

Additional file 1

C3G deregulation uncovers a dual role in B-cell lymphoma: tumor suppression and enhanced metastasis via Rap1 and Rac2 signaling

Running title: C3G dual function in B-cell lymphoma

Alba Morán-Vaquero^{1,2,3}, Óscar Herranz^{1,2,3}, Ana Dávila-Hidalgo^{1,2,3}, Antonio Rodríguez-Blázquez^{1,2,3}, Cristina Fernández-Infante^{1,2,3}, Ignacio García-Tuñón^{1,2,4}, Elena Vuelta^{1,2,4}, Femke van der Meer⁵, Coert Margadant⁵, Carmen Guerrero^{1,2,3*} and José M. de Pereda^{1*}

¹Centro de Investigación del Cáncer, Consejo Superior de Investigaciones Científicas (CSIC), Universidad de Salamanca, 37007 Salamanca, Spain.

²Instituto de Investigación Biomédica de Salamanca (IBSAL), Salamanca, Spain.

³Departamento de Medicina, Universidad de Salamanca, Salamanca, Spain.

⁴Departamento de Biomedicina y Biotecnología. Universidad de Alcalá, Alcalá de Henares, Spain.

⁵Institute of Biology, Leiden University, Gorlaeus Building, Einsteinweg 55, 2333 CC Leiden, The Netherlands.

*Correspondence and equal contribution: C. Guerrero and JM. de Pereda, Centro de Investigación del Cáncer, Campus Unamuno s/n, Salamanca, Spain. Tel.: +34 923294801; e-mail: cguerrero@usal.es; jm.depereda@csic.es

Supplementary Tables

Table S1. Primary antibodies used in western blot.

Antibody	Host	Supplier	Reference	RRID	Dilution
β-actin	Mouse	Sigma-Aldrich	A5441	AB_476744	1:1000
β-tubulin (2-28-33)	Mouse	Sigma-Aldrich	T5293	AB_477580	1:1000
Bcl-xL	Rabbit	Cell Signaling	2762	AB_10694844	1:1000
C3G (F-5)	Mouse	Santa Cruz Biotechnology	sc-376992	n.a.	1:1000
C3G (G-9)	Mouse	Santa Cruz Biotechnology	sc-393836	n.a.	1:1000
C3G (G-4)	Mouse	Santa Cruz Biotechnology	sc-17840	AB_628203	1:1000
Cdc42	Rabbit	Proteintech	10155-1-AP	AB_2078096	1:1000
ERK1 (K-23)	Rabbit	Santa Cruz Biotechnology	sc-94	AB_2140110	1:1000
p-ERK (E-4)	Mouse	Santa Cruz Biotechnology	sc-7383	AB_627545	1:1000
GFP (FL)	Rabbit	Santa Cruz Biotechnology	sc-8334	AB_641123	1:1000
GST (B-14)	Mouse	Santa Cruz Biotechnology	sc-138	AB_627677	1:1000
polyHis	Mouse	Sigma-Aldrich	H1029	AB_260015	1:1000
MEK1/2	Rabbit	Cell Signaling	9122	AB_823567	1:1000
phospho-MEK1/2	Rabbit	Cell Signaling	9121	AB_331648	1:1000
PP2A, C sub (1D6)	Mouse	Merck	05-421	AB_309726	1:1000
Rac1	Mouse	Cytoskeleton	ARC03	AB_10709099	1:500
Rac2	Rabbit	Proteintech	10735-1-AP	AB_2176127	1:1000
c-Raf (D43BJ)	Rabbit	Cell Signaling	53745	AB_2799444	1:1000
Rap1 (E-6)	Mouse	Santa Cruz Biotechnology	sc-398755	AB_2884025	1:800
pan-Ras (C-4)	Mouse	Santa Cruz Biotechnology	sc-166691	AB_2154229	1:1000
Talin (C-20)	Goat	Santa Cruz Biotechnology	sc-7534	AB_661610	1:500

n.a.: Not available

Table S2. Secondary antibodies used in western blot.

Antibody	Host	Supplier	Reference	RRID	Dilution	Detection
anti-mouse Ig (H+L) DyLight 800	Goat	ThermoFisher	SA5-35521	AB_2556774	1:5000	Odyssey
anti-mouse IgG (H+L) DyLight 680	Goat	ThermoFisher	SA5-35518	AB_614942	1:5000	Odyssey
anti-rabbit IgG (H+L) DyLight 800	Goat	ThermoFisher	SA5-10036	AB_2556616	1:10000	Odyssey
anti-rabbit IgG (H+L) DyLight 680	Goat	ThermoFisher	SA5-35568	AB_614946	1:10000	Odyssey
anti-goat AF680 IgG (H+L)	Donkey	ThermoFisher	A-21084	AB_2535741	1:5000	Odyssey
anti-mouse IgG-HRP	Sheep	Cytiva	NXA931	AB_772209	1:5000	ECL
anti-rabbit IgG-HRP	Goat	Santa Cruz Biotechnology	sc-2004	AB_631746	1:10000	ECL

Table S3. Sequence of forward and reverse primers used in RT-qPCR to quantify the expression of the indicated mouse genes.

Gene	Forward primer (5'-3')	Reverse primer (5'-3')
N-terminal <i>Rapgef1</i>	GTGAGCAAAGAGGCAAGAGA	CACAGCACTGGTGGACATAA
C-terminal <i>Rapgef1</i>	ATTTCCACAGCCACGAGATAG	CTCTTCTCCTCATTCTGCTCTT
<i>Araf</i>	TGGCGATGTAGCTGTGAAAG	CTGTGTGATGATGGCAAACC
<i>Braf</i>	ATGCCCTTCAGCAAAGAGAA	ATCTTGCGGGTACCACTGTC
<i>Craf</i>	CTACACCCCATGCCTTCACT	GCTGAAGGTGAGGCTGATTC
<i>Phlda1</i>	GGGCTACTGCTCATAACCGC	AAAAGTGCAATTCCTTCAGCTTG
<i>Spry2</i>	TCCAAGAGATGCCCTTACCCA	GCAGACCGTGGAGTCTTTCA
<i>Dusp6</i>	GCGTCGGAAATGGCGATCT	ATGTGTGACGACTCGTACAGC
<i>Dusp4</i>	CGTGCGCTGCAATACCATC	CTCATAGCCACCTTTAAGCAGG
<i>Etv4</i>	CGGAGGATGAAAGCGGATAC	TCTTGGAAGTGACTGAGGTCC
<i>Etv5</i>	TCAGTCTGATAACTTGGTGCTTC	GGCTTCCTATCGTAGGCACAA
<i>Dock1</i>	CAGGAAGCATAAATACCTCGCC	CAGCTCATCCGATTGTCTTTGT
<i>Cyria</i>	GAGCGAGAGATATGGAACCAGA	AGTGCTTTTTCAAGACGGATGG
<i>Armex3</i>	CTGGAGCCTGCTATTGCATTT	TCAGACCAGTCATTATACCTGGC
<i>Armex6</i>	TGGGAAGAAGTGAGGGGAAC	GTCGAGCCATTGCTGTGAAAT
<i>Nol4l</i>	GGACCAGCCATTAACTGT	GCTGTATGGGGGTGACTCG
<i>Actb</i>	CCACCATGTACCCAGGCATT	CAGCTCAGTAACAGTCCGCC

Supplementary figures

Figure S1

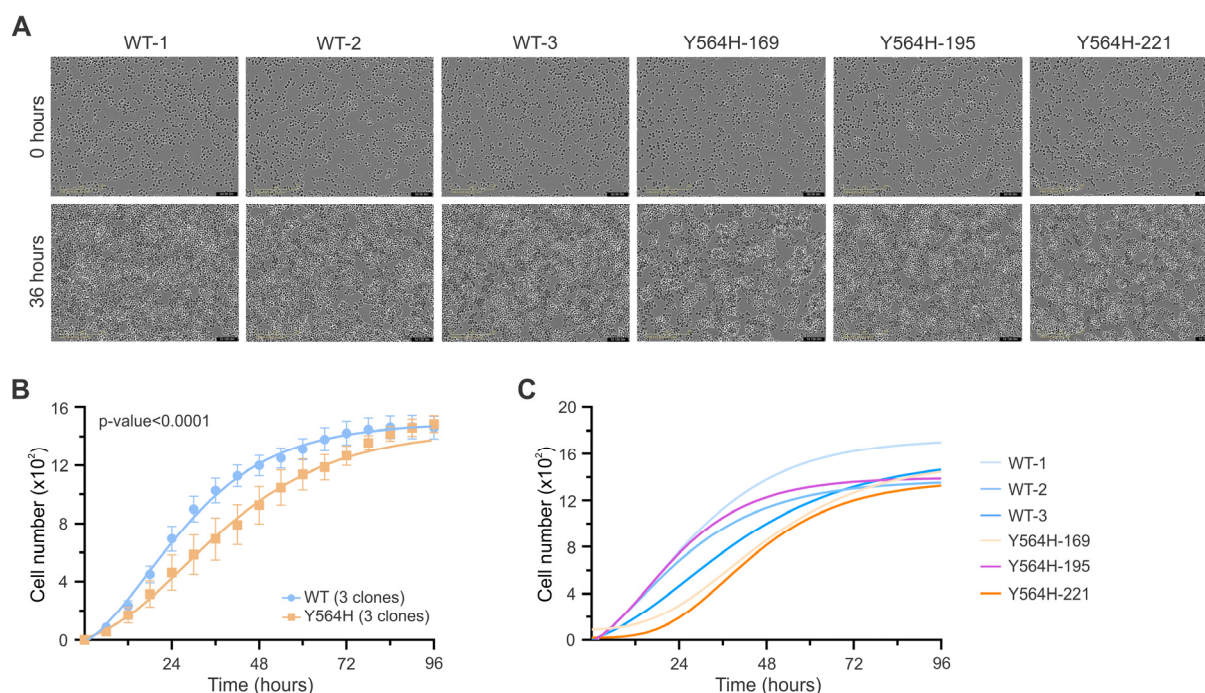


Figure S1. C3G-Y564H mutation slows down A20 cell proliferation. The number of live cells was quantified using live-cell imaging of label-free cell module with an Incucyte SX5 instrument (Sartorius). Images were captured at 6-hour intervals over 4 days, analyzing three A20-C3G-WT and three A20-C3G-Y564H cell clones. **(A)** Representative images at 0 hours (upper panels) confirm the initial seeding density, while images at 36 hours (lower panels) highlight the most pronounced differences among clones. **(B)** Time course of cell numbers presented as the mean $\pm$ SEM of grouped data from three A20-C3G-WT and three A20-C3G-Y564H clones. Images were processed using the Non-Adherent Cell-by-Cell Analysis software module. Lines represent a 4-parameter Gompertz growth model fitted to the experimental data. **(C)** Growth curves of individual clones, each fitted with a 4-parameter Gompertz growth model. Two independent experiments were performed per clone, with experimental triplicates and four images taken per condition.

Figure S2

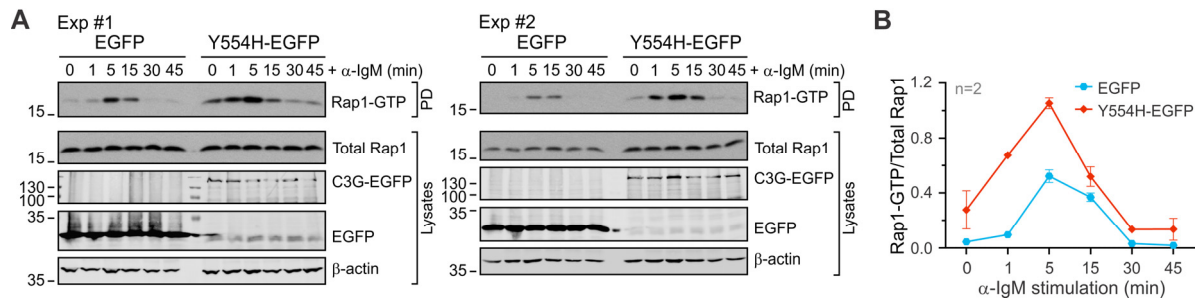


Figure S2. Expression of C3G-Y554H in A20 cells increases Rap1 activation. A20 cells were electroporated with the pEF1-C3GY554H-EGFP construct to generate C3G-Y554H-expressing clones. **(A)** Two Rap1-GTP pull-down experiments are shown, including C3G protein levels. **(B)** Quantification of Rap1-GTP levels demonstrates an increase in C3G-Y554H-overexpressing A20 cells compared with control cells.

Figure S3

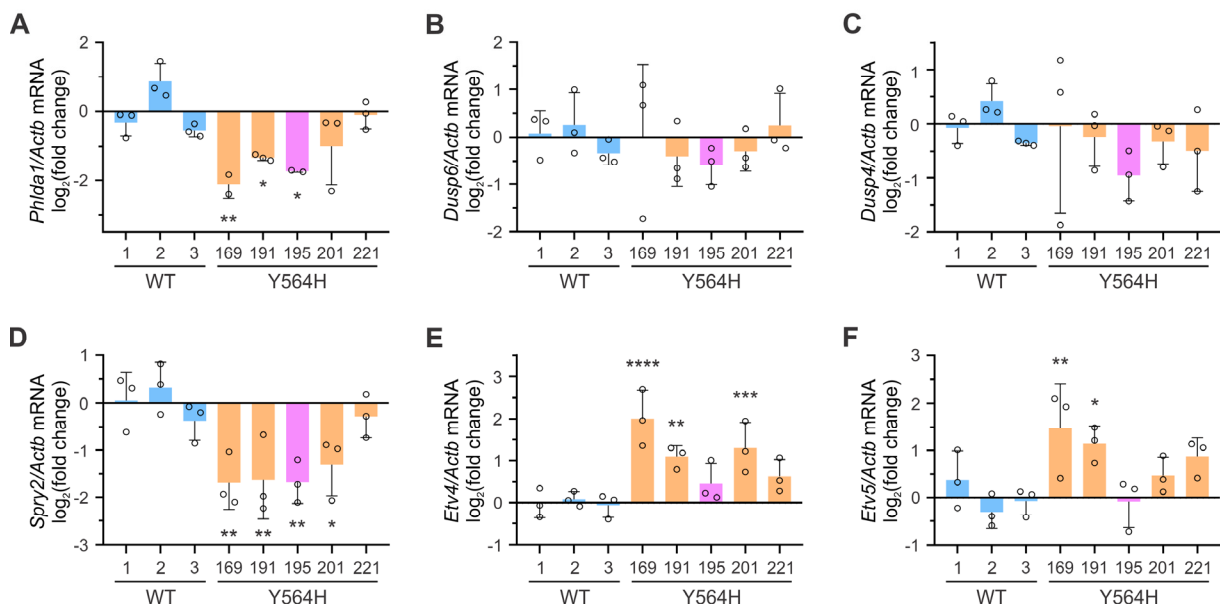


Figure S3. A20-C3G-Y564H cells display altered expression of ERK1/2-regulated genes. mRNA expression levels of **(A)** *Phlda1*, **(B)** *Dusp6*, **(C)** *Dusp4*, **(D)** *Spry2*, **(E)** *Etv4*, and **(F)** *Etv5* were analyzed in A20-C3G-Y564H and A20-C3G-WT clones by RT-qPCR. Gene expression levels were relative to *Actb*. Bar charts represent the mean $\pm$ SEM of the $\log_2$ (fold change) from three independent experiments, with individual data points shown. * p <0.05, ** p <0.01, *** p <0.001, **** p <0.0001.

Figure S4

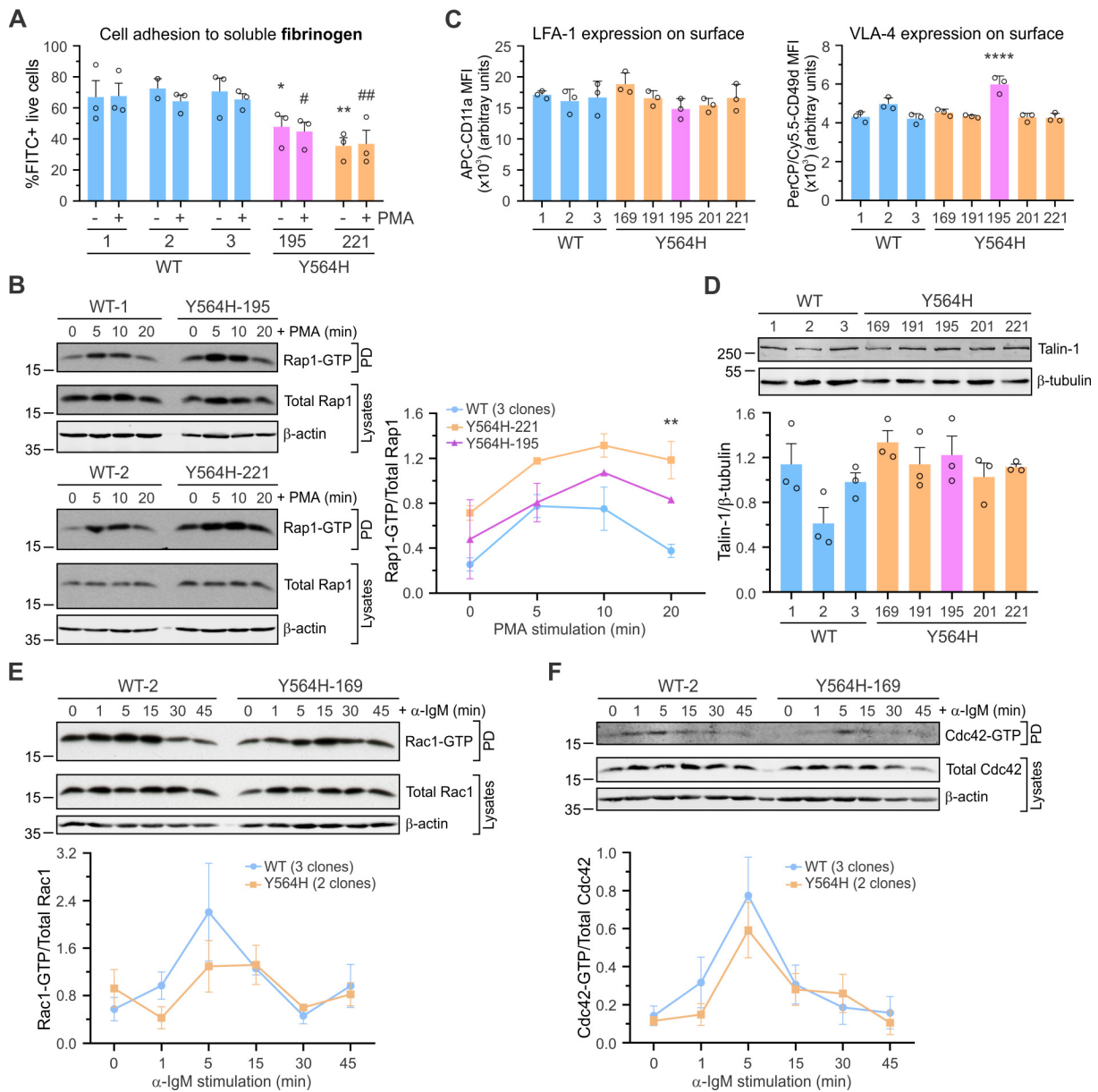


Figure S4. A20-C3G-Y564H cells exhibit reduced fibrinogen binding with no changes in LFA-1, VLA-4, or Talin-1 levels or in Rac1 and Cdc42 activity. (A) Cells were stimulated with 20 nM PMA and incubated with Alexa Fluor 488-conjugated human fibrinogen. The bar chart represents the mean $\pm$ SEM of the percentage of live cells bound to soluble fibrinogen (FITC+). Three independent experiments were performed with each clone. * $p < 0.05$, ** $p < 0.01$ versus unstimulated A20-C3G-WT cells; # $p < 0.05$, ## $p < 0.01$ versus PMA-stimulated A20-C3G-WT cells. (B) Rap1 activation time course after stimulation with 10 ng/mL PMA. Rap1-GTP level were detected by pull-down followed by western blot. Total Rap1 and β -actin expression were analyzed as references. Graphs represents the mean $\pm$ SEM of grouped data from three A20-C3G-WT and the indicated A20-C3G-Y564H clones, with at least two experiments in each case. ** $p < 0.01$ when comparing A20-C3G-WT with A20-C3G-Y564H-221 cells.

(C) Flow cytometry analysis of LFA-1 (α L β 2) (left) and VLA-4 (α 4 β 1) (right) integrin levels on the cell surface using anti-mouse APC-CD11a and anti-mouse PerCP/Cyanine5.5-CD49d conjugated antibodies, respectively. Bar charts represent the mean $\pm$ SD of the mean fluorescence intensity (MFI) from three independent experiments. **** p <0.0001. (D) Talin-1 protein levels were analyzed by western blot. A representative experiment is shown. The bar chart represents the mean $\pm$ SEM of Talin-1 levels, relativized to β -tubulin, from three independent experiments. (E, F) A20 cells were stimulated with 10 μ g/ml anti-IgM, and (E) Rac1-GTP or (F) Cdc42-GTP levels were detected at different times. Representative experiments are shown. Total Rac1, Cdc42, and β -actin levels in cell lysates were used as loading controls. Graphs represents the mean $\pm$ SEM of grouped data from three A20-C3G-WT and two A20-C3G-Y564H clones, with at least one experiment performed per clone.

Figure S5

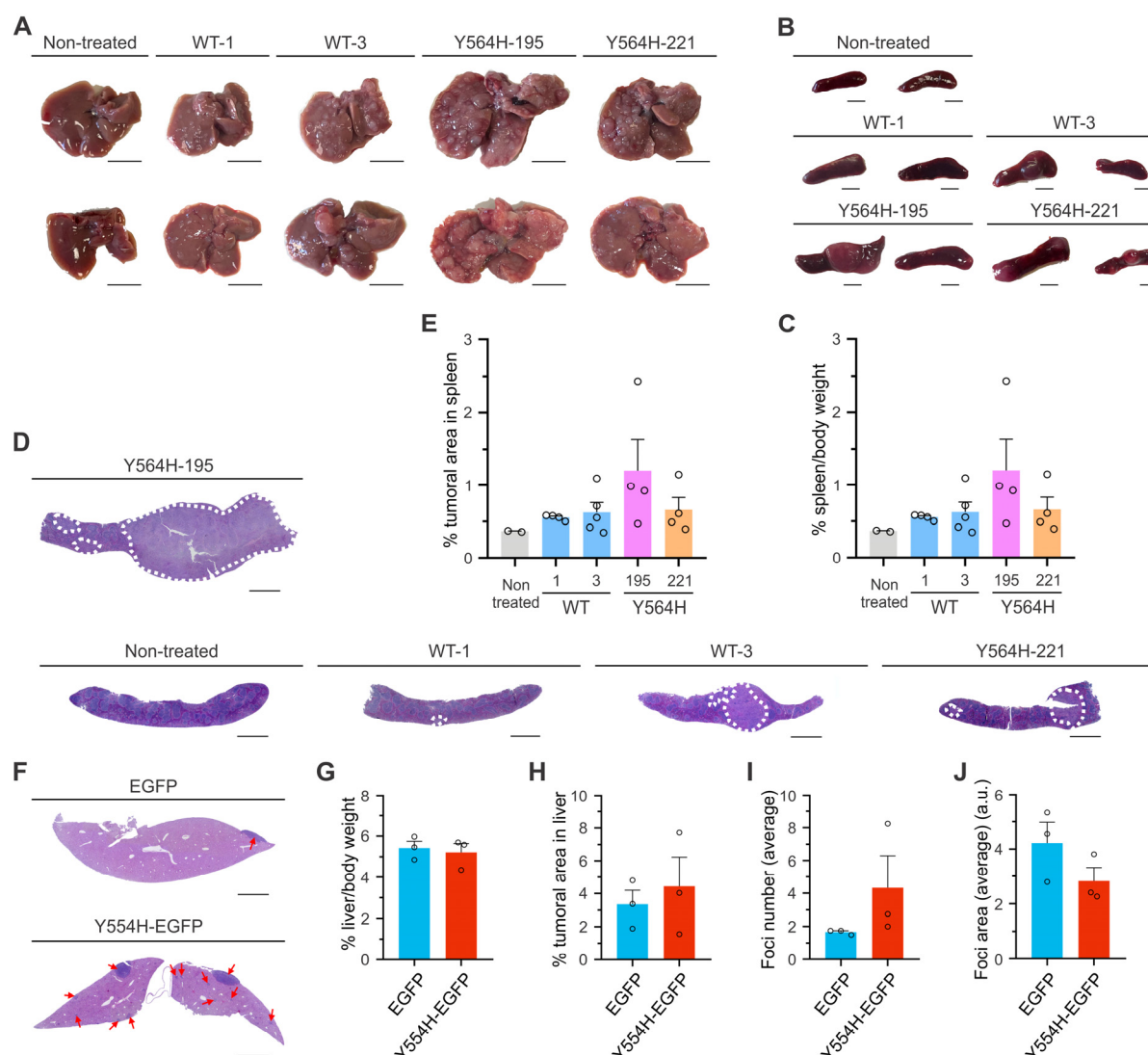


Figure S5. Analysis of tumor growth and metastasis in the liver and spleen of BALB/c mice injected with A20-C3G-Y564H cells, A20-C3G-Y554H-EGFP cells or their corresponding control cells. A total of $\sim 5 \times 10^5$ A20 cells were injected into the tail vein of BALB/c mice, and tumors were allowed to grow for 21 days. Mice were sacrificed, and liver and spleen were dissected for analysis. **(A)** Images of the livers from mice injected with A20-C3G-WT cells (clones -1 and -3) or A20-C3G-Y564H cells (clones -195 and -221) are shown. The images on the left show the livers of non-injected BALB/c mice, used as controls. Two representative images are provided for each clone, one from each independent experiment. Scale bar: 1 cm. **(B)** Images of the spleens from the same mice whose livers are shown in **(A)**. Scale bar: 0.5 cm. **(C)** Bar chart displaying the mean $\pm$ SEM of spleen weight as a percentage of total body weight. **(D)** Representative sections of spleens from mice injected with A20-C3G-WT (clones -1 and -3) or A20-C3G-Y564H cells (clones -195 and -221). A spleen section from a

non-treated BALB/c mouse is included as a control. Sections were stained with hematoxylin and eosin, with tumor areas outlined by dotted white lines. Images were captured using a Leica DM6 B microscope with a 5X objective. Scale bar: 0.2 cm. **(E)** Bar chart showing the mean $\pm$ SEM of tumor cell area as a percentage of the total spleen section area. Four mice injected with each cell clone were analyzed in four independent experiments. **(F)** Representative liver section from mice injected with A20-EGFP or A20-C3G-Y554H-EGFP cells, 21 days post-injection. Histopathological analysis was performed using hematoxylin and eosin staining. Tumor foci are indicated with red arrows. Scale bar: 0.2 cm. **(G, H)** Percentage of **(G)** liver weight to total body weight (mean $\pm$ SEM) and **(H)** tumor cell area as a percentage of the total liver section area (mean $\pm$ SEM). **(I, J)** Graphs displaying the mean $\pm$ SEM of the **(I)** average foci number and **(J)** average foci area.