

Supplementary Information

Engineered Self-Regulating Macrophages for Targeted Anti-inflammatory Drug Delivery

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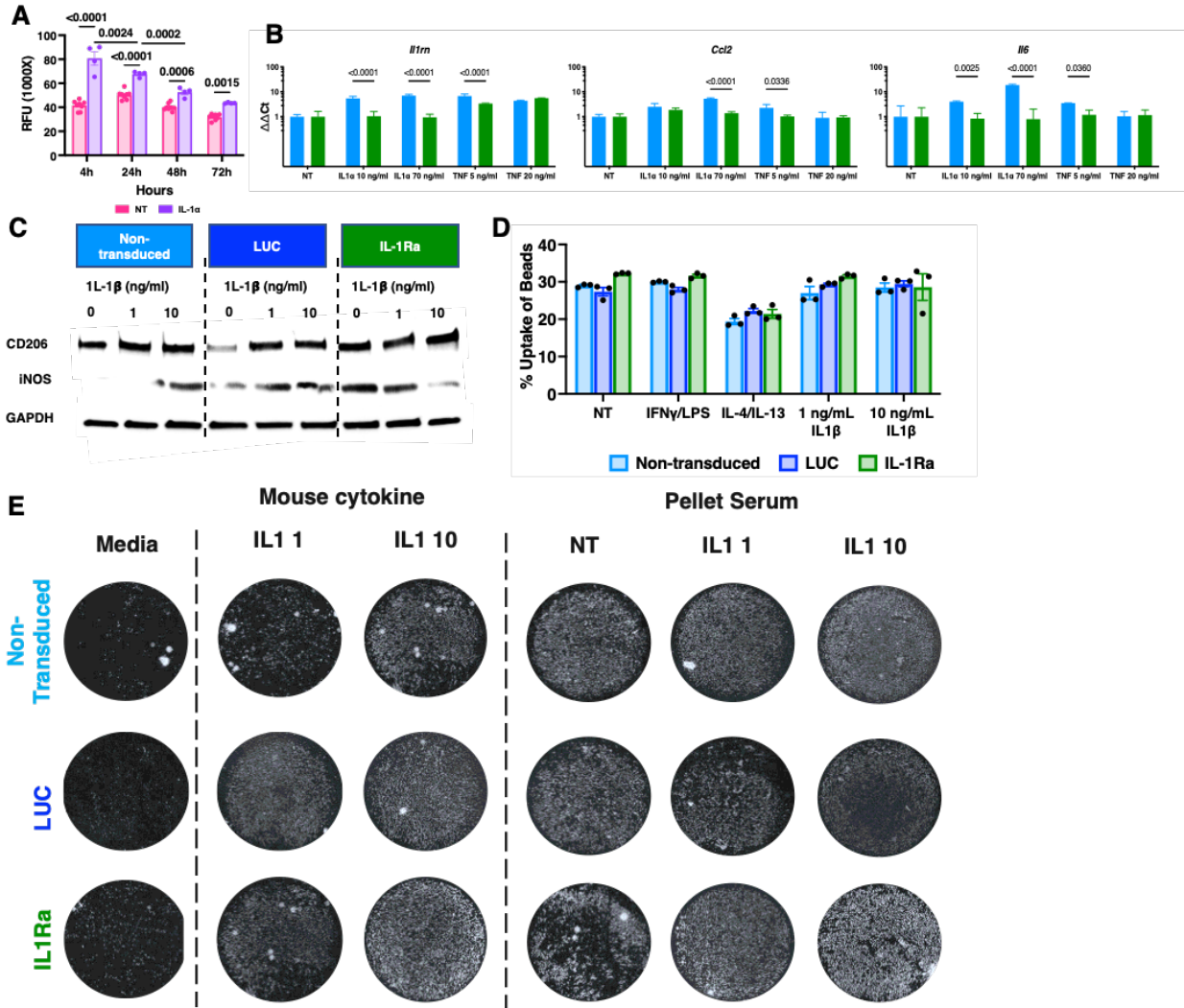
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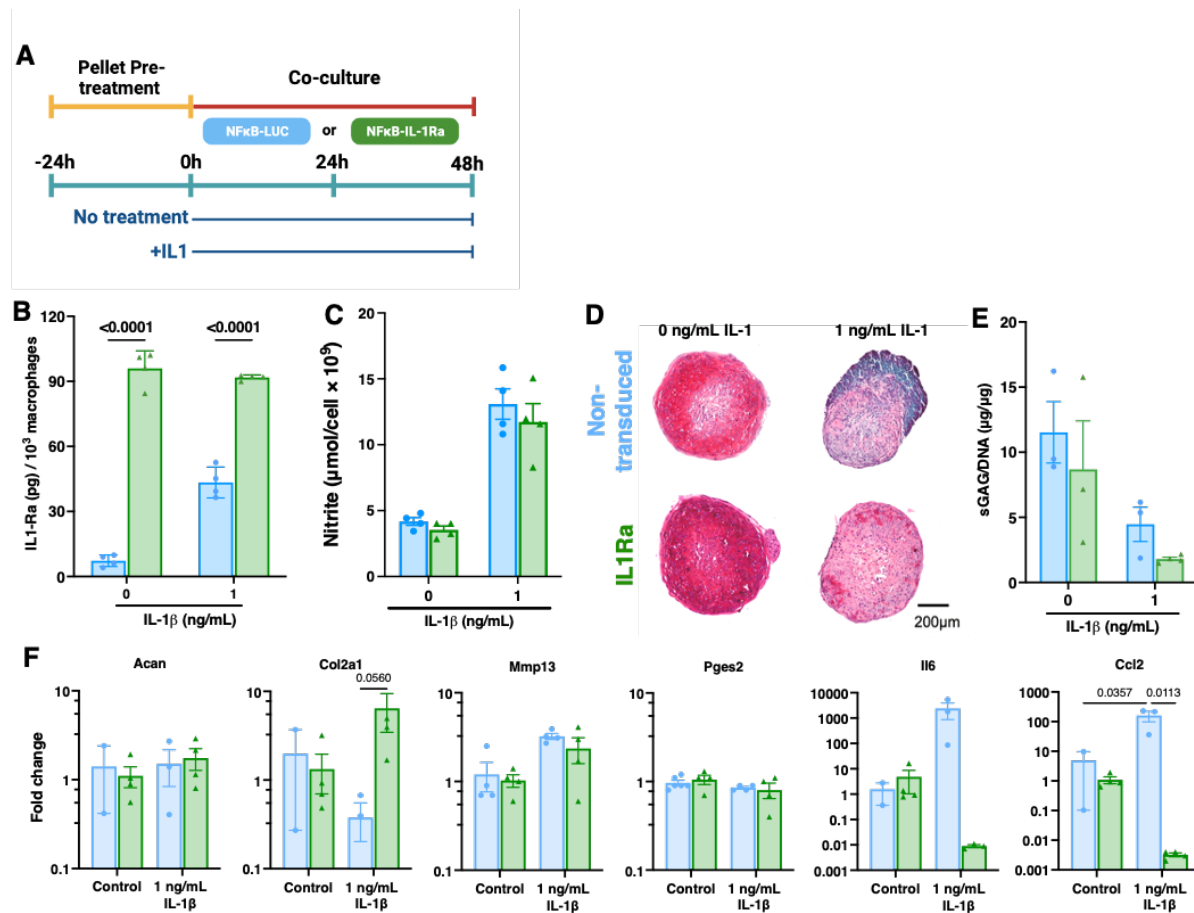
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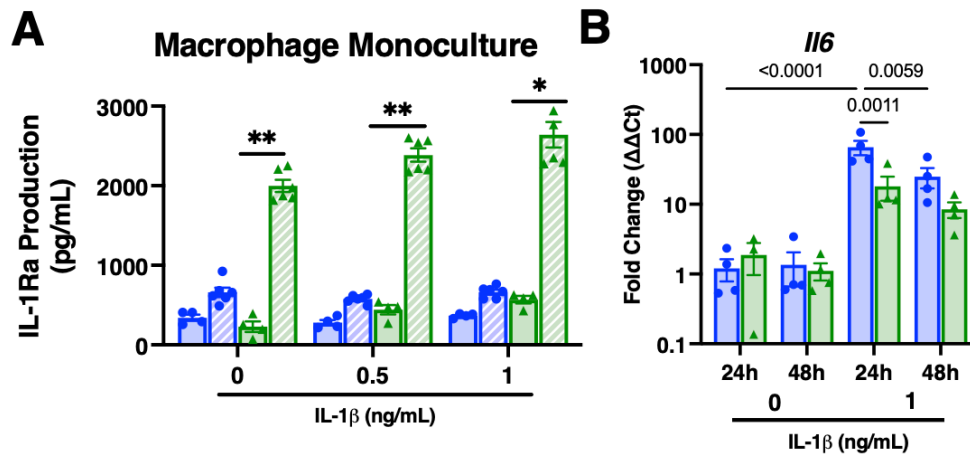
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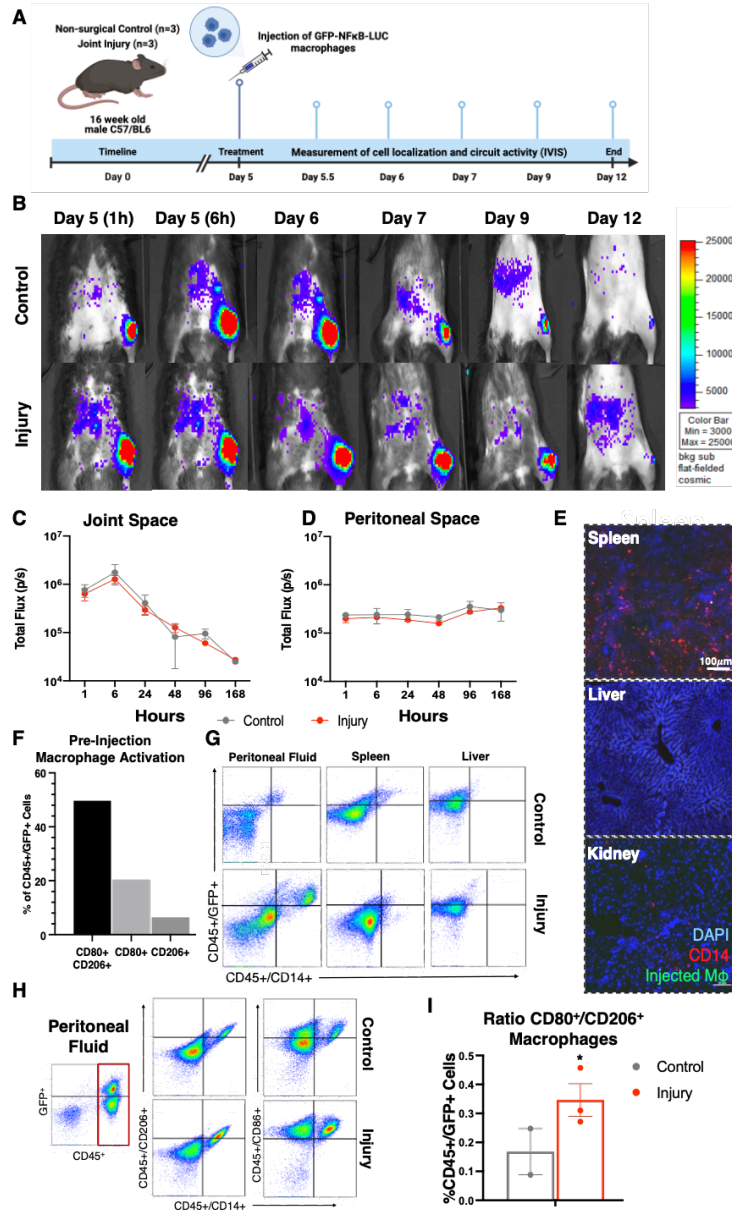
Supplemental Figure 1. Dynamic self-regulation of the NF-κB synthetic promoter in monolayer. (A) LUC macrophages demonstrate circuit activation by 4 hours which is resolved by 72 hours following treatment with IL-1 (n=5). (B) Gene expression of IL-1Ra BMDMs in monoculture in reference to non-transduced, non-treated BMDMs when challenged with either no treatment (NT), 10 or 70 ng/mL IL-1α, or 5 or 20 ng/mL of TNF for 48h normalized to GAPDH and R18S (n=4) demonstrated significant reduction in NF-κB responsive genes by IL-1Ra macrophages for both medium and high doses (n=4). (C) Western blot analysis revealed that NRE-IL-1Ra macrophages possessed a similar phenotype in comparison to non-transduced and NRE-LUC macrophages in response to IL-1β (n=3). Full replicates are shown in Supplemental Figure 5. (D) Flow cytometry of engineered macrophages as compared to non-transduced controls demonstrates little change in phagocytosis of FITC labeled latex beads following a 4-hour incubation (n=3). (E) Migration of BMDMs in response to both cytokine and conditioned media stimuli for non-transduced and engineered macrophages over 16 hours (n=3). Data was assessed using a one or two-way ANOVA with Tukey's post hoc test; values represent mean with standard error of the mean (SEM) (n = 3-4); NT, no treatment; RFU, relative luminescence unit; IL-1, Interleukin-1; TNF - tumor necrosis factor.



Supplemental Figure 2. IL-1Ra macrophages offer moderate chondroprotection against inflammatory mediators and inflammatory driven matrix degradation in response to IL-1β. (A) Timeline of co-culture experimental conditions. (B) LUC/IL-1Ra macrophage production of IL-1Ra in co-culture over 48 hours, as measured by ELISA, following stimulation with 0.5 or 1 ng/ml of IL-1β. (C) Concentration of nitric oxide (NO₂-) secreted into the media normalized to cell number following treatment with 0.5 or 1 ng/ml of IL-1β. (D) Safranin-O (red)/fast green(blue)/hematoxylin-stained tissue sections of miPSC derived cartilage co-cultured with LUC or IL-1Ra macrophages. (E) Amount of sGAG per pellet normalized to DNA content and secreted into the media during culture. (F) Gene expression of miPSC derived cartilage pellets in co-culture in reference to non-treated (NT) control when challenged with 1 ng/mL IL-1β for 48h normalized to GAPDH and R18S (n=3-4) demonstrated significant reduction in NF-κB responsive genes and some chondroprotective effects when cultured with IL-1Ra macrophages. Data was assessed using a two-way ANOVA with Tukey's post hoc test; * p < 0.05, ** p < 0.01, *** p < 0.001, **** p < 0.0001; values represent mean with standard error of the mean (SEM) (n = 3-4).



Supplemental Figure 3. IL-1Ra macrophages offer moderate chondroprotection against inflammatory mediators and inflammatory driven matrix degradation in response to IL-1 β . (A) LUC/IL-1Ra macrophage production of IL-1Ra in co-culture over 48 hours, as measured by ELISA, following stimulation with 0.5 or 1 ng/ml of IL-1 β . (B) Gene expression of engineered macrophages in monoculture in reference to non-treated control when challenged with 0 or 1 ng/mL IL-1 β for 48h normalized to GAPDH and R18S (n=3-4). Data was assessed using a two-way ANOVA with Tukey's post hoc test; * p < 0.05, ** p < 0.01, *** p < 0.001; values represent mean with standard error of the mean (SEM) (n = 3-4).



Supplemental Figure 4. Engineered macrophages can maintain their presence in the joint for at least 7 days. (A) Timeline of experimental conditions (B-D) NF-κB-LUC macrophages were given by interarticular injection into mice after receiving a joint injury or into non-surgical controls. Localization and activation of the engineered macrophages was monitored through IVIS imaging and quantified for the joint space and peritoneal fluid over time starting 1 hour following injection and continuing for 7 days (E) Macrophage activation was examined for inflammatory marker CD80 and anti-inflammatory marker CD206. At 7 days following injection, mouse tissues were examined for injected macrophages through (F) immunofluorescence as well as (G) flow cytometry on cells isolated from dissociated tissues (n=3). (H) Peritoneal fluid possessed the only notable population of engineered macrophages and was examined for macrophage activation post-injection based on the same markers, CD80 and CD206, which are (I) quantified.

Supplemental Table 1. qPCR primers

Gene	Forward	Reverse
<i>Ccl2</i>	GGC TCA GCC AGA TGC AGT TAA	CCT ACT CAT TGG GAT CAT CTT GCT
<i>IL1RN</i>	GTC CAG GAT GGT TCC TCT GC	TCT TCC GGT GTG TTG GTG AG
<i>IL6</i>	GAG GAT ACC ACT CCC AAC AGA CC	AAG TGC ATC ATC GTT GTT CAT ACA
<i>R18s</i>	CGG CTA CCA CAT CCA AGG AA	GGG CCT CGA AAG AGT CCT GT
<i>Acan</i>	GCA TGA GAG AGG CGA ATG GA	CTG ATC TCG TAG CGA TCT TTC TTC T
<i>Col2a1</i>	TCC AGA TGA CTT TCC TCC GTC TA	AGG TAG GCG ATG CTG TTC TTA CA
<i>GAPDH</i>	CAT GGC CTT CCG TGT TCC TA	TGT CAT CAT ACT TGG CAG GTT TCT
<i>TNFa</i>	GGT GCC TAT GTC TCA GCC TCT T	GCC ATA GAA CTG ATG AGA GGG AG
<i>PGES2</i>	AAG ACA TGT CCC TTC TGC	CCA AGA TGG GCA CTT TCC
<i>MMP13</i>	TCA AGG CTA TGC ACA CTG GT	CAC TAT GGT CTT TTC AAT GCC TAA
<i>ADAMPTS4</i>	CAG TCA CCT CTA AGC CAA AGAAA	CTT CCG GCG TAG GAT GTG AG

Supplemental Table 2. Flow cytometry antibodies

Conjugate	Specificity	Manufacturer	Catalog Number
Sytox Blue	Live/Dead	Thermo Scientific	S34857
PerCP-Cy5.5	anti-mouse F4/80	Biolegend	123127
PE	anti-mouse CD163	Biolegend	156703
PE	anti-mouse CD14	Biolegend	123309
PE	anti-mouse CD206	BioLegend	141705
PE	anti-mouse CD163	Biolegend	156703
PE	anti-mouse CD80	Biolegend	104707
PE	anti-mouse CD45	Biolegend	103105
PE	anti-mouse CD301	Biolegend	145703
PE/Cy7	anti-mouse CD206	BioLegend	141719
PE/Cy7	anti-mouse CD11b	Biolegend	101215
PE/Cy7	anti-mouse CD80	Biolegend	104733
PE/Cy7	anti-mouse Cx3CR1	Biolegend	149015
APC	anti-mouse CD206	BioLegend	141707
APC	anti-mouse CD11b	Biolegend	101211
APC	anti-mouse CD45	Biolegend	103111