

SUPPLEMENTARY FIGURES

Figure S1

Expression of miR-214 in cells, tumors and metastases.

(A-B) Correlations between miR-214 and miR-148b expression and stroma/immune cell composition in TCGA-SKCM (A) and TCGA-BRCA (B), for metastasis or primary samples, are presented as “scores”, based on different evaluation methods, in plots. Non-significant relationships: p -value >0.05 . Estimates: *EDec Immune/Stromal* = Epigenomic Deconvolution of stromal/immune percentage (1); *LUMP* = Leukocytes Unmethylation for Purity, which averages 44 non-methylated immune-specific CpG sites (2); *IHC* = estimated by image analysis of haematoxylin and eosin stained slides produced by the Nationwide Children's Hospital Biospecimen Core Resource (3); *ABSOLUTE* = based on somatic copy-number data (4). **(C-H)** miR-214 expression levels were measured for the indicated cells, primary tumors or metastasis by qRT-PCR analysis. **(C-D)** Human tumor and stroma cells; **(E-F)** mouse B16-F10 or human MA-2 melanoma cells in culture or their derived total primary subcutaneous (sc) tumors or dissected metastases (mets) following sc or tail vein injections in syngeneic or immunosuppressed mice, respectively; **(G-H)** B16-F10 or MA-2 cells not treated (NT) or co-cultured with MEFs (G) or hMSCs (H) or treated with Conditioned Medium (CM) derived from hMSCs (H) for 24 h. Results are shown as fold changes (mean $\pm$ SD of triplicates) relative to the median of miR-214 levels, normalized on U6 or U44 RNA. At least 2 independent experiments (with triplicates) were performed and either representative results (C, E, F) or pooled results of two independent experiments (D, G) are shown. MEFs=murine embryo fibroblasts; hMSCs=human mesenchymal stem cells; SD=standard deviation. * $P\leq0.05$; ** $P\leq0.01$; *** $P\leq0.001$.

Figure S2

Generation and characterization of the miR-214 overexpressing mouse model.

(A) Schematic overview of the strategy used to generate a conditional miR-214 overexpressing (miR-214^{over}) mouse model. The sequence of miR-214 precursor was cloned downstream of a constitutive active strong promoter (CAGGS) followed by a transcriptional STOP element (STOP) flanked by two LoxP sites (miR-214^{over} plasmid). This construct was specifically targeted in the 3' UTR of the Collagene alpha 1 (ColA1) genomic locus in previously engineered mouse Embryonic Stem Cells (mESCs) using the flippase (FLP) site-directed recombination technology. Recombinant ESCs were positively selected using hygromycin, driven by a PGK (phosphoglycerate kinase) promoter. (B) miR-214 expression levels were measured in miR-214^{wt} and miR-214^{over} total embryos (E12.5) by qRT-PCR analysis for the indicated number (n) of animals. (C) miR-214 expression levels were evaluated in miR-214^{wt} (n=5) and miR-214^{over} (n=3) total embryos (E12.5) by Northern Blot analysis using specific miR-214 and U6 small nucleolar RNA digoxigenin-labeled LNA probes. (D) *In situ* hybridization analysis for miR-214 expression on Formalin-Fixed Paraffin Embedded (FFPE) miR-214^{wt} and miR-214^{over} total embryos (E12.5), using digoxigenin-labeled LNA probes. Representative images of miR-214 expression in vertebrae (a, b) and liver (c, d) are shown; scale bar: 200 μ m. (E) Results were calculated as fold changes (mean $\pm$ SD of triplicates) relative to the median of miR-214 levels for the indicated number (n) of embryos, normalized on U6 small nucleolar RNA levels. SD=standard deviation. *P $\leq$ 0.05; **P $\leq$ 0.01; ***P $\leq$ 0.001.

Figure S3

***In vivo* tumor growth and metastatic dissemination.**

(A) *In vivo* tumor growth and metastatic dissemination of B16-F10 cells injected subcutaneously in miR-214^{wt} and miR-214^{over} syngeneic mice, 45 days post-inoculation. Tumors were removed 15 days post-injection. Relative primary tumor weight and relative number of lung metastases are presented as mean $\pm$ SEM for the indicated number (n) of mice. Representative images of lung metastases are also shown. Arrows point to metastases. (B) Analysis of S100b expression in a lung lobe derived

from B16-F10-xenografts grown in miR-214^{wt} and miR-214^{over} mice (as in A) by qRT-PCR analysis. Results were calculated as fold changes (mean $\pm$ SD of triplicates) relative to the median of samples for the indicated number (n) of mice, normalized on 18S rRNA levels. SD=standard deviation. *P $\leq$ 0.05; **P $\leq$ 0.01; ***P $\leq$ 0.001.

Figure S4

Characterization of miR-214-overexpressing stroma cells.

(A-B) miR-214 expression levels were measured in miR-214^{wt} and miR-214^{over}-derived CAFs (A) or MEFs (B) by qRT-PCR analysis. (C) Representative pictures of miR-214^{wt} or miR-214^{over} CAFs isolated from mice; scale bar: 150 μ m. (D) Evaluation of N-cadherin, E-cadherin, α -SMA and loading control protein expression in different miR-214^{wt} or miR-214^{over} CAF preparations by Western Blot analysis. (E-F) Proliferation assays for miR-214^{wt} or miR-214^{over} CAFs (E) or MEFs (F). Results are indicated as mean $\pm$ SEM of proliferation rate, measured by Optical Density (OD) at 0h–72h (t0-t72). (G-H) Transwell Migration assays for miR-214^{wt} or miR-214^{over} CAFs (G) or MEFs (H). (A, B) Results were calculated as fold changes (mean $\pm$ SD of triplicates) relative to the median of miR-214 levels for the indicated number (n) of MEF and CAF preparations, normalized on U6 small nucleolar RNA levels. For (E-H) Results are shown as mean $\pm$ SEM of the area covered by migrated cells. All experiments were performed as triplicates and representative results are shown. CAFs=cancer associated fibroblasts; MEFs=murine embryo fibroblasts; SEM= standard error of mean; α -SMA=*alpha* Smooth Muscle Actin. *P $\leq$ 0.05; **P $\leq$ 0.01; ***P $\leq$ 0.001.

Figure S5

Characterization of miR-214-sponged stromal cells.

(A-B) miR-214 expression levels assessed in NIH3T3 mouse fibroblasts (A) or HS5 human bone marrow stromal cells (B) previously transduced with a miR-214sponge (miR-214^{sponge}) or an empty (control) expressing vector by qRT-PCR analysis. Results are shown as fold changes (mean $\pm$ SD of

triplicates) relative to controls, normalized on U6. At least 2 independent experiments (with triplicates) were performed and representative results are shown. **(C-D)** Proliferation assays for the same NIH3T3 and HS5 used in (A-B). Results are indicated as mean $\pm$ SEM of the proliferation, measured by Optical Density (OD) at 0h–72h (t0-t72). **(E-F)** Wound healing assays for the same NIH3T3 and HS5 used in (A-B). Migration speed was evaluated after 6h (E) or 24h (F) as mean $\pm$ SEM of distance/time covered by migrated cells ($\mu\text{m/h}$ of 10 pictures/duplicates). **(C-F)** All experiments were performed at least twice (with duplicates/triplicates) and representative results are shown. SEM= standard error of mean. SD=standard deviation. * $P\leq 0.05$; ** $P\leq 0.01$; *** $P\leq 0.001$.

Fig. S6

Stroma miR-214 enhances transendothelial migration of tumor cells.

(A-B) Transendothelial migration through a HUVECs monolayer on top of a porous membrane for B16-F10 and EO771 cells pretreated for 24h with Extracellular Vesicles (EVs) derived from miR-214^{wt} and miR-214^{over} CAFs. Top: representative pictures of HUVECs monolayer before plating and B16-F10 and EO771 migrated cells. Bottom: results expressed as mean $\pm$ SEM of the area (pixels) covered by migrated cells. 2 independent experiments (with triplicates) were performed and pooled results are shown. CAFs=cancer associated fibroblasts; SEM= standard error of mean. * $P\leq 0.05$; ** $P\leq 0.01$; *** $P\leq 0.001$.

Fig. S7

Stroma miR-214 does not influence tumor cell proliferation.

(A-F) Proliferation assays for B16-F10 (A, C) or EO771 (B, D) or MA-2 (E) or 4175-TGL (F) cells pretreated for 24h with Conditioned Medium (CM) derived from either miR-214^{wt} or miR-214^{over} CAFs (A-B) or alternatively from NIH3T3 (C-D) or HS5 cells (E-F) previously transduced with a miR-214sponge (miR-214^{sponge}) or an empty (control) expressing vector. All results are presented as mean $\pm$ SEM of the proliferation rate, measured by Optical Density (OD) at 0h–72h (t0-t72). At least

two independent experiments as triplicates were performed, and representative results are shown. MEFs=murine embryo fibroblasts; CAFs=cancer associated fibroblasts; SEM= standard error of mean. * $P \leq 0.05$; ** $P \leq 0.01$; *** $P \leq 0.001$.

Figure S8

Characterization of Extracellular Vesicles derived from stroma cells.

(A-H) Evaluation of number and size (nm) of Extracellular Vesicles (EVs) derived from miR-214^{wt}, miR-214^{over} or miR-214^{ko} MEFs and CAFs (A-D) or alternatively from NIH3T3 and HS5 (E-H) cells previously transduced with a miR-214sponge (miR-214^{sponge}) or an empty (control) expressing vector by Nanosight analysis. Values are shown as mean $\pm$ SEM for the number (n) of indicated preparations. **(I-K)** Evaluation of EV and cell of origin markers (CD63, CD140a CD140b, CD73, CD44, α -SMA, Fap) in EVs derived from the indicated cells by FACS analysis shown as percentage (%) of signal intensity referred to mean $\pm$ SEM. **(L-M)** Electron microscopy images of EVs derived from miR-214^{wt}, miR-214^{over} or miR-214^{ko} CAFs (L) or alternatively from HS5 cells previously transduced with a miR-214sponge (miR-214^{sponge}) or an empty (control) expressing vector (M); scale bar: 100 nm. MEFs=murine embryo fibroblasts; CAFs=cancer associated fibroblasts; SEM= standard error of mean. * $P \leq 0.05$; ** $P \leq 0.01$; *** $P \leq 0.001$.

Figure S9

miR-214 and miR-148b expression in tumor cells is modulated by stroma cell conditioned medium.

(A-H) miR-214 and miR-148b expression levels measured in mouse B16-F10 (A-C) and EO771 (B-D) or human MA-2 (E-G) or 4175-TGL (F-H) cells pre-treated for 24h with the Conditioned Medium (CM) derived from miR-214^{wt} and miR-214^{over} MEFs or alternatively from HS5 cells previously transduced with a miR-214sponge (miR-214^{sponge}) or an empty (control) expressing vector by qRT-PCR analysis. All results are shown as fold changes (mean $\pm$ SD of triplicates) relative to the median

of samples, normalized on U6. At least 2 independent experiments (with triplicates) were performed and pooled results of two or three independent experiments are shown. MEFs=murine embryo fibroblasts; SD=standard deviation. * $P \leq 0.05$; ** $P \leq 0.01$; *** $P \leq 0.001$.

Figure S10

The transfer of stroma miR-214 to tumor cells affects a pro-metastatic pathway.

(A-C) miR-214 or miR-148b expression levels in EO771 (A) and 4175-TGL (B, C) cells pre-treated for 24-48h with Extracellular Vesicles (EVs) derived from NIH3T3 (A) or HS5 (B, C) previously transduced with a miR-214sponge (miR-214^{sponge}) or an empty (control) expressing vector by qRT-PCR analysis. All results are shown as fold changes (mean $\pm$ SD of triplicates) relative to the median of samples, normalized on U6. At least 2 independent experiments (with triplicates) were performed and pooled results of two or three independent experiments are shown. (D-F) TFAP2C, ALCAM and ITGA5 protein expression in B16-F10 (D), EO771 (E) and 4175-TGL (F) pre-treated for 24-48h with Extracellular Vesicles (EVs) derived from miR-214^{over} or miR-214^{ko} CAFs or alternatively from NIH3T3 or HS5 cells previously transduced with a miR-214sponge (miR-214^{sponge}) or an empty (control) expressing vector. Protein modulations were calculated relative to the indicated controls, normalized on the loading control and expressed as percentages. CAFs=cancer associated fibroblasts; SD=standard deviation * $P \leq 0.05$; ** $P \leq 0.01$; *** $P \leq 0.001$.

Figure S11

Metastasis formation in a miR-214^{ko} background.

(A) *In vivo* tumor growth and metastatic dissemination of EO771 cells injected subcutaneously in miR-214^{wt} and miR-214^{ko} syngeneic mice, 30 days post-inoculation. Relative primary tumor weight (measured in grams) and number of lung metastases is shown in the graphs as mean $\pm$ SEM, for the indicated number of mice (n). Representative images of lung metastases are also shown. (B) Analysis of S100b expression in a lung lobe derived from B16-F10-xenografts grown in miR-214^{wt} and miR-

214^{ko} mice by qRT-PCR analysis. Results were calculated as fold changes (mean $\pm$ SD of triplicates) relative to the median of samples for the indicated number (n) of mice, normalized on 18S rRNA levels. SD=standard deviation. *P $\leq$ 0.05; **P $\leq$ 0.01; ***P $\leq$ 0.001.

Figure S12

miR-214-rich extracellular vesicles (EVs) promote melanoma cell dissemination.

(A) Transendothelial migration through a HUVECs monolayer on top of a porous membrane for B16-F10 cells pretreated for 24h with EVs derived from miR-214^{wt}, miR-214^{over} or miR-214^{ko} CAFs. Top: representative pictures of HUVECs monolayer before plating and B16-F10 migrated cells. Bottom: results expressed as mean $\pm$ SEM of the area (pixels) covered by migrated cells. 3 independent experiments (with quadruplicates) were performed and pooled results are shown. **(B)** miR-214 expression levels in blood-isolated EVs of miR-214^{ko} mice 15 minutes after tail vein injections of EVs derived from miR-214^{wt}, miR-214^{over} and miR-214^{ko} CAFs, measured by qRT-PCR analysis. Results are shown as fold changes (mean $\pm$ SD of triplicates) relative to the median of all samples, normalized on U6 for the indicated number (n) of mice. EVs= extracellular vesicles; CAFs=cancer associated fibroblasts; SEM= standard error of mean; SD=standard deviation. *P $\leq$ 0.05; **P $\leq$ 0.01; ***P $\leq$ 0.001.

Figure S13

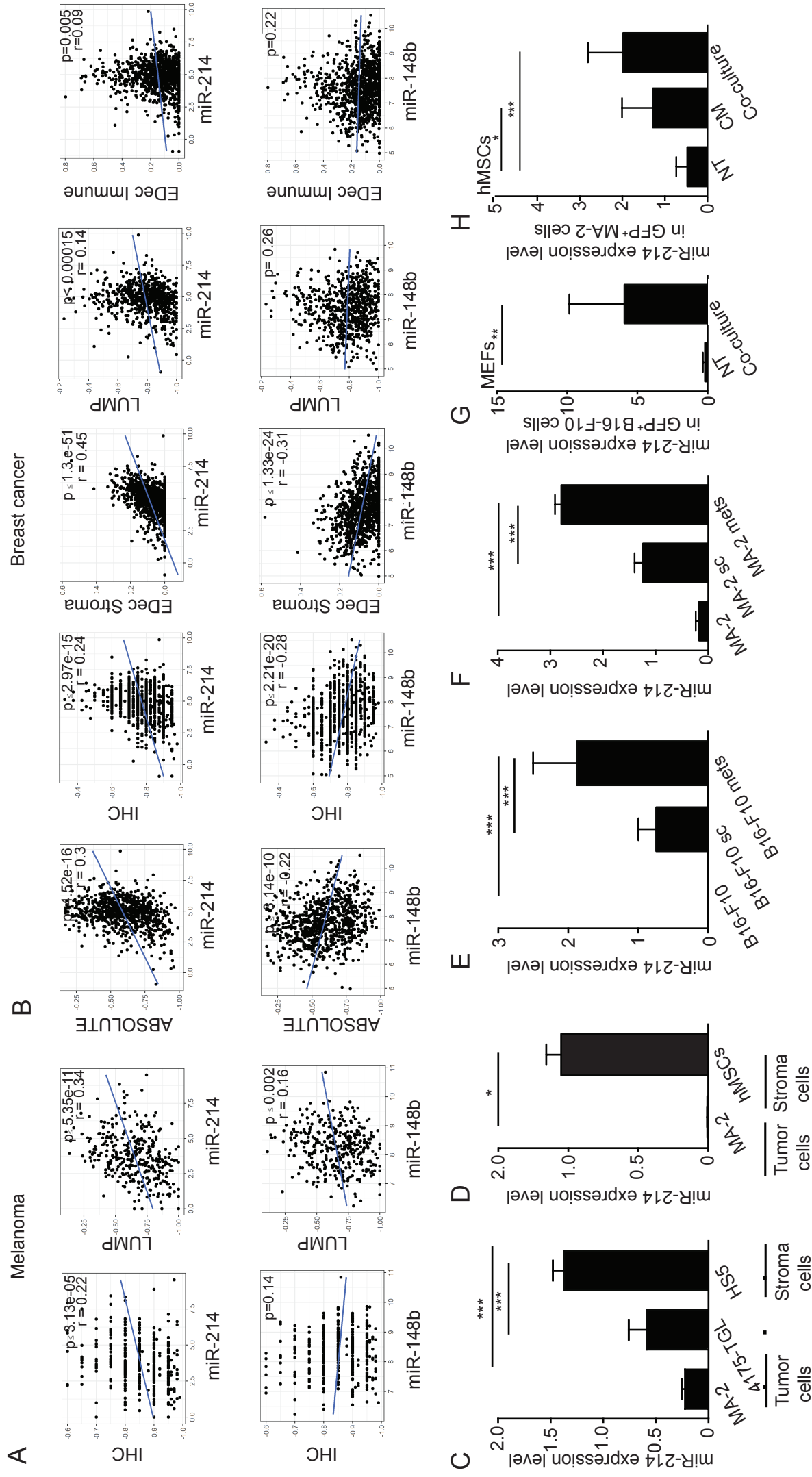
Correlations between miR-214 and IL6/STAT3 axis in human tumors.

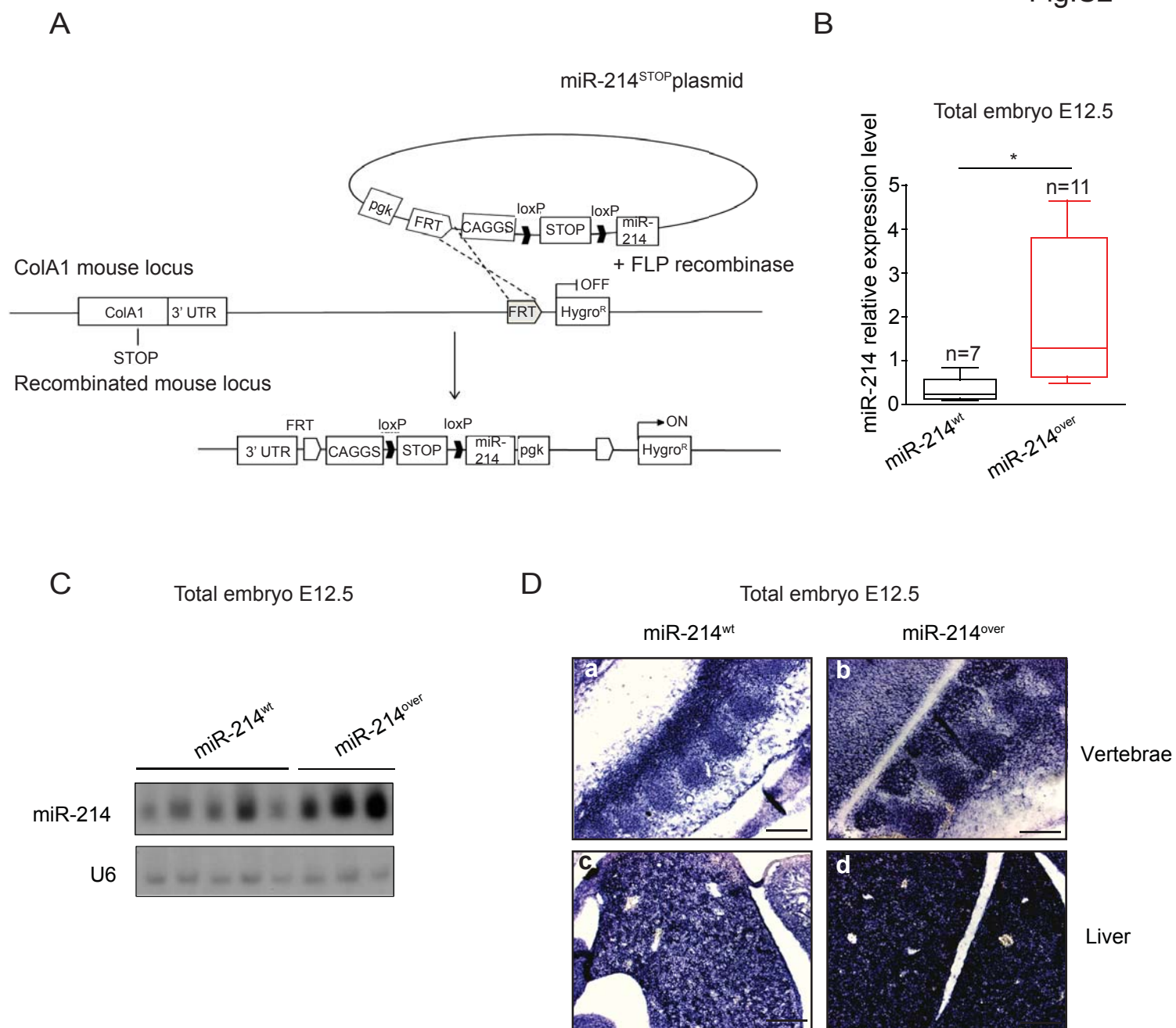
(A-B) Correlations between miR-214 expression and different IL-6 and Stat3-related signatures in TCGA-SKCM (A) or TCGA-BRCA (B) datasets. Samples are presented in plots. Non-significant relationships: p-value>0.05. *AZARE_sig* (5); *DAUER_sig* (6); *ALVAREZ_sig* (7); *TH_sig* (8); *Jak/Stat* (9, 10) M11564; *Stat3_sig_down* (8).

References

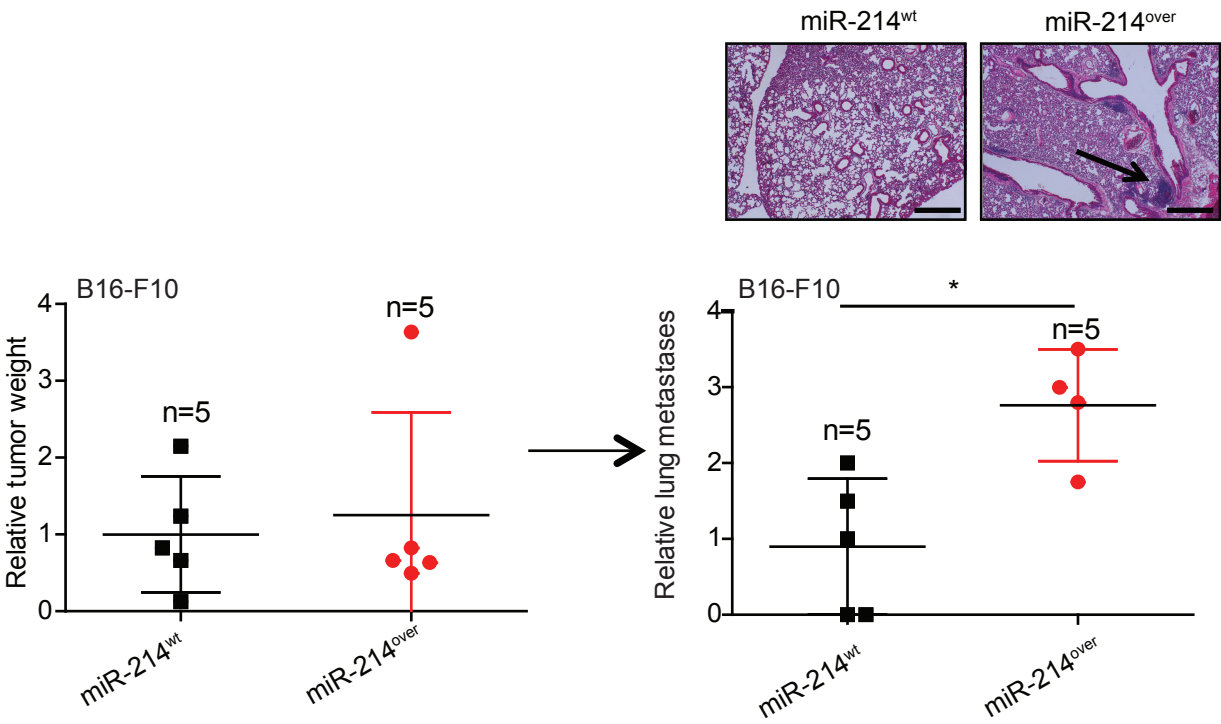
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Fig. S1





A



B

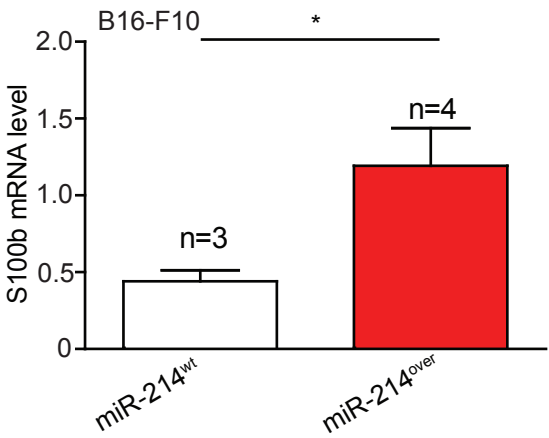


Fig. S4

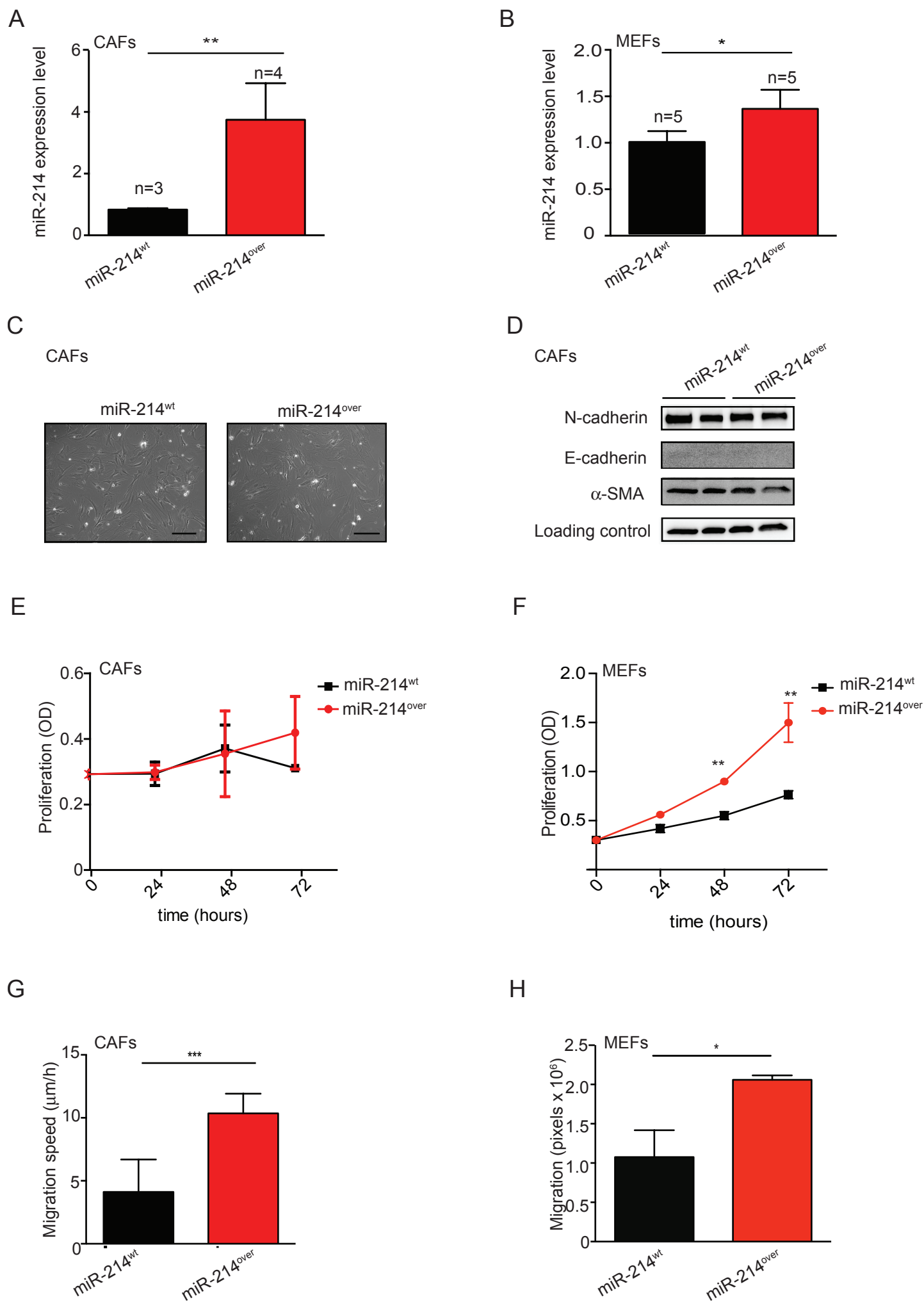
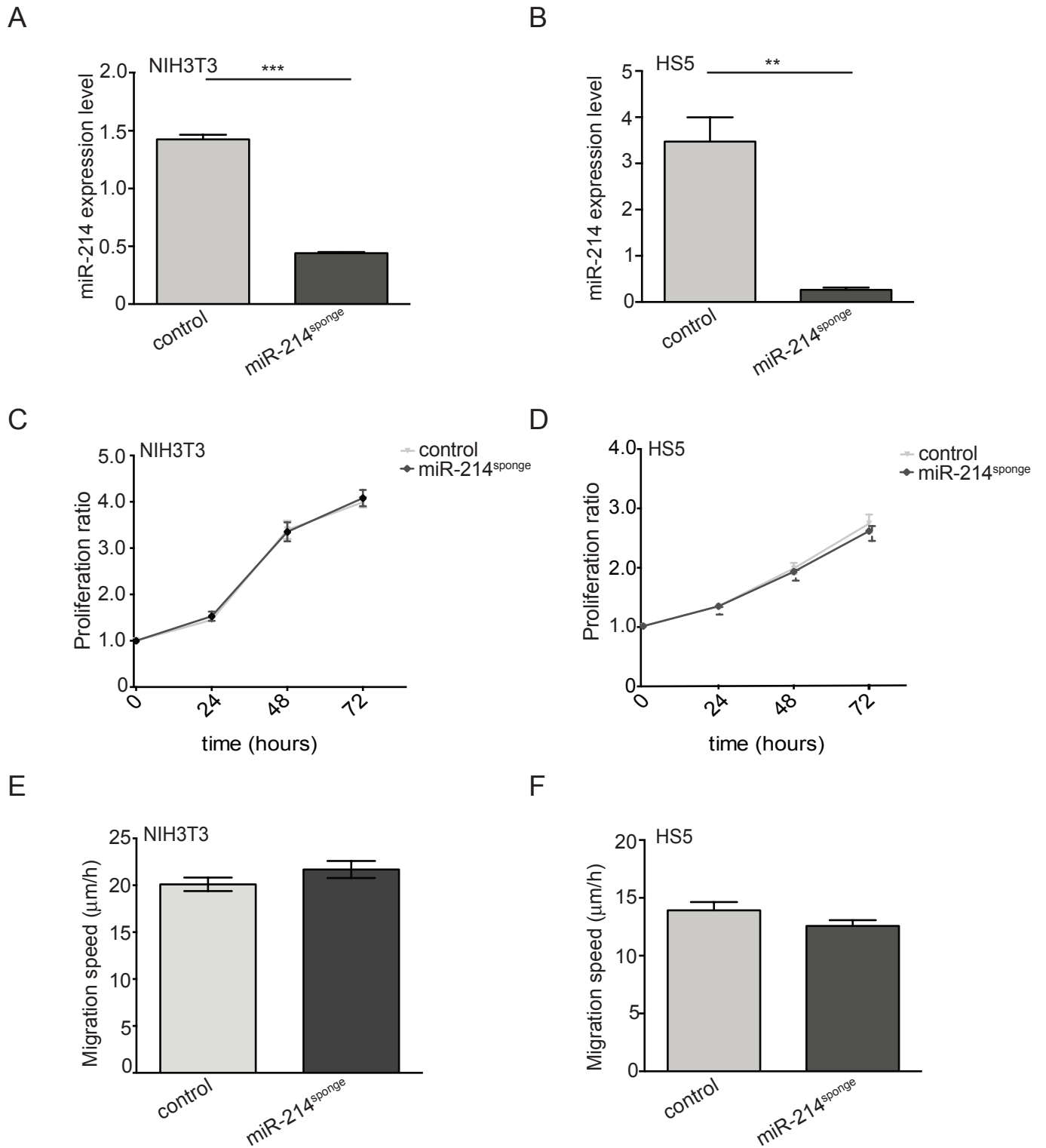


Fig. S5



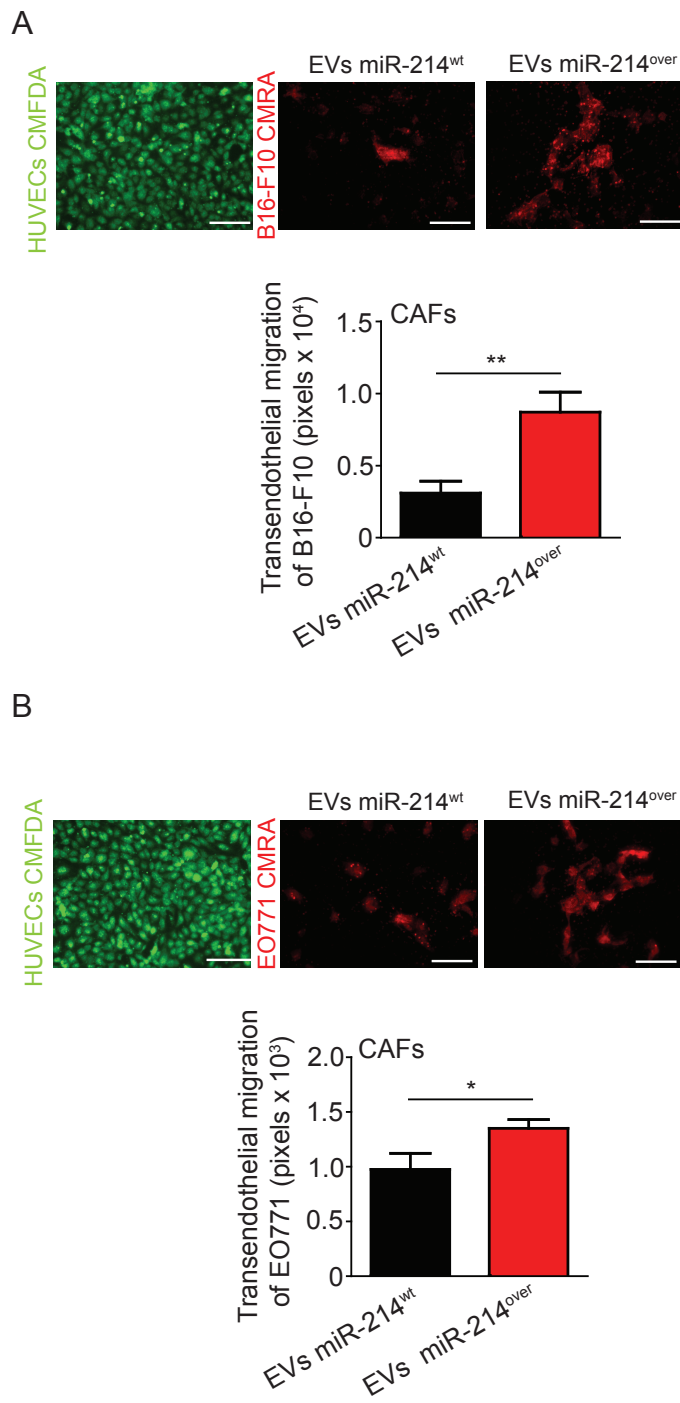


Fig. S7

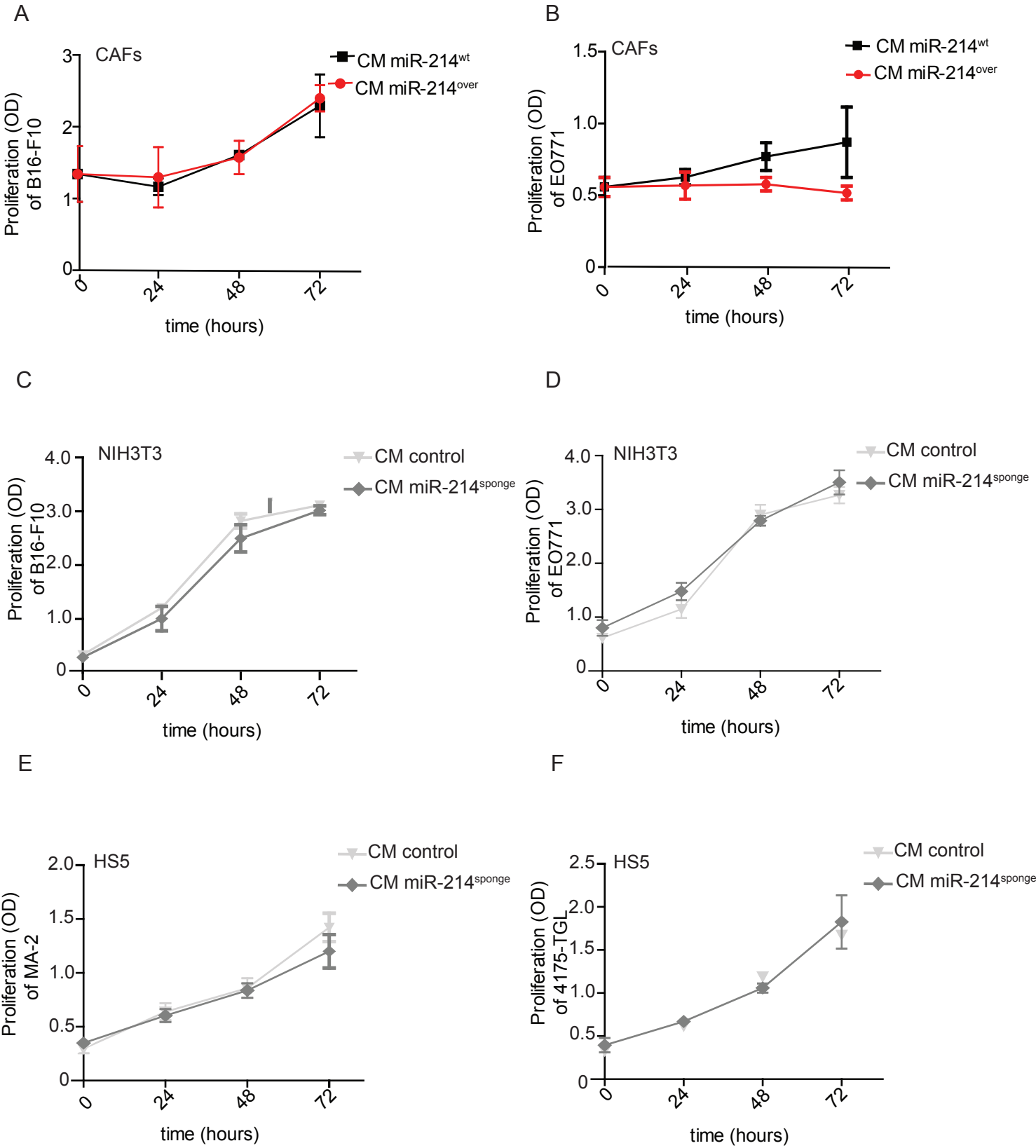


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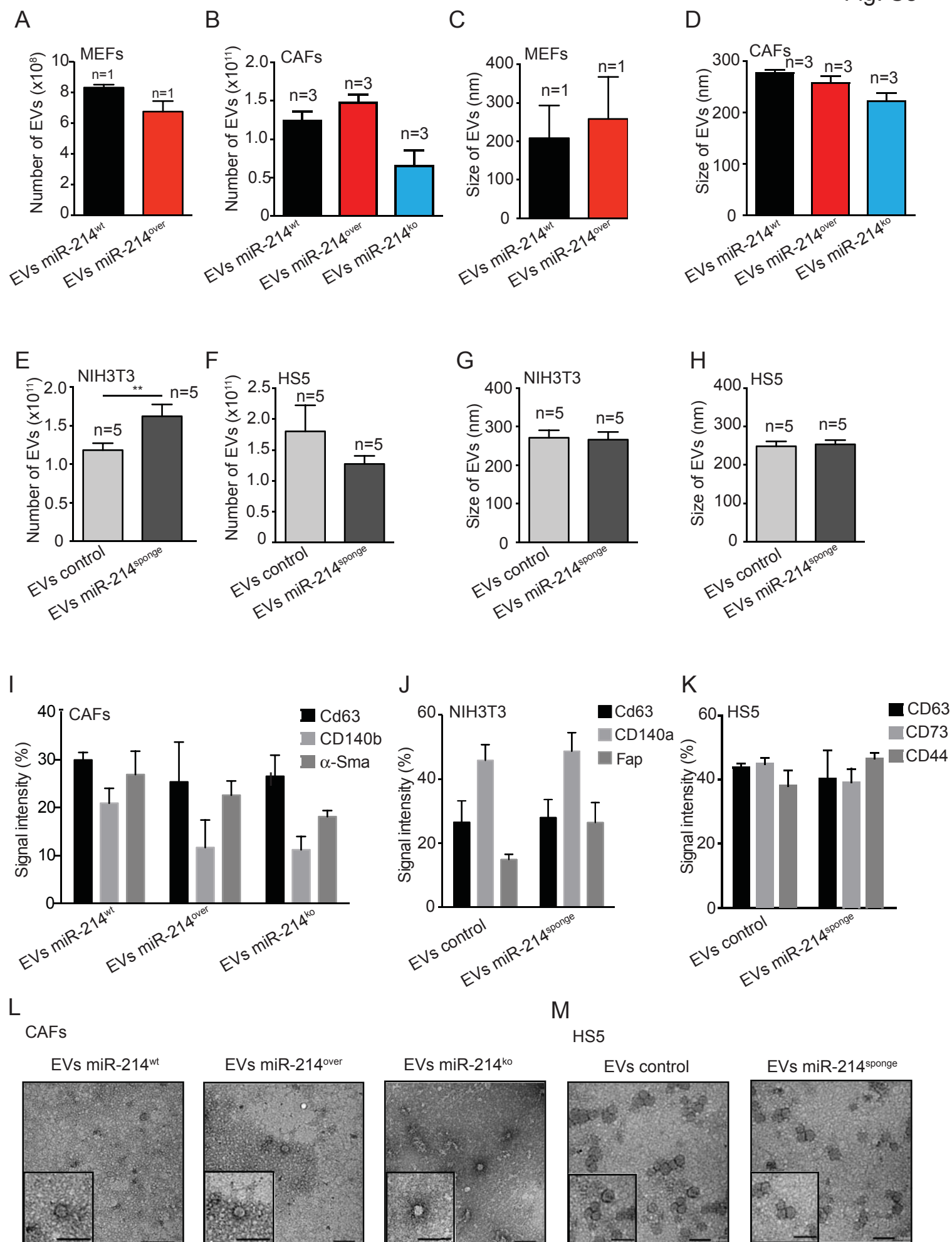
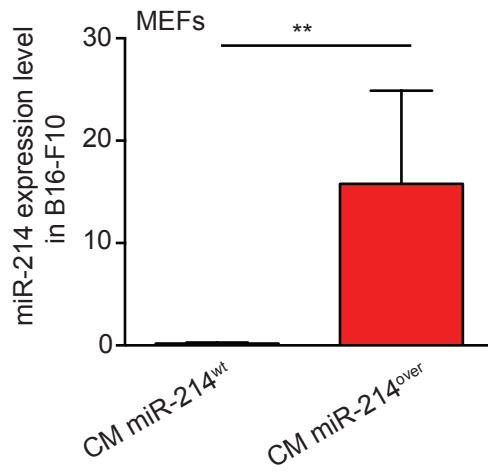
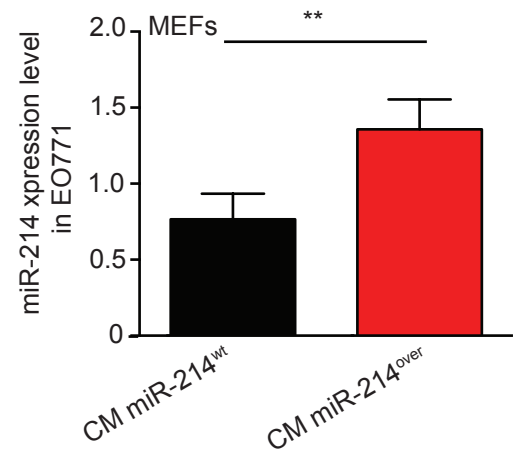


Fig. S9

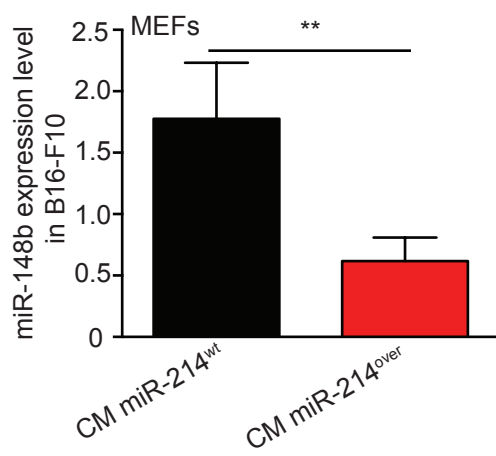
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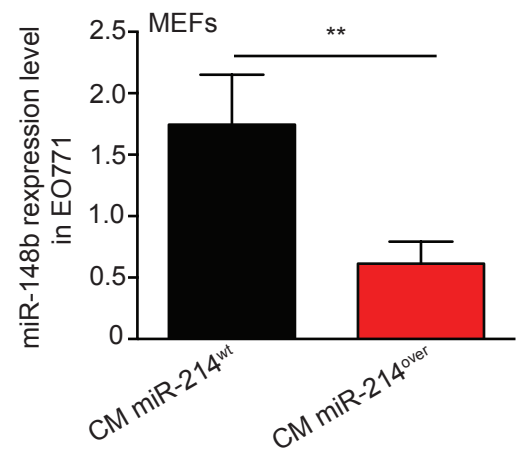
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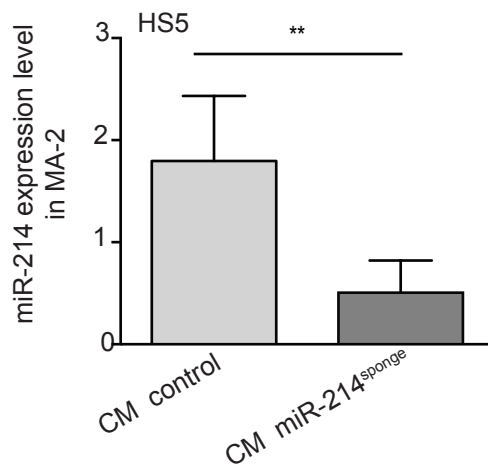
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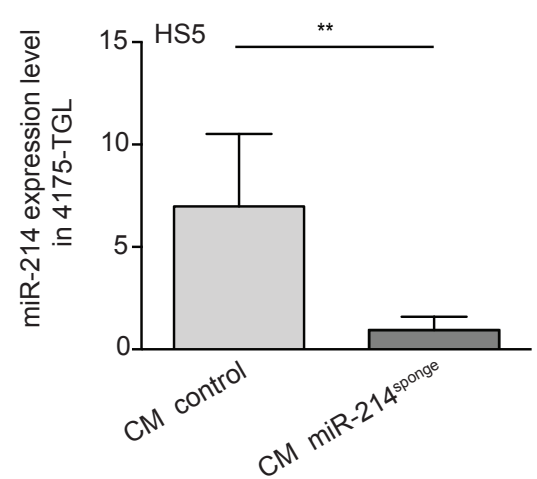
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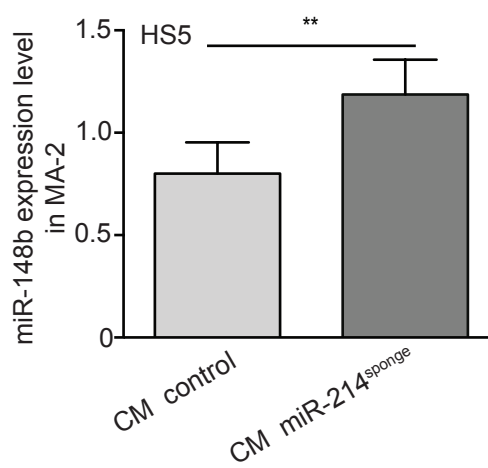
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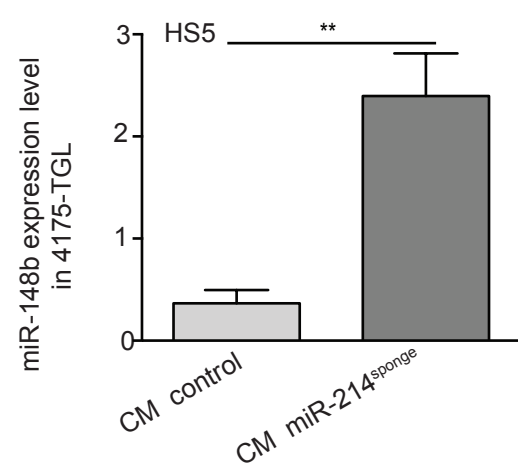
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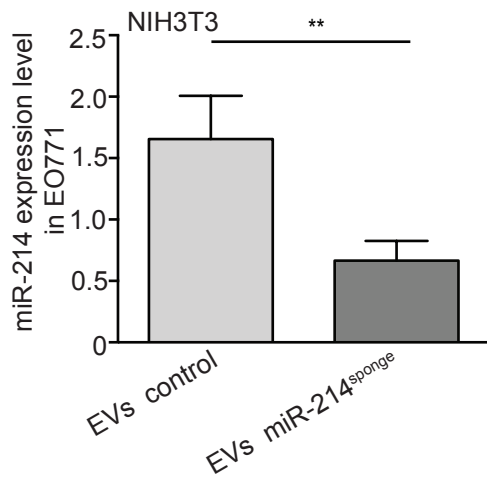
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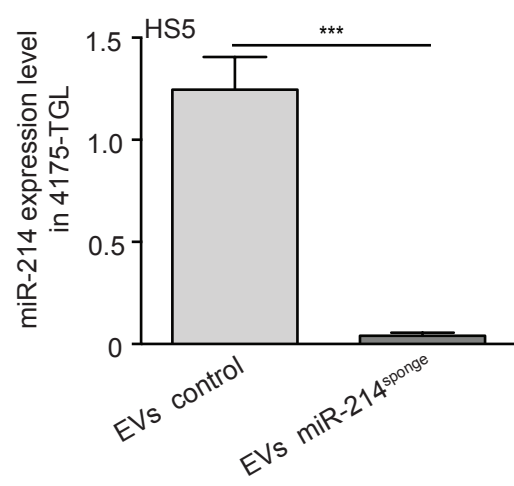
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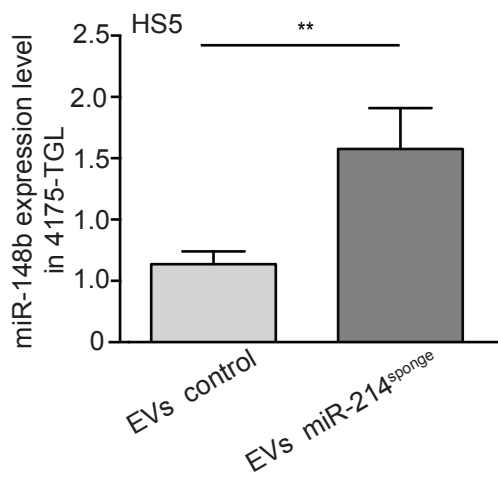
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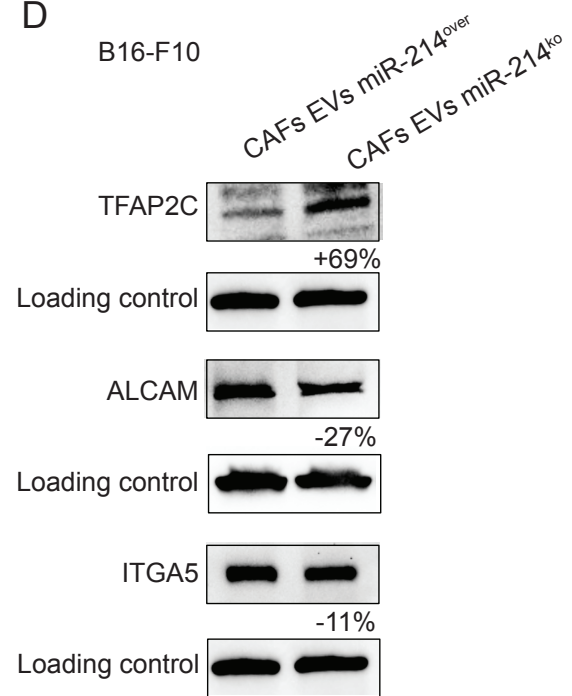
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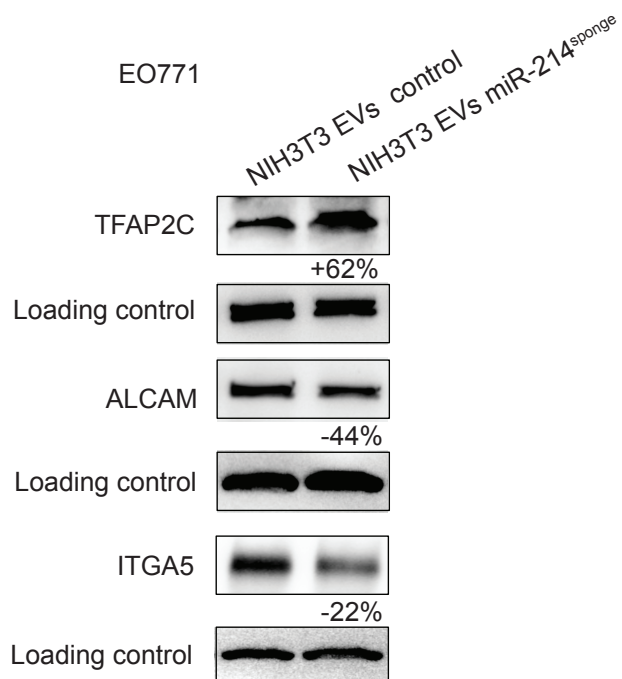
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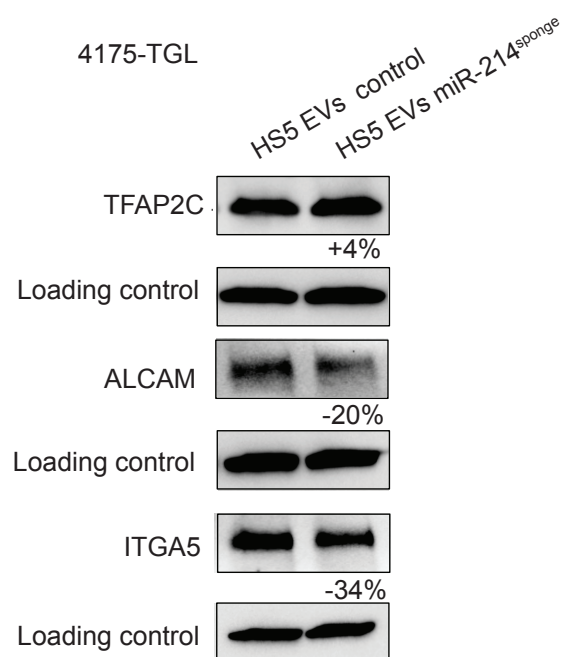
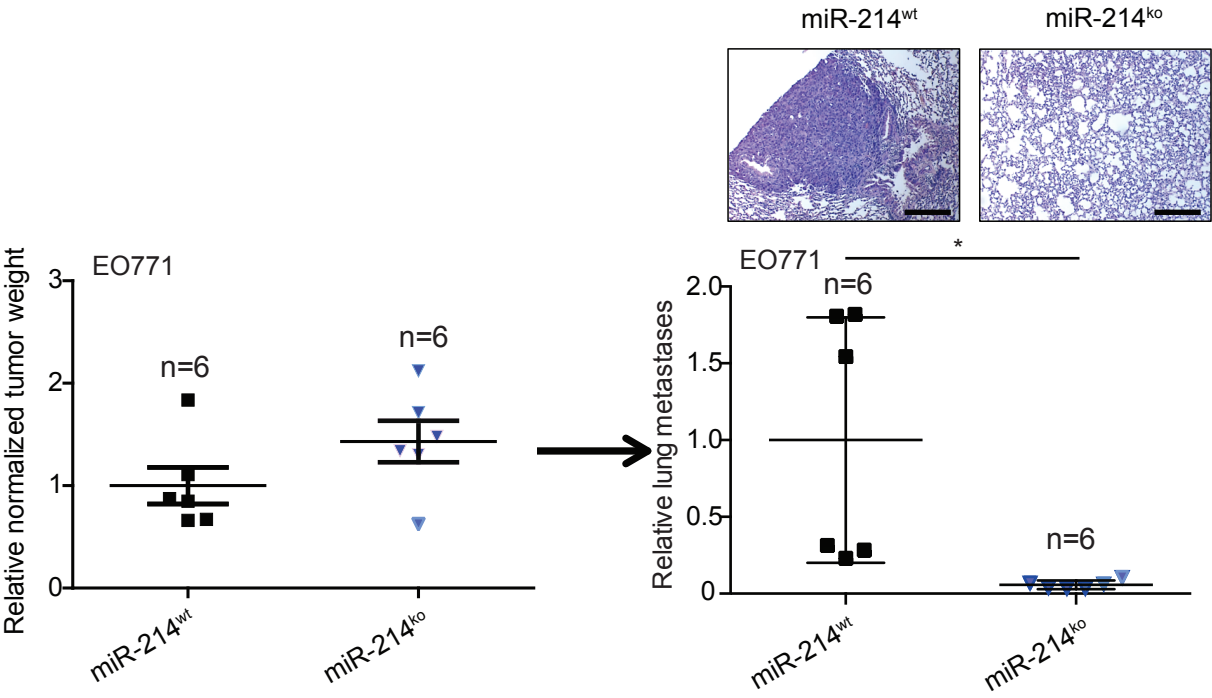
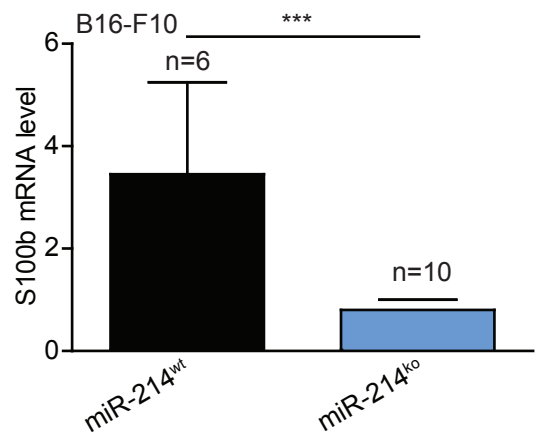


Fig. S11

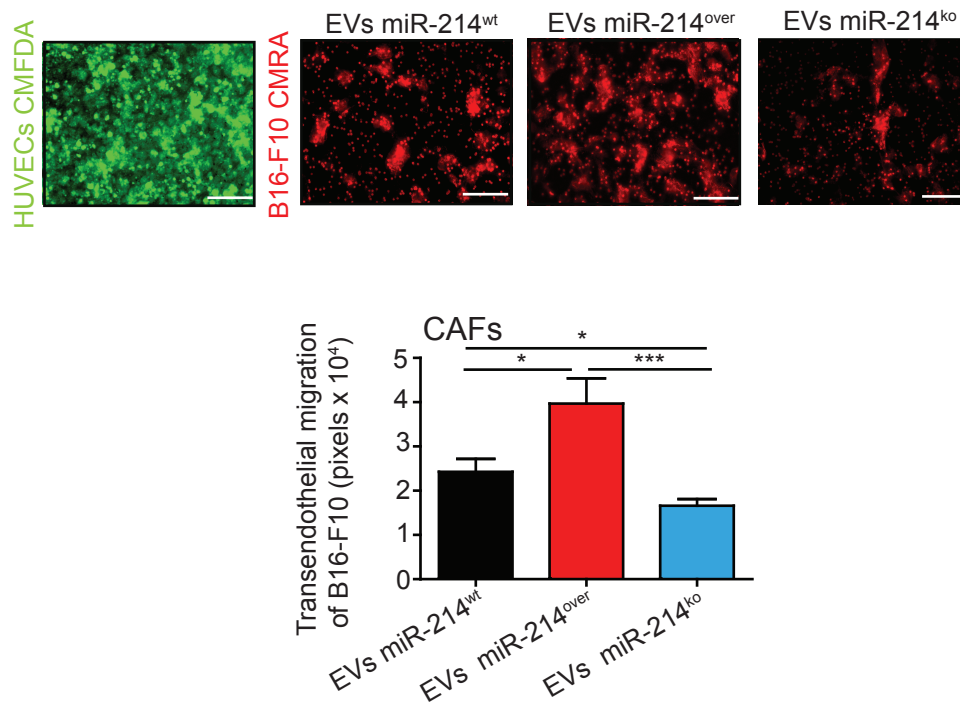
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B



A



B

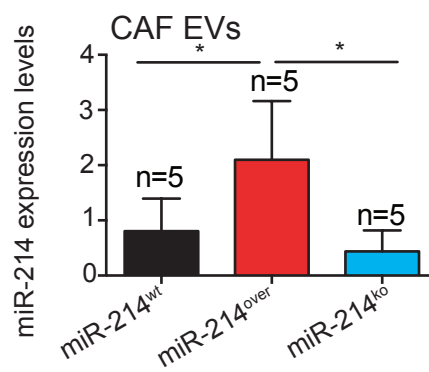
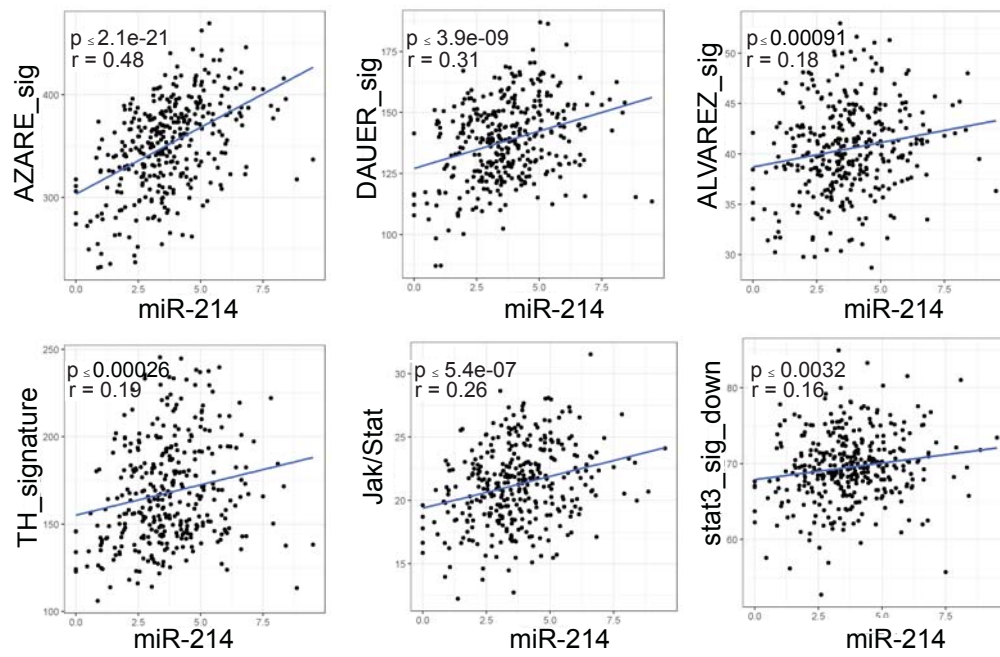


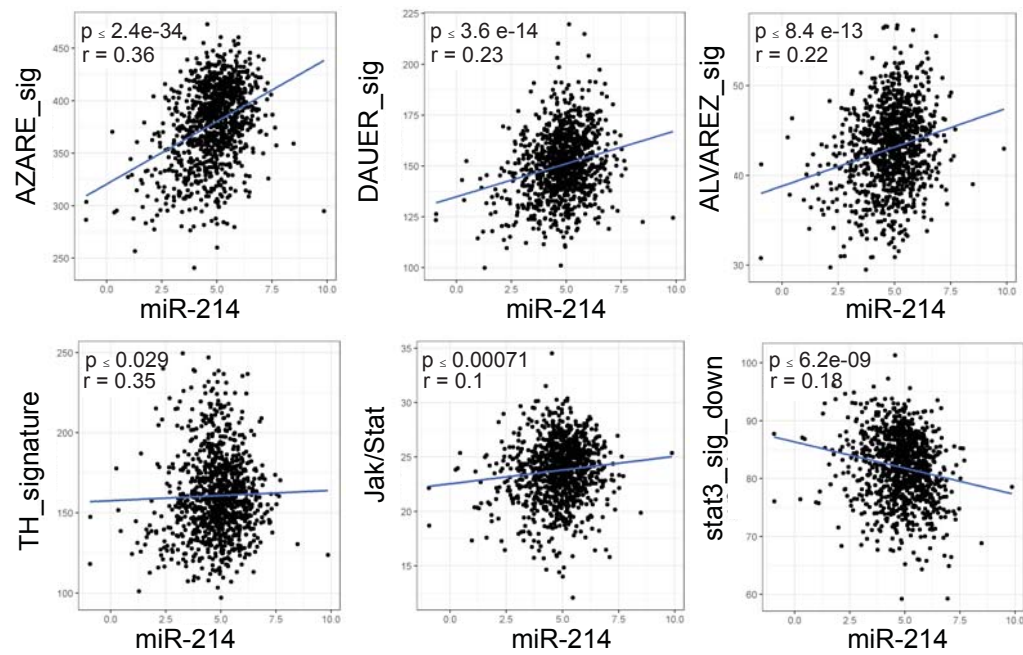
Fig. S13

A



Melanoma

B



Breast cancer