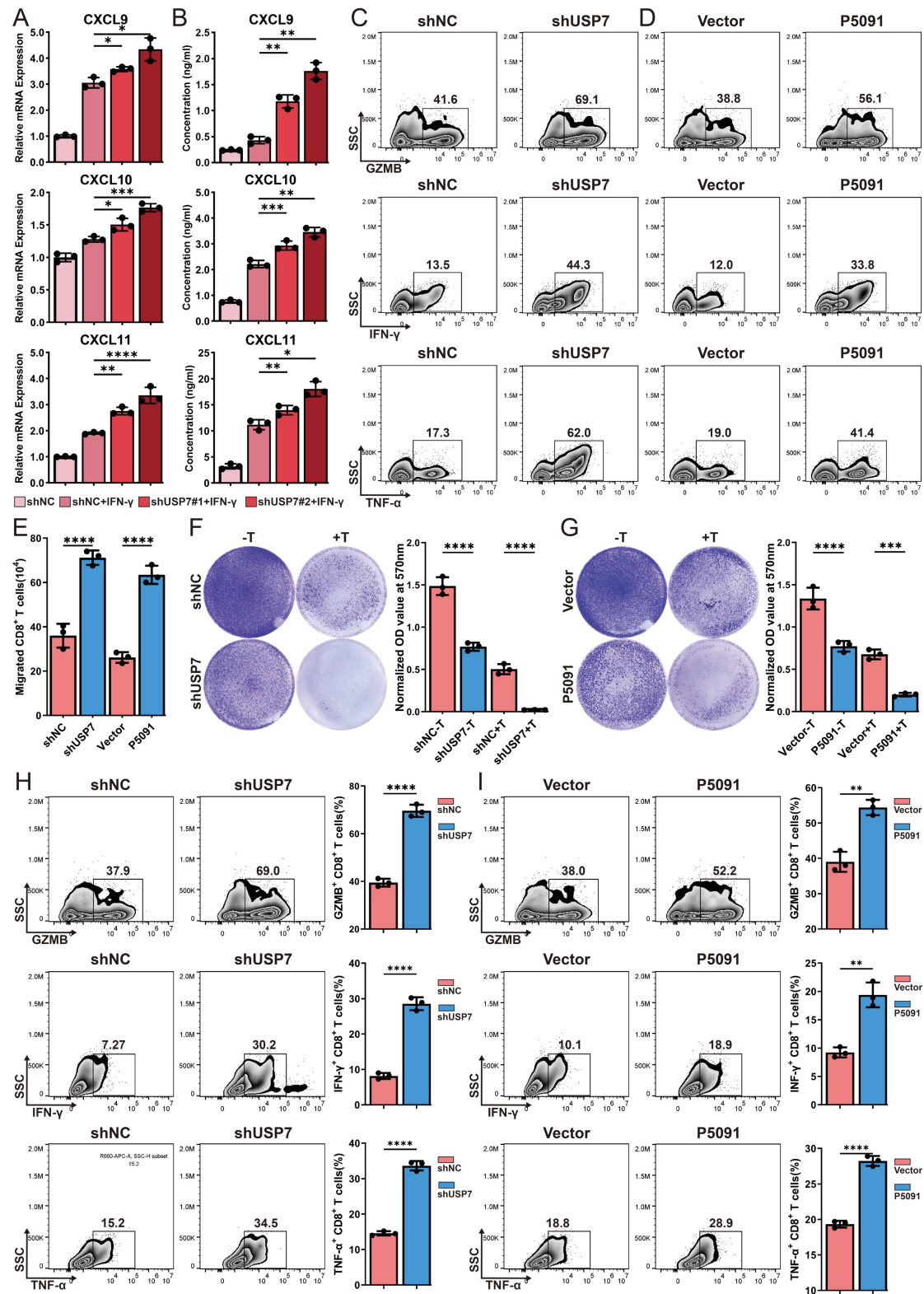


Figure S8. Related to Figure 6. (A). CXCL9, CXCL10 and CXCL11 mRNA expression measured by RT-PCR in the following groups: negative control, negative control + IFN- γ , and USP7 knockdown + IFN- γ in SW1116 cells (B). CXCL9, CXCL10 and CXCL11 protein levels quantified by ELISA in the following groups: negative control, negative control + IFN- γ , and USP7 knockdown + IFN- γ in SW1116 cell supernatants (C). Representative contour plots of GZMB⁺, IFN- γ ⁺, and TNF- α ⁺ CD8⁺ T cells after co-culture with SW480 cells bearing USP7 knockdown or a negative control (D) Representative contour plots of GZMB⁺, IFN- γ ⁺, and TNF- α ⁺ CD8⁺ T cells after co-culture with SW480 cells pretreated with P5091 or a vehicle control (E). Number of migrated CD8⁺ T cells following their co-culture with SW1116 cells that underwent either USP7 knockdown (vs. negative control) or pharmacological inhibition with P5091 (vs. vehicle control) (F). Representative images and quantitative analysis of T cell-mediated tumor cell-killing in SW1116 cells with USP7 knockdown versus negative control (G). Representative images and quantitative analysis of T cell-mediated killing of SW1116 cells pretreated with P5091 or vehicle control (H). Representative contour plots and proportions of GZMB⁺, IFN- γ ⁺, and TNF- α ⁺ CD8⁺ T cells after co-culture with USP7-knockdown or negative control SW1116 cells (I). Representative contour plots and proportions of GZMB⁺, IFN- γ ⁺, and TNF- α ⁺ CD8⁺ T cells after co-culture with SW1116 cells following P5091 or vehicle control pretreatment.

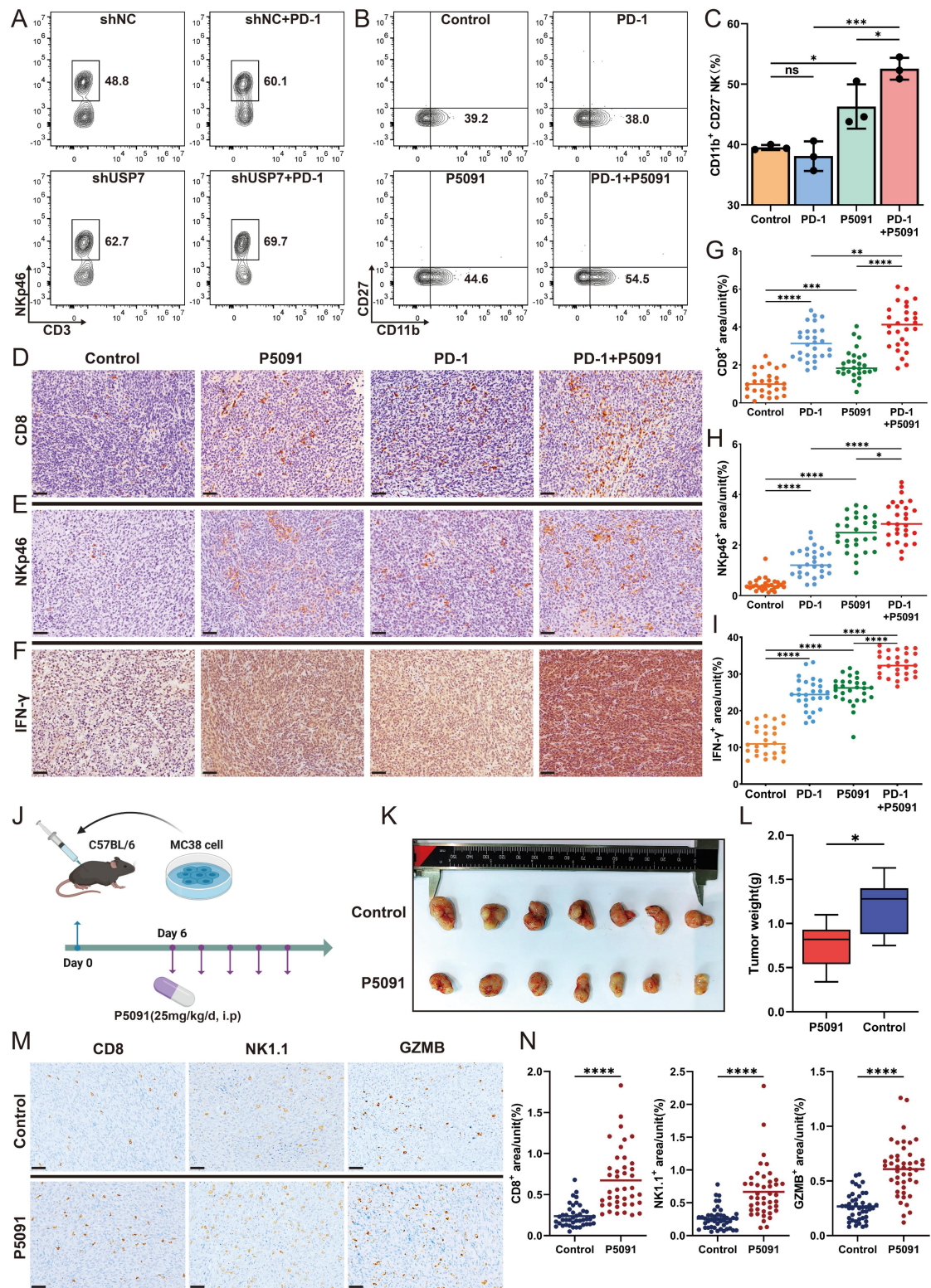


Figure S9. Related to Figure 7&8. (A). Representative contour plots of CD3-NKp46⁺ NK cells in tumors across different groups. (B). Representative contour plots of CD27-CD11b⁺ NK cells in tumors across different groups. (C). Proportions of CD27-CD11b⁺ NK cells in tumors across different groups. (D-F) Representative IHC images of (D) CD8, (E) NKp46 and (F) IFN- γ in tumors from different groups. (G-I) Statistic analysis of (G) CD8, (H) NKp46 and (I) IFN- γ expression in tumors from different groups. (J). Schematic diagram of animal study. C57BL/6 mice was subcutaneously injected with MC38 cells, drug injection (P5091, 25mg/kg/d, i.p.) began on day 6, and tumors were harvested on day 21 (K). Tumor images and between P5091-treated and control groups (L). Tumor weights in P5091-treated and control groups. (M). Representative IHC staining for CD8, NK1.1 and GZMB in P5091-treated and control tumors. (N). Quantification of IHC staining for CD8, NK1.1 and GZMB in P5091-treated and control tumors.

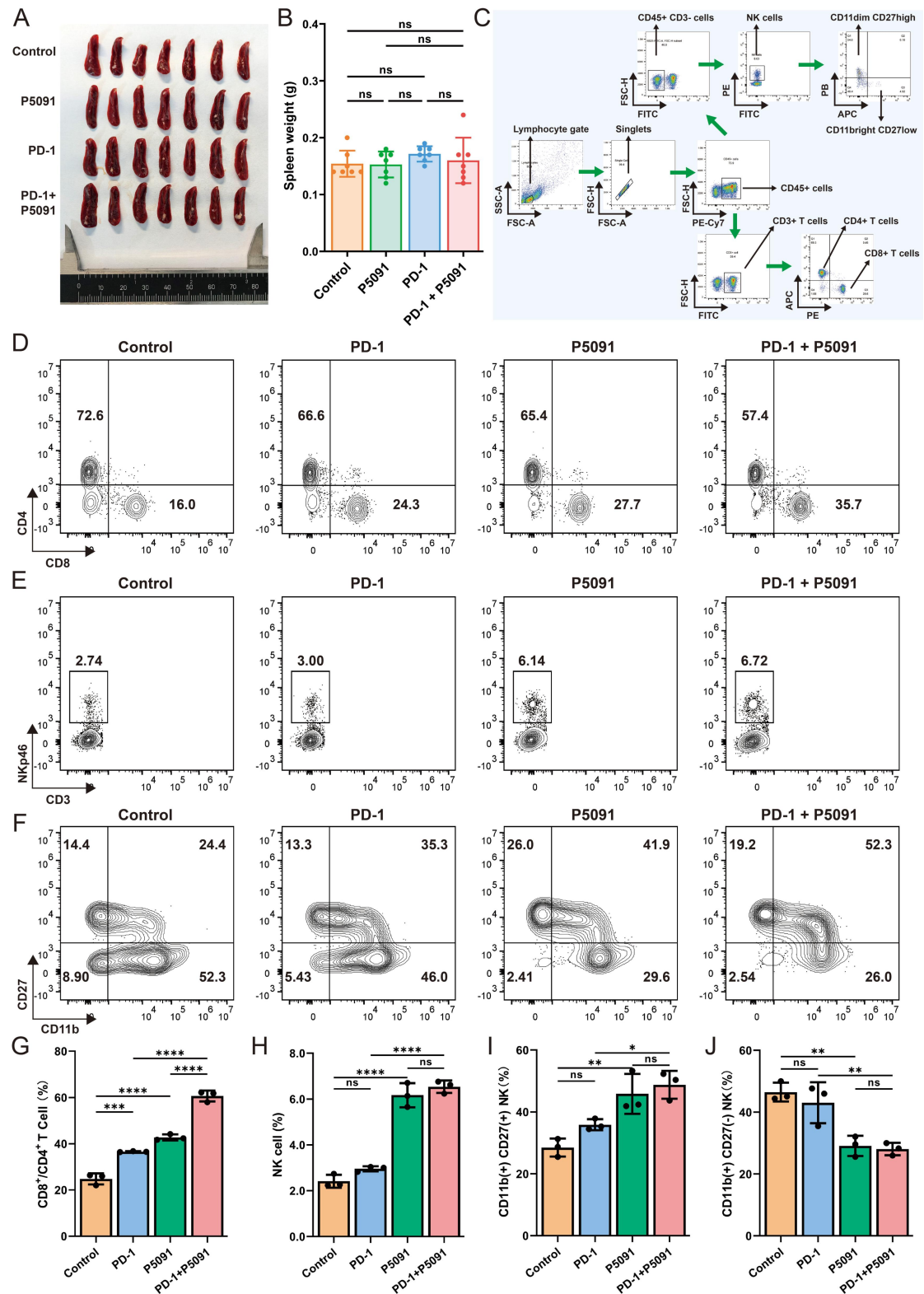


Figure S10. Related to Figure 8. Spleen images (A) and weights (B) of different groups after CT26 mice sacrifice (C) Flow cytometry gating strategy for identifying immune cell populations in tumor tissue: CD4⁺ T

cells (CD3⁺CD4⁺), CD8⁺ T cells (CD3⁺CD8⁺), NK cells (CD3⁻NKp46⁺), and CD11b⁺CD27⁻ NK cells. Lymphocytes were gated based on SSC-A vs. FSC-A, and singlets were selected using FSC-A vs. FSC-H (D). Representative flow plots of CD8⁺ and CD4⁺ T cells in spleens across different groups (E). Representative flow plots of NK cells in spleens across different groups (F). Representative flow plots of NK cell subtypes in spleens across different groups (G). Ratios of CD8⁺ to CD4⁺ T cells in spleens across different groups (H). Proportions of NK cells of spleens in different groups (I). Proportions of CD11b⁺CD27⁺ NK cells across different groups (J). Proportions of CD11b⁺CD27⁻ NK cells across different groups.

Table S1. Correlations of IR-DUBs with infiltrated CD8⁺T / NK cells in the TCGA cohort

	ssGSEA	MCP-counter	TIMER	CIBERSORT	EPIC	xCell	TIP	quanTIseq
USP10	-	-	-	-	-	-	-	○
USP7	-	○	-	-	-	-	-	○
BRCC3	-	○	-	-	-	-	-	○
EIF3F	-	-	-	○	○	-	-	-
UCHL5	-	-	○	○	-	-	-	○
USP54	-	○	-	-	-	○	-	○
COPS6	-	-	-	○	○	-	○	-
USP30	-	-	○	-	-	-	○	○
USPL1	-	○	○	-	-	-	-	○
EIF3H	-	-	○	○	-	-	○	○
TAF1D	-	-	○	○	-	-	○	○
USP36	-	-	○	○	-	-	○	○
USP40	-	-	○	○	-	-	○	○
OTUD3	-	○	-	○	-	○	○	○
PAN2	-	-	○	○	-	○	○	○
USP21	-	○	○	○	○	-	-	○
MINDY2	+	+	+	+	+	+	+	+
CYLD	+	+	+	+	+	+	+	+
UCHL1	+	+	+	○	+	+	+	+
JOSD1	+	+	+	○	○	+	○	+
UFSP2	+	○	+	○	○	+	○	+
OTUD1	+	○	+	○	○	+	○	+

“+”: positive correlation, “-”: negative correlation, “○”: Not significant

Heatmap showing the correlation of 15 proteins. The color scale ranges from -0.2 (blue) to 1.0 (red). A legend on the right indicates significance levels: ***: P < 0.001, **: P < 0.01, *: P < 0.05.

Table S3. Online clinical cohorts used in this study

Cohort	Sample description	platform
TCGA-COAD&READ	529 MSS CRC samples 90 MSI CRC samples	Illumina HiSeq 2000
GSE39582	444 MSS CRC samples 75 MSI CRC samples	GPL570
GSE37892	130 MSS CRC samples	GPL570
Pooled_cohort 1 (GSE13067+GSE13294+ GSE18088+GSE75316)	222 MSS CRC samples 119 MSI CRC samples	GPL570
Pooled_cohort 2 (GSE26682+GSE35896+GSE39084)	252 MSS CRC samples 39 MSI CRC samples	GPL570

Table S4. Clinical features of CRC patients in the Local cohort 1(LC1)

Features		n(%)	
Gender			
	Male	43	(57)
	Female	33	(43)
Age			
	≤60	36	(47)
	>60	40	(53)
TNM Stage			
	I-II	39	(51)
	III-IV	37	(49)
MSI status			
	MSI-H	9	(12)
	MSS	67	(88)
Location			
	Distal	34	(45)
	Proximal	42	(55)

Table S5. Clinical features of CRC patients in the Local cohort 2(LC2)

Features		n(%)	
Gender			
	Male	36	(68)
	Female	17	(32)
Age			
	≤60	21	(40)
	>60	32	(60)
TNM Stage			
	I	18	(34)
	II	18	(34)
	III	17	(32)
MSI status			
	MSI-H	7	(13)
	MSS	46	(87)
Location			
	Distal	26	(49)
	Proximal	27	(51)

Table S6. Clinical features of CRC patients with anti-PD-1 treatment

ID	Gender	Age	Stage	MSI status	Treatment	Response	OS	Status
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Patient 1	Female	65	IV	MSS	anti-PD-1	PR	103.5395	Dead
Patient 2	Male	71	IV	MSS	anti-PD-1	PR	426.5968	Dead
Patient 3	Male	66	IV	MSS	anti-PD-1	PR	1099.638	Dead
Patient 4	Male	62	IV	MSS	anti-PD-1	PR	770.5662	Dead
Patient 5	Female	66	IV	MSS	anti-PD-1	PR	677.5172	Dead
Patient 6	Male	59	IV	MSS	anti-PD-1	SD	245.6667	Dead
Patient 7	Female	75	IV	MSS	anti-PD-1	PD	80.50017	Dead
Patient 8	Male	71	IV	MSS	anti-PD-1	SD	328.5172	Dead
Patient 9	Male	57	IV	MSS	anti-PD-1	PD	14.50624	Dead
Patient 10	Male	66	IV	MSS	anti-PD-1	PD	154.5888	Dead

Table S7. Sequences of primers for qRT-PCR.

Gene		Sequence
Human GAPDH	5'Primer	GAGAAGGCTGGGGCTCATTT
	3'Primer	AGTGATGGCATGGACTGTGG
Human USP7	5'Primer	CCCTCCGTGTTTTGTGCGA
	3'Primer	AGACCATGACGTGGAATCAGA
Human CXCL9	5'Primer	TGAGAAAGGGTCGCTGTTCC
	3'Primer	GGGCTTGGGGCAAATTGTTT
Human CXCL10	5'Primer	ACTGCCATTCTGATTTGCTGC
	3'Primer	ATGCAGGTACAGCGTACAGT
Human CXCL11	5'Primer	GAGTGTGAAGGGCATGGCTA
	3'Primer	ATAAGCCTTGCTTGCTTCGAT