

Figure S1

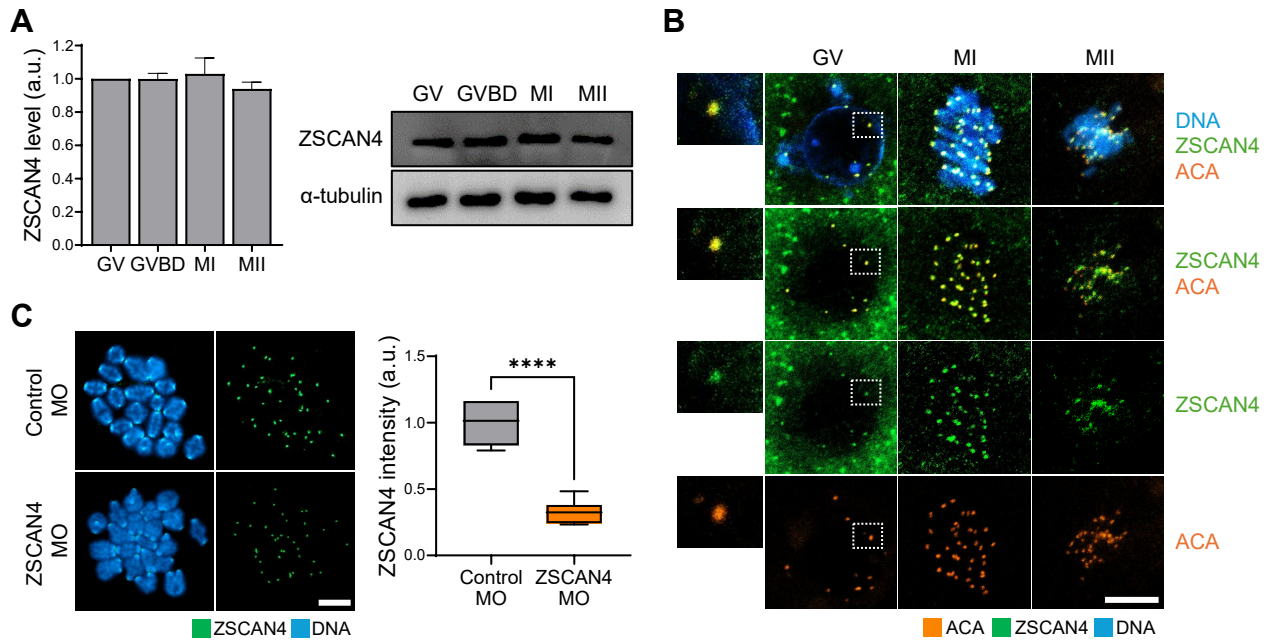


Figure S1. Expression and localization of ZSCAN4 in mouse oocytes. (A) Immunoblot analysis of ZSCAN4 expression during meiotic maturation. Oocytes were collected after 0, 2, 8, and 16 h of culture, corresponding to the GV, GVBD, MI, and MII stages, respectively, and subjected to immunoblot analysis with ZSCAN4 antibody. Each lane includes 50 oocytes, and α -tubulin was used as a loading control. Data are presented as the mean $\pm$ SEM of two independent experiments. (B) Subcellular localization of ZSCAN4 during mouse oocyte meiotic maturation. Immunofluorescence staining was performed using ZSCAN4 and ACA antibodies. Scale bar, 10 μ m. (C) Representative images of chromosome spreads at the MI stage, stained with ZSCAN4 antibody. Oocytes were injected with control MO or ZSCAN4 MO for ZSCAN4 depletion. Scale bar, 10 μ m. Quantification of ZSCAN4 intensity. Data are presented as the mean $\pm$ SEM of three independent experiments. ****p<0.00001.

Figure S2

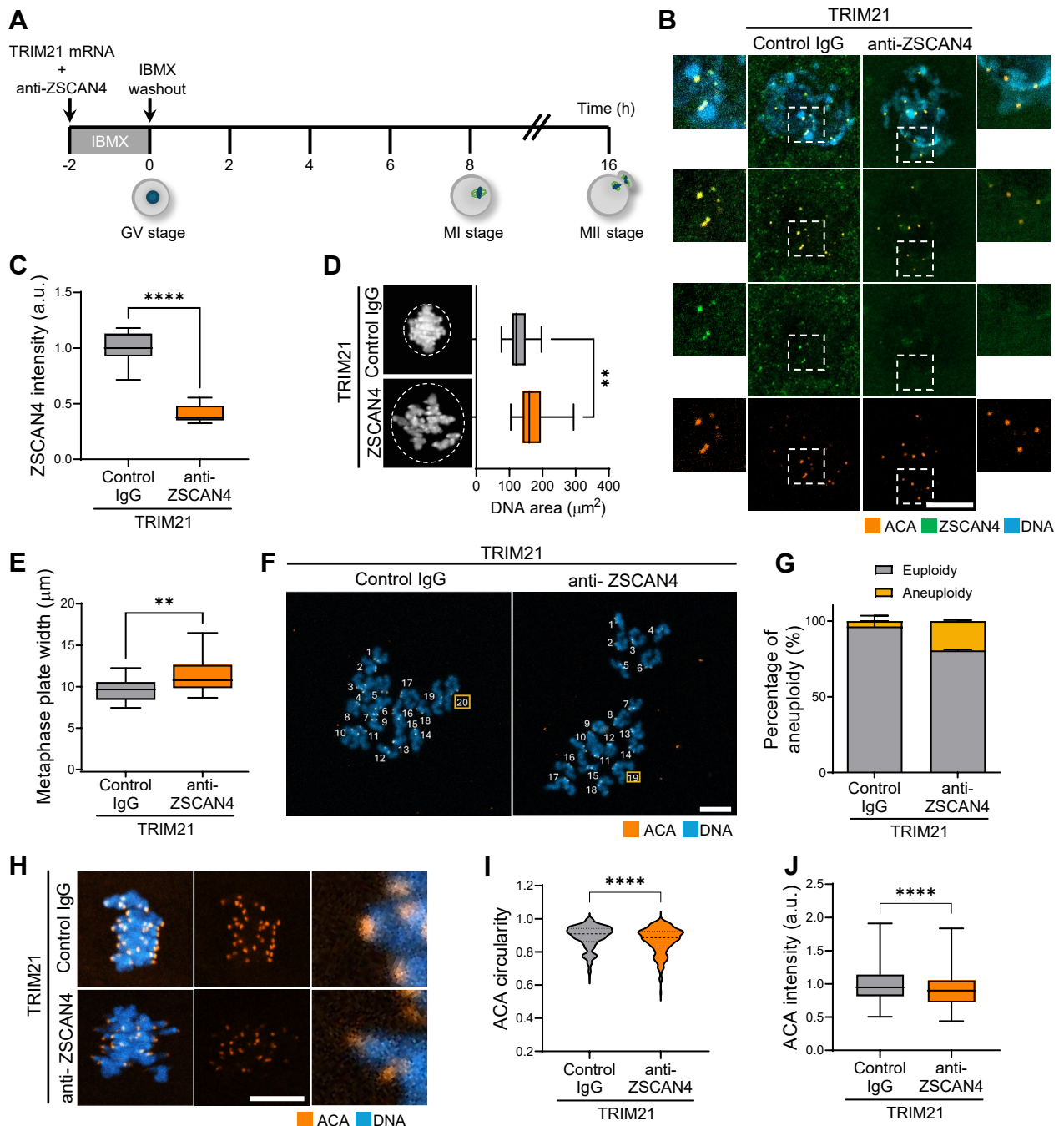


Figure S2. Trim-Away-mediated depletion of ZSCAN4 in mouse oocytes. (A) Schematic diagram depicting the Trim-Away approach for depleting ZSCAN4. GV oocytes were injected with TRIM21 mRNA and ZSCAN4 antibody; control oocytes were injected TRIM21 mRNA and IgG. After 2 h recovery in IBMX-supplemented medium, oocytes were released from IBMX and cultured for 8 h and 16 h to reach the MI and the MII stage, respectively. (B) Representative images of control and ZSCAN4-depleted GV oocytes stained with ZSCAN4 and ACA antibodies. Scale bar, 10 μ m. (C) Quantification of ZSCAN4 intensity. Data are presented as the mean $\pm$ SEM of two independent experiments. **** p <0.00001. (D) Representative images and quantification of DNA area at the MI stage. DNA area was measured as the area of a circle that includes all the aligned chromosomes. Data are presented as the mean $\pm$ SEM of two independent experiments. ** p <0.001. (E) Quantification of metaphase plate width. Data are presented as the mean $\pm$ SEM of two independent experiments. ** p <0.001. (F) Representative images of chromosome spread stained with ACA antibody in control and ZSCAN4-depleted oocytes. (G) Quantification of percentage of aneuploidy. Data are presented as the mean $\pm$ SEM of two independent experiments. (H) Representative images of control and ZSCAN4-depleted oocytes stained with ACA antibody. (I-J) Quantification of ACA circularity and ACA intensity. Data are presented as the mean $\pm$ SEM of three independent experiments. **** p <0.00001.

Figure S3

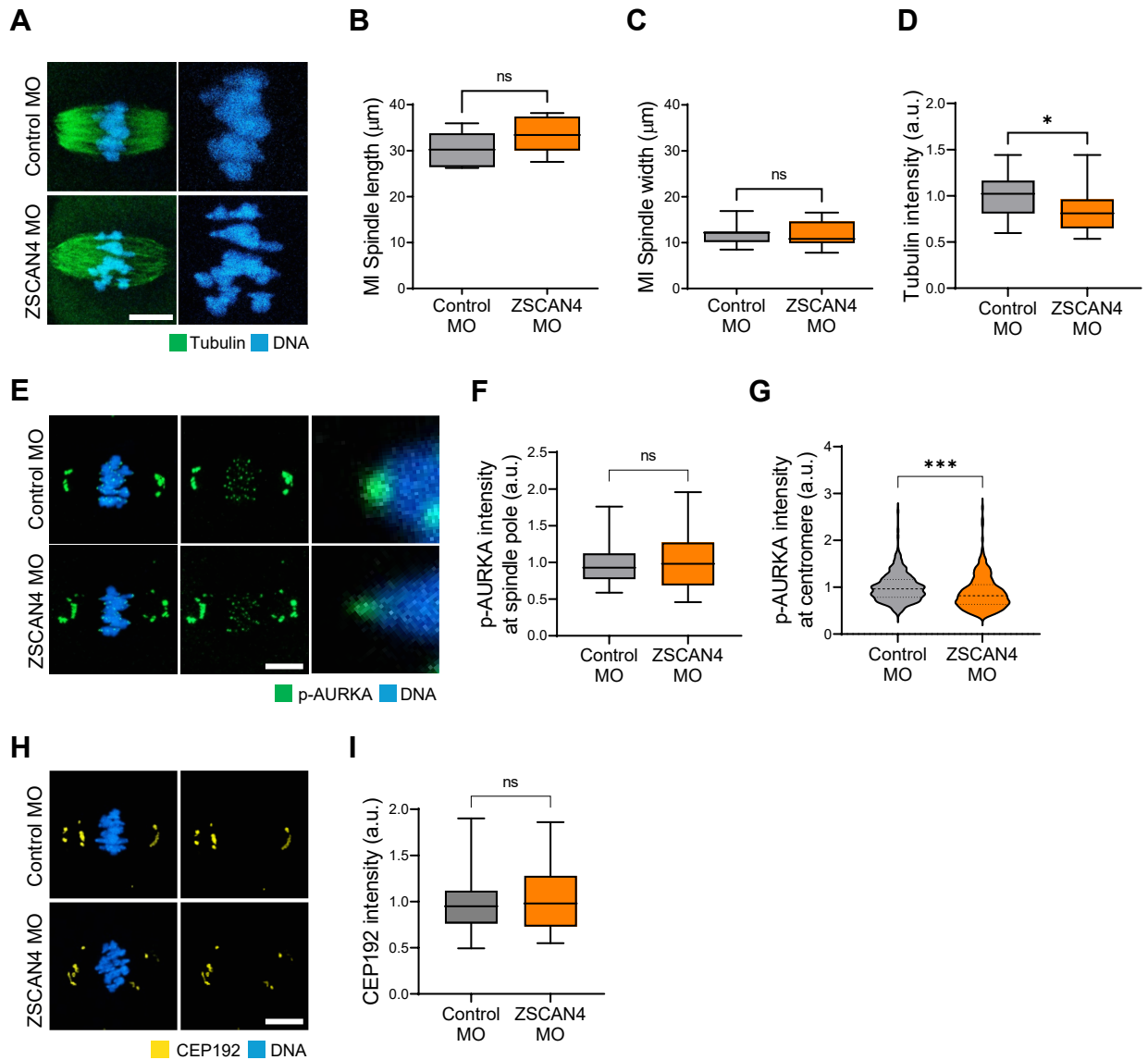


Figure S3. No discernible difference in spindle morphology and pole integrity after ZSCAN4 depletion. (A) Representative images of control and ZSCAN4-depleted oocytes stained with acetylated alpha-tubulin antibody. Scale bar, 10 μm . (B–D) Quantification of spindle length, width, and intensity of control and ZSCAN4-depleted oocytes at the MI stage. Spindle length was measured as the distance between the two spindle poles, and spindle width was measured as the widest part of the barrel-shaped spindle. Data are presented as the mean $\pm$ SEM of three independent experiments. ns, not significant. * $p < 0.05$. (E) Representative images of control and ZSCAN4-depleted oocytes stained with phosphorylated aurora kinase A (p-AURKA) antibody. Scale bar, 10 μm . (F–G) Quantification of p-AURKA intensity at the spindle pole and centromere, respectively. Data are presented as the mean $\pm$ SEM of two independent experiments. ns, not significant. *** $p < 0.0001$. (H) Representative images of control and ZSCAN4-depleted oocytes stained with CEP192 antibody. Scale bar, 10 μm . (I) Quantification of CEP192 intensity at the spindle pole. Data are presented as the mean $\pm$ SEM of two independent experiments. ns, not significant.

Figure S4

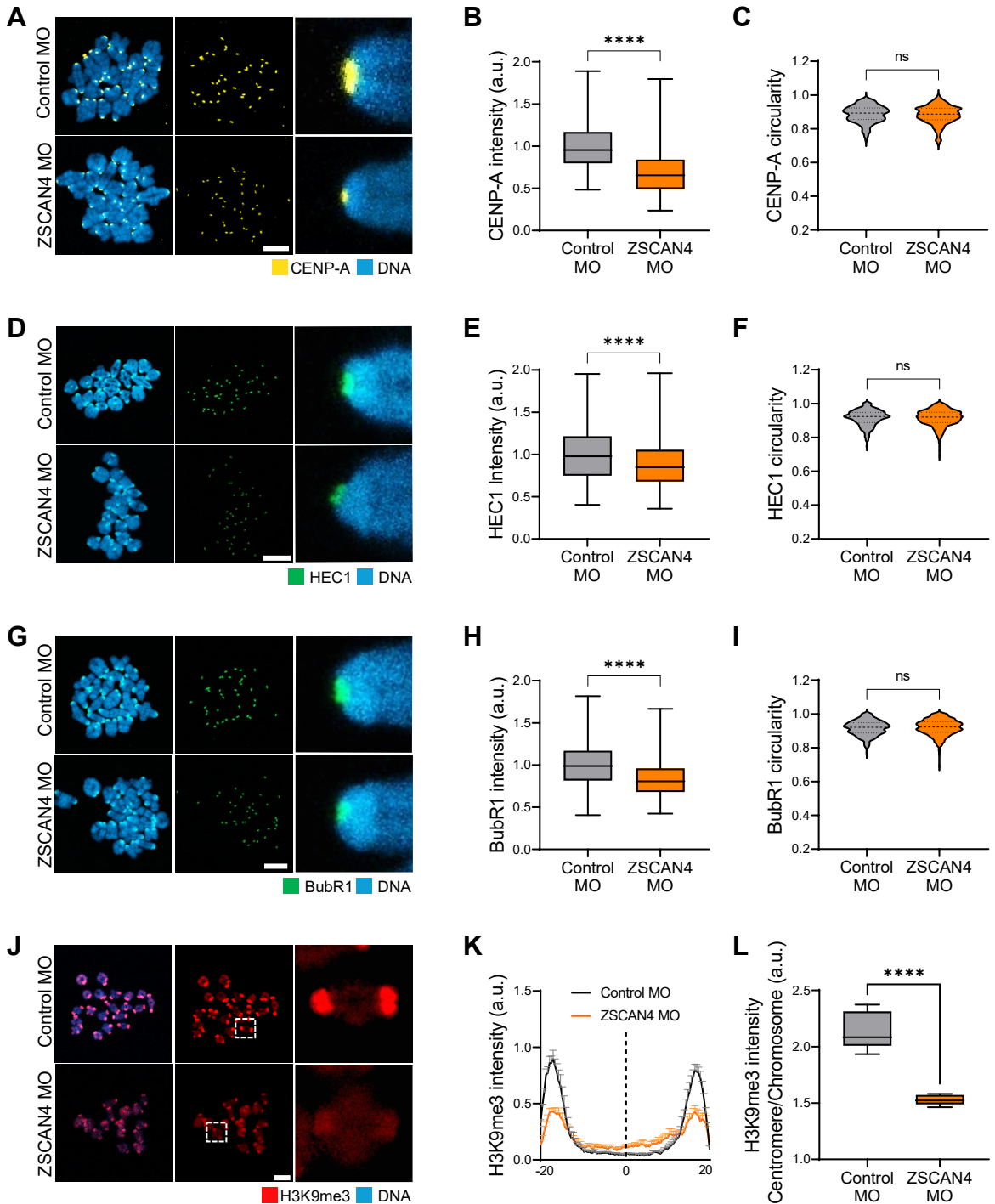


Figure S4. Decreases in CENP-A, HEC1, BubR1, and H3K9me3 after ZSCAN4 depletion. (A, D, G) Representative chromosome spread images of control and ZSCAN4-depleted oocytes stained with CENP-A (or HEC1 or BubR1) at the MI stage. Scale bar, 10 μ m. (B, E, H) Quantification of CENP-A (or HEC1 or BubR1) intensity. Data are presented as the mean $\pm$ SEM of two independent experiments. **** $p < 0.00001$. (C, F, I) Quantification of CENP-A (or HEC1 or BubR1) circularity. Data are presented as the mean $\pm$ SEM of two independent experiments. ns, not significant. (J) Representative chromosome spread images of control and ZSCAN4-depleted oocytes stained with H3K9me3 antibody. Scale bar, 10 μ m. (K) Plot graph depicting H3K9me3 intensity from one chromosome end (-20) to the opposite end (20). Ten representative chromosomes from control and ZSCAN4-depleted oocytes were selected to depict the plot. (L) Quantification of the ratio intensity of H3K9me3 at centromeres over each chromosome (centromere/chromosome). Data are presented as the mean $\pm$ SEM of two independent experiments. **** $p < 0.00001$.

Figure S5

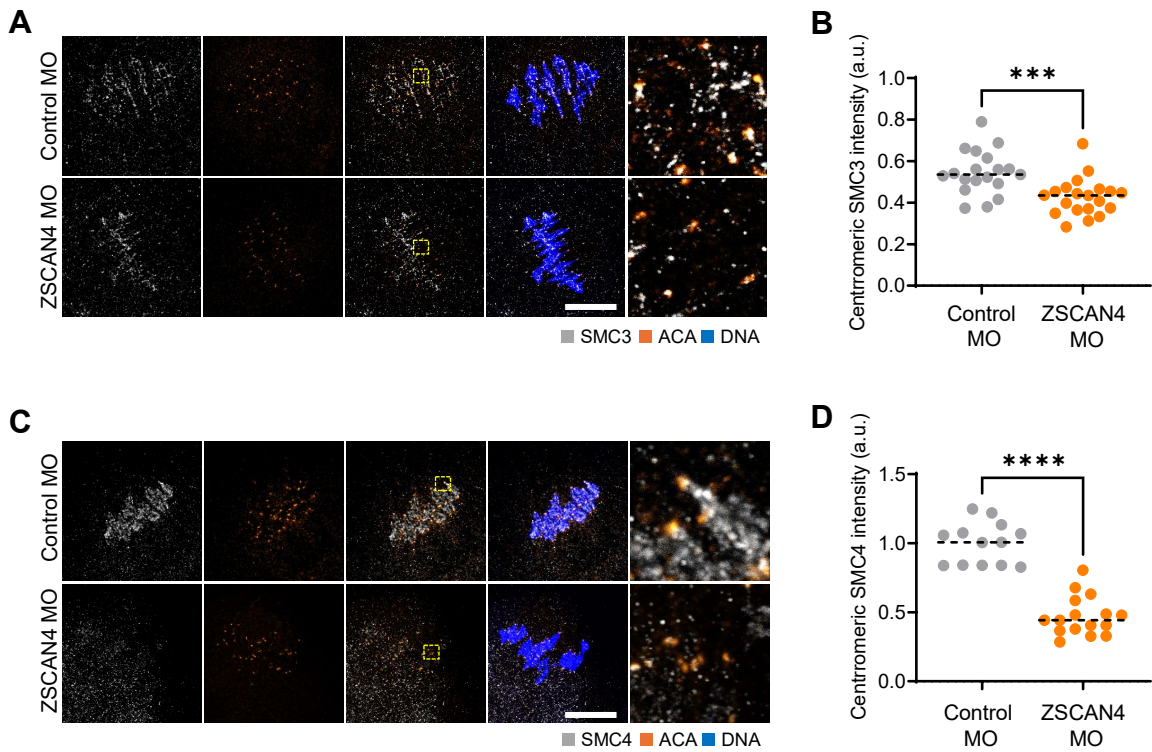


Figure S5. Decreases in SMC3 and SMC4 levels after ZSCAN4 depletion. (A, C) Representative images of control and ZSCAN4-depleted oocytes stained with SMC3 or SMC4 antibody. Regions outlined by yellow box are enlarged and shown in right panel. Scale bar, 10 μ m. (B, D) Quantification of SMC3 or SMC4 intensity. Data are presented as the mean $\pm$ SEM of two independent experiments. *** $p < 0.0001$, **** $p < 0.00001$.

Figure S6

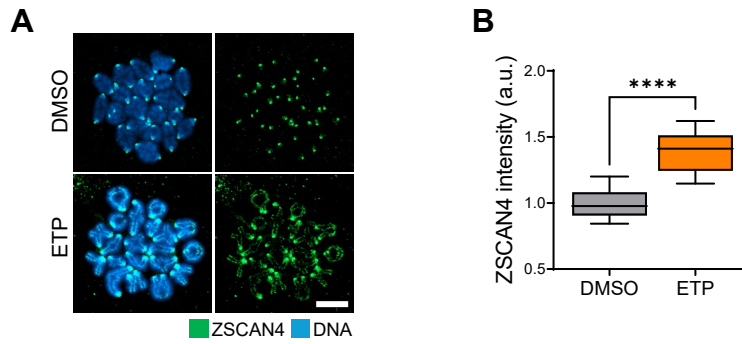


Figure S6. Increases in ZSCAN4 levels after DNA damage induction. (A) Representative images of chromosome spread stained with ZSCAN4 and ACA antibodies in DMSO- or ETP-treated oocytes. Scale bar, 10 μ m. (B) Quantification of ZSCAN4 intensity. Data are presented as the mean $\pm$ SEM of three independent experiments. **** $p < 0.00001$.

Figure S7

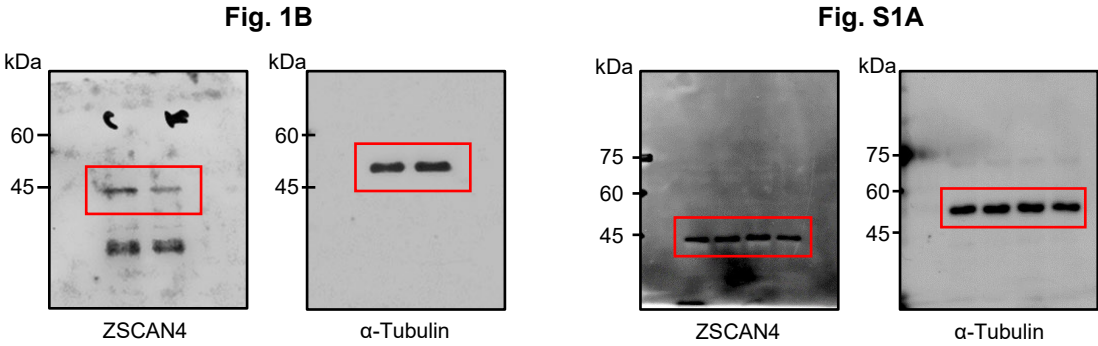


Figure S7. Full-length immunoblot images. Uncropped full-length images of Western blotting membranes presented in Figure 1B and the Supplementary Figure S1A.