

Supplementary information:
Semi-supervised prediction of protein fitness for data-driven protein
engineering

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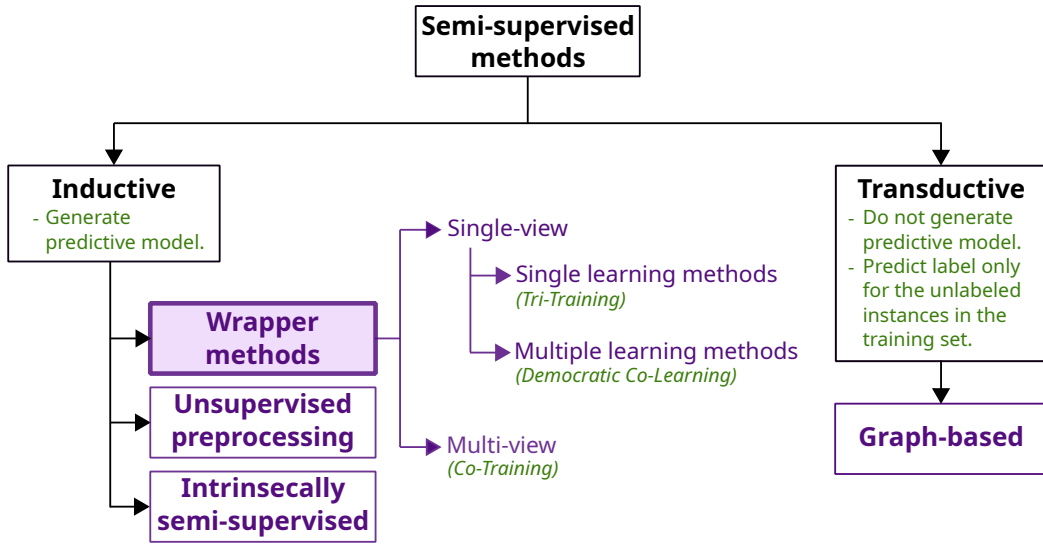


Figure S1: General taxonomy of semi-supervised learning classifiers described in [18].

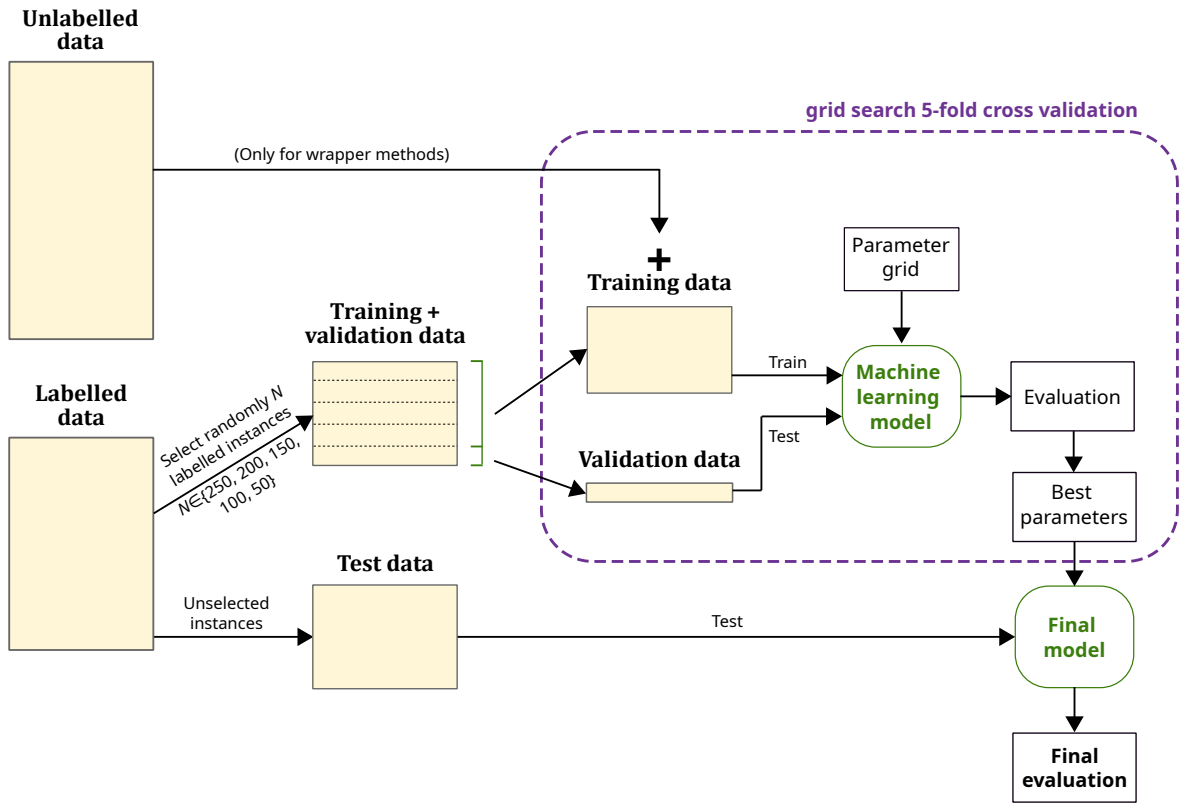


Figure S2: One iteration of the evaluation process when only few labelled instances available are used for training. The whole process, including the 5-fold cross validation for parameters tuning, is repeated 10 times for each of the possible values of N . Grid search via 5-fold cross validation is performed only for wrapper methods. The rest use default parameters for the model.

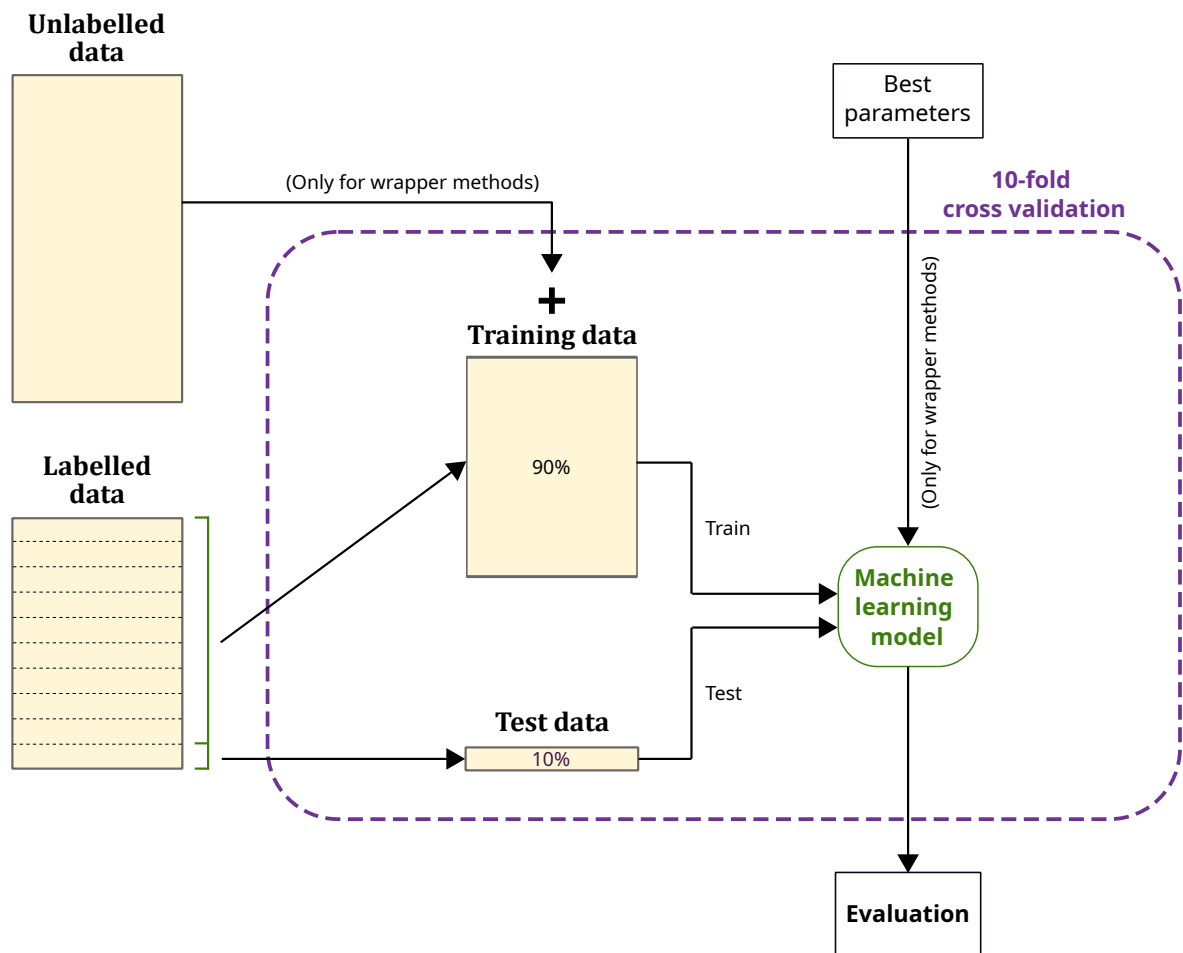


Figure S3: **One iteration of the evaluation process when all the labelled instances available are used for training.** 10-fold cross-validation is used. Best parameters are calculated for each dataset as the mode of the best parameters found using only few labelled instances.

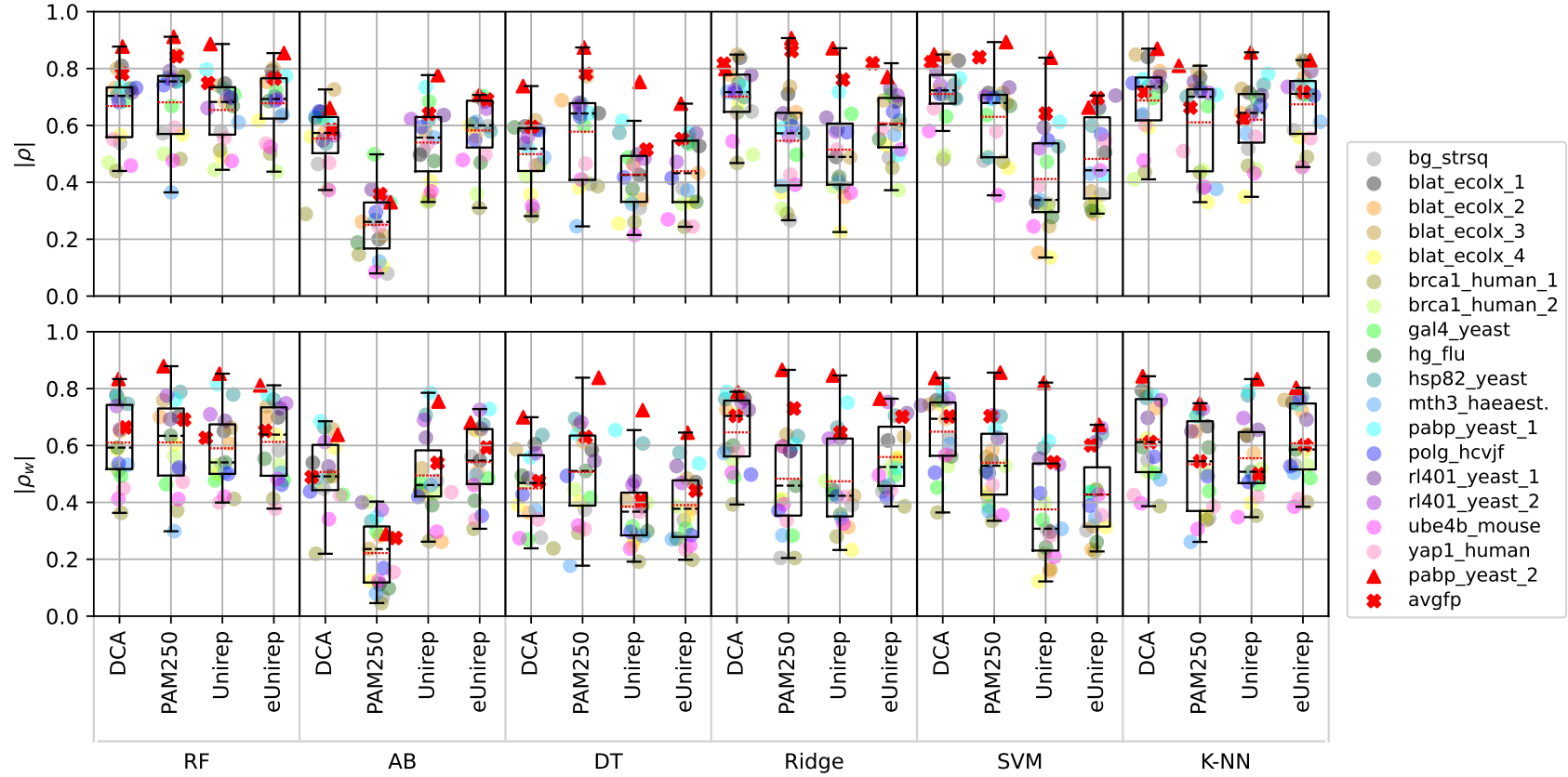


Figure S4: **Absolute Spearman's ρ and absolute Weighted Spearman's ρ_w of the methods in strategies 0 and 1, using all available labelled sequences on each dataset for training and validation.** Coloured dots represent the different datasets. Dataset abbreviations in the legend are defined in Table S1. Lines indicating median values in black and average values in red.

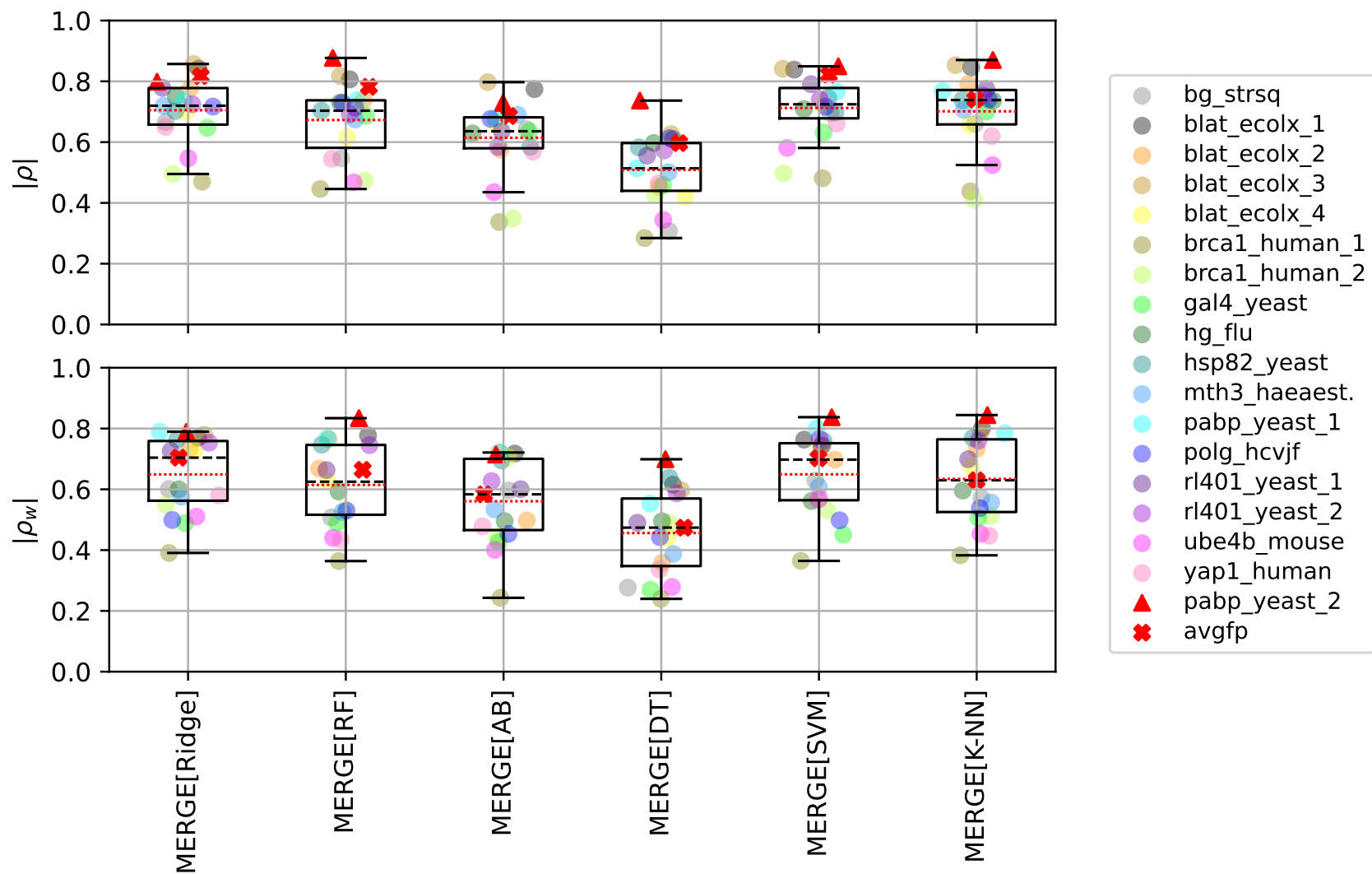


Figure S5: **Absolute Spearman's ρ and absolute Weighted Spearman's ρ_w of the methods in strategy 2, using all available labelled sequences on each dataset for training and validation.** Coloured dots represent the different datasets. Dataset abbreviations in the legend are defined in Table S1. Lines indicating median values in black and average values in red.

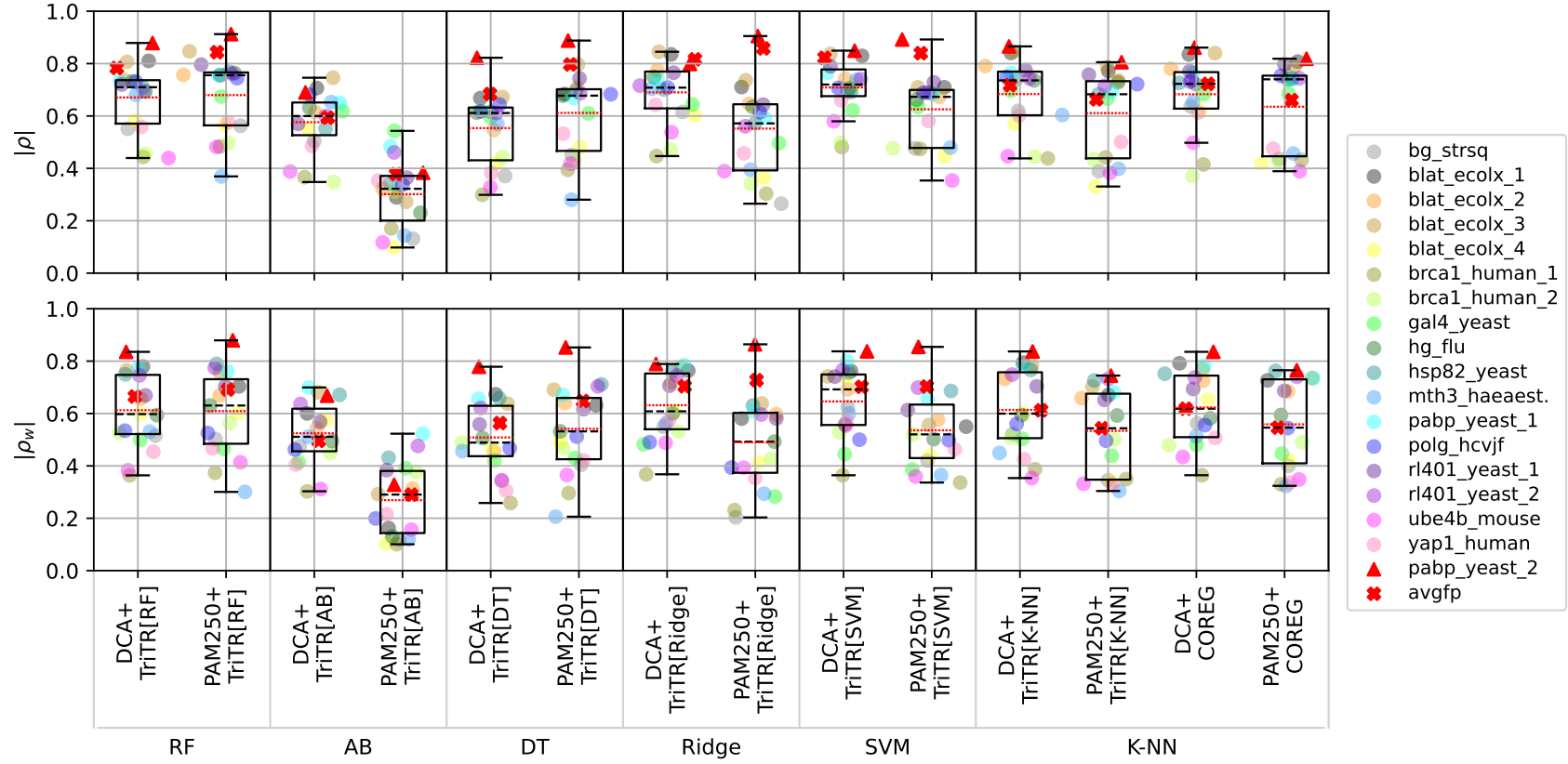


Figure S6: **Absolute Spearman's ρ and absolute Weighted Spearman's ρ_w of the methods in strategies 3 and 4, using all available labelled sequences on each dataset for training and validation.** Coloured dots represent the different datasets. Dataset abbreviations in the legend are defined in Table S1. Lines indicating median values in black and average values in red.

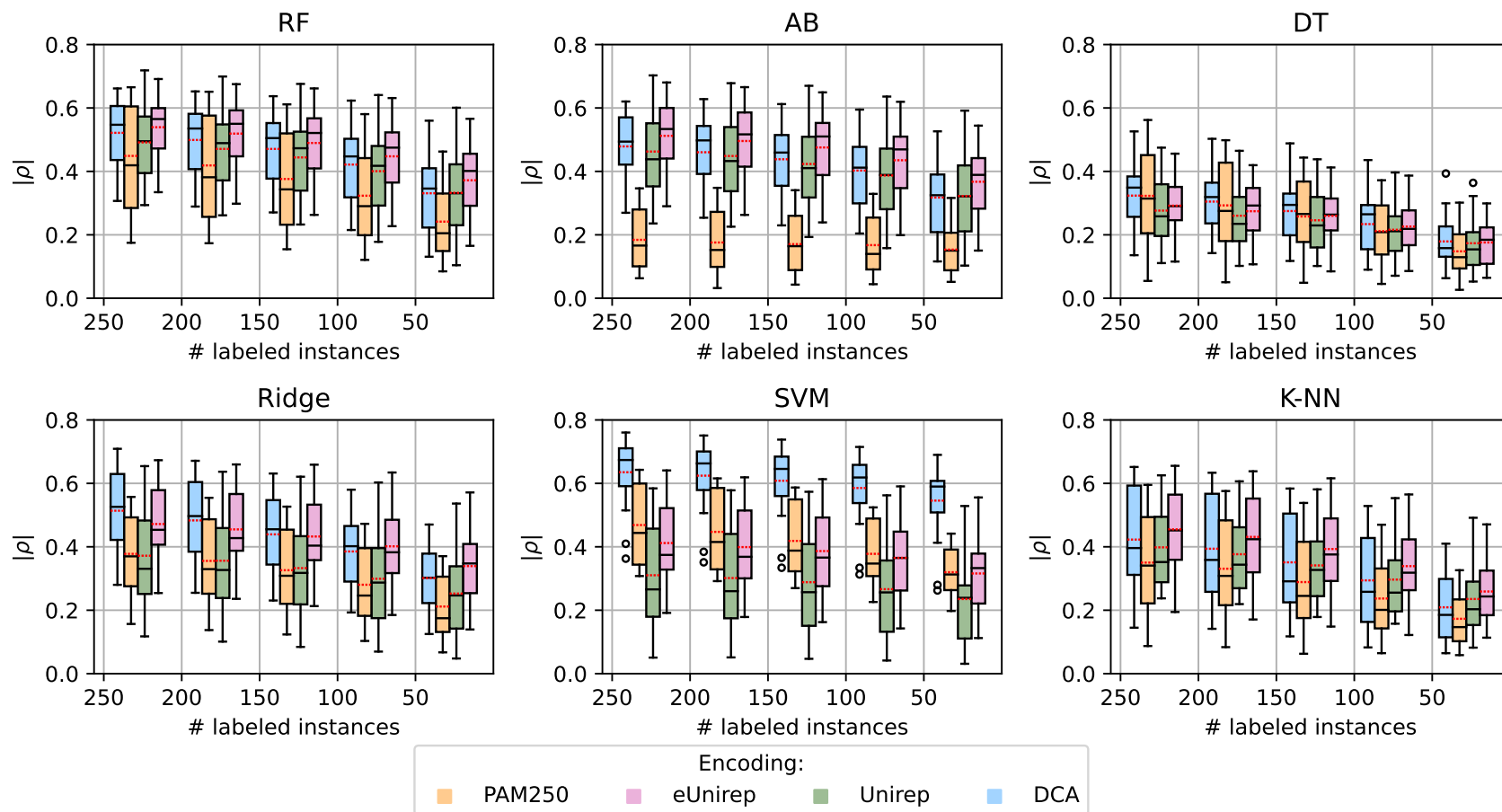


Figure S7: **Absolute Spearman's ρ of the methods in strategies 0 and 1, as a function of the number of labelled sequences used for training.** Coloured dots represent the different datasets. Dataset abbreviations in the legend are defined in Table S1. Lines indicating median values in black and average values in red.

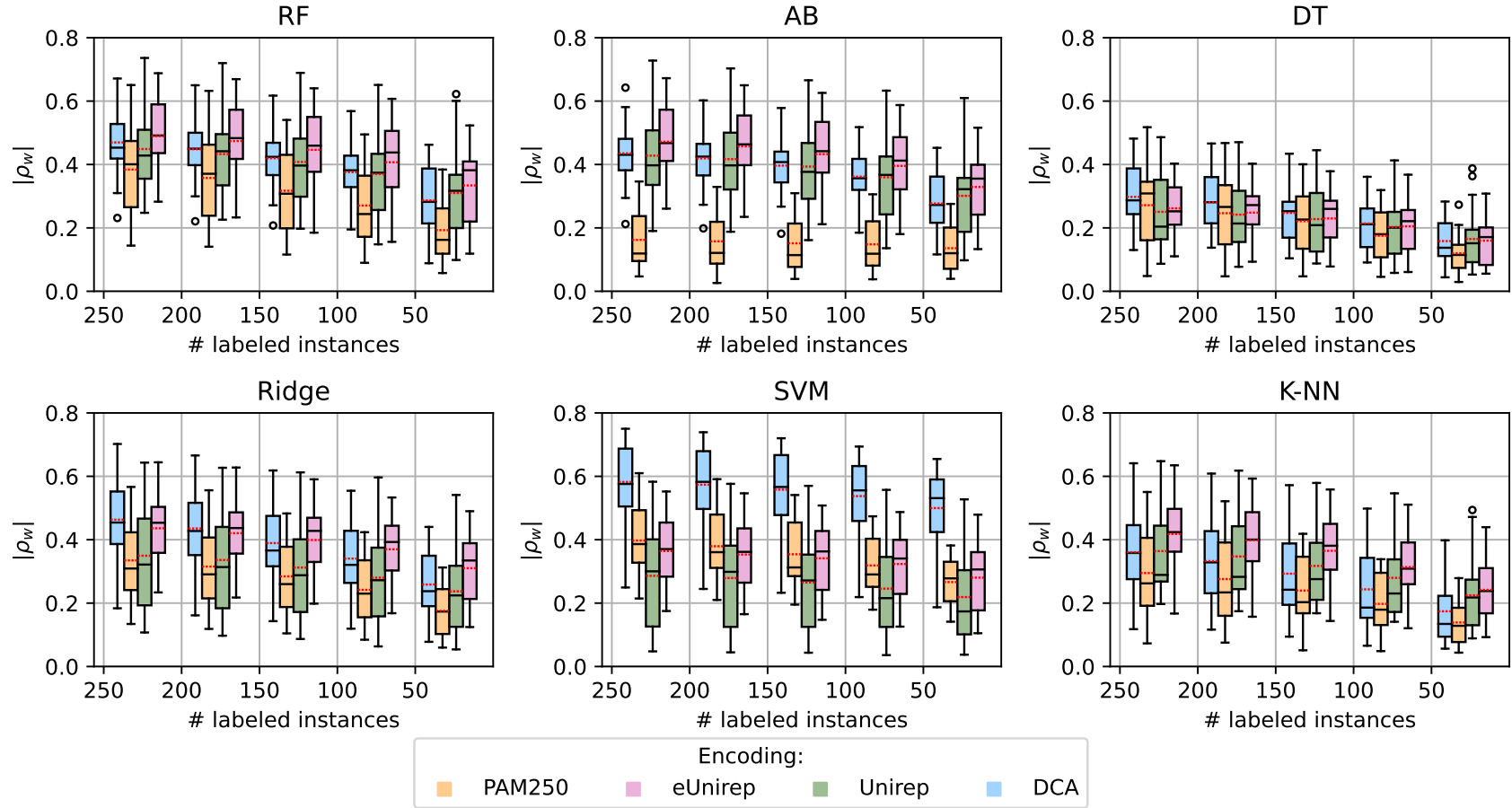


Figure S8: **Absolute Weighted Spearman's ρ_w of the methods in strategies 0 and 1, as a function of the number of labelled sequences used for training.** Coloured dots represent the different datasets. Dataset abbreviations in the legend are defined in Table S1. Lines indicating median values in black and average values in red.

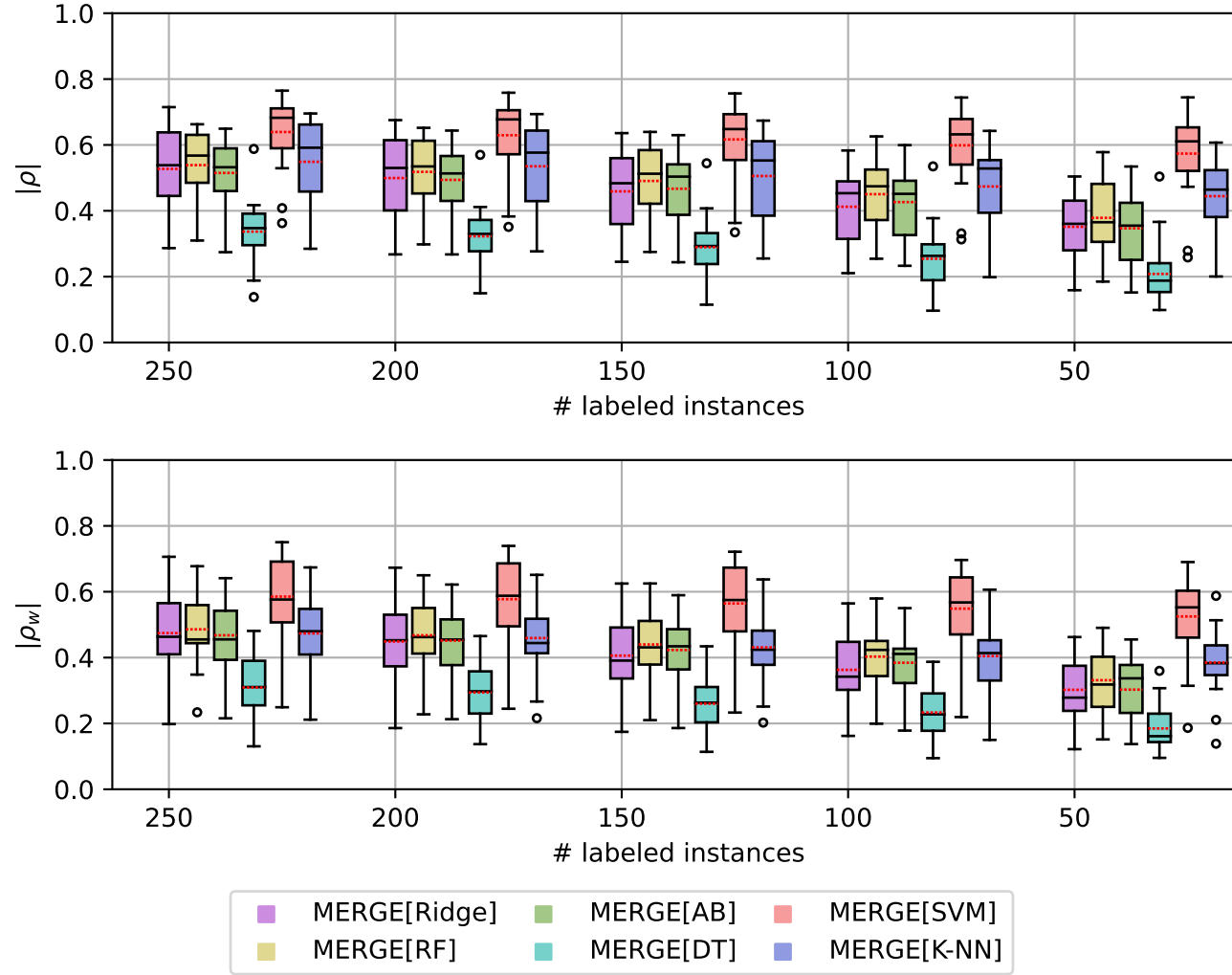


Figure S9: **Absolute Spearman's ρ and absolute Weighted Spearman's ρ_w of the methods in strategy 2, as a function of the number of labelled sequences used for training.** Coloured dots represent the different datasets. Dataset abbreviations in the legend are defined in Table S1. Lines indicating median values in black and average values in red.

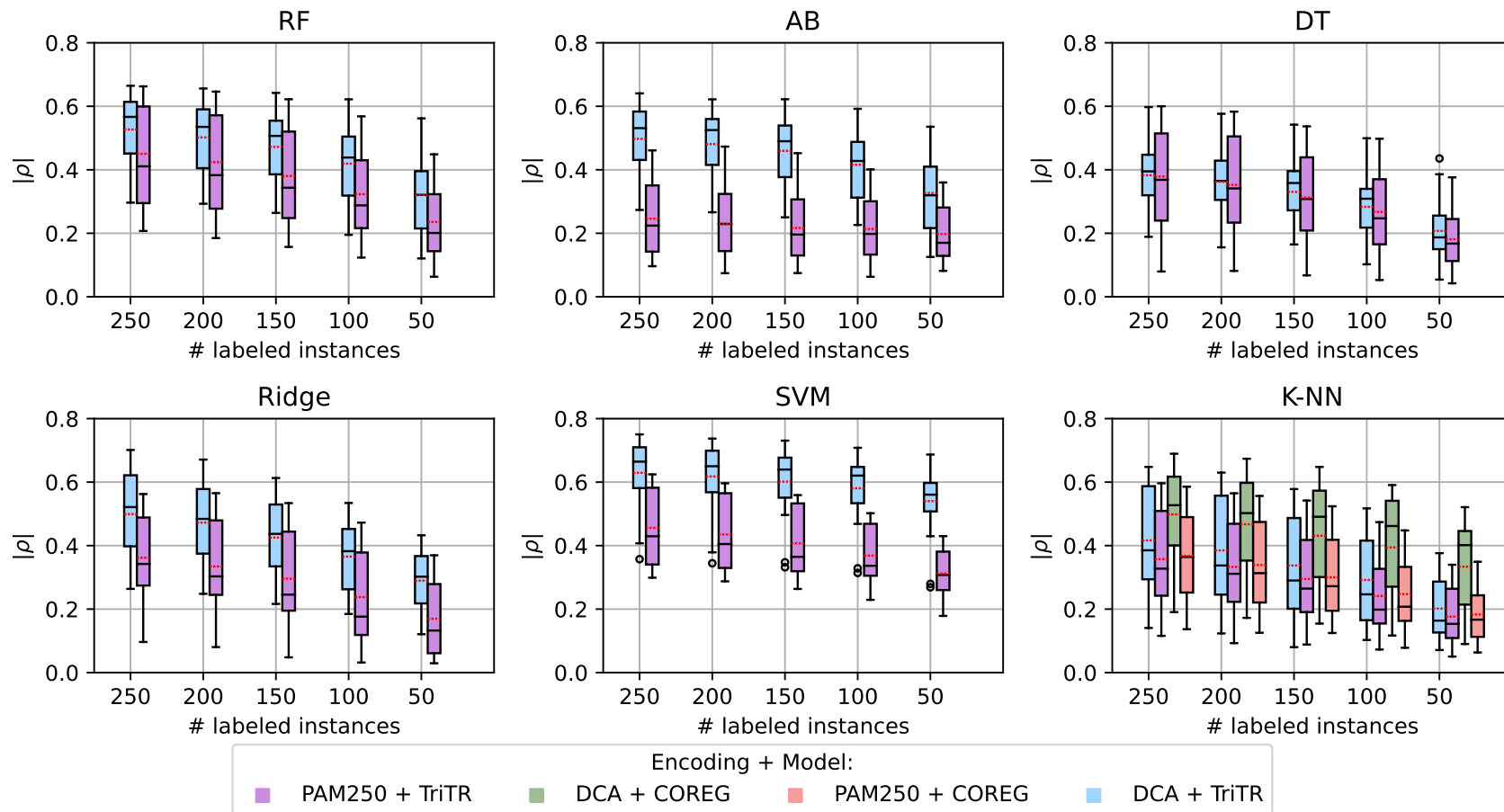


Figure S10: **Absolute Spearman's ρ of the methods in strategies 3 and 4, as a function of the number of labelled sequences used for training.** Coloured dots represent the different datasets. Dataset abbreviations in the legend are defined in Table S1. Lines indicating median values in black and average values in red.

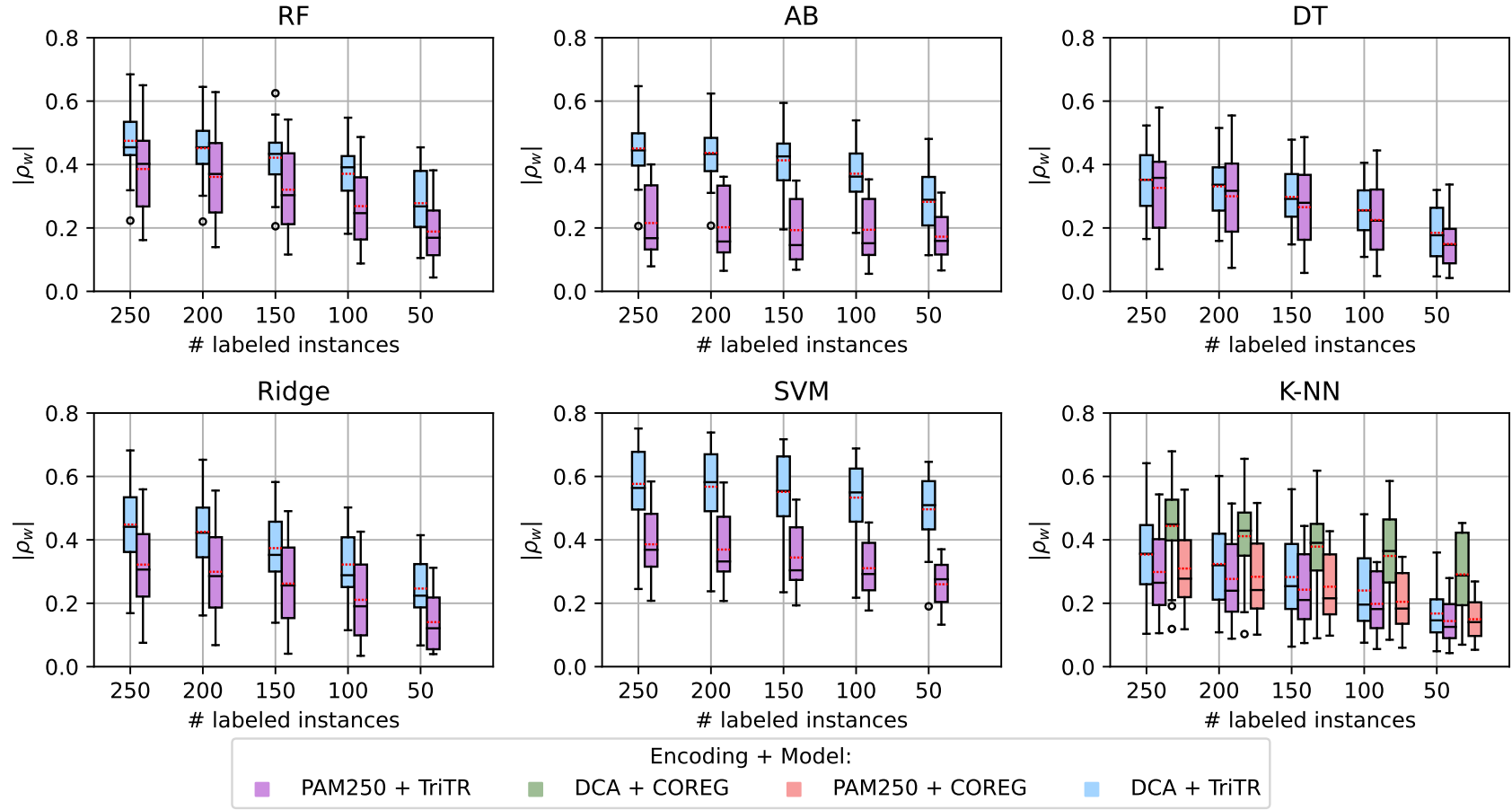


Figure S11: **Absolute Weighted Spearman's ρ_w of the methods in strategies 3 and 4, as a function of the number of labelled sequences used for training.** Coloured dots represent the different datasets. Dataset abbreviations in the legend are defined in Table S1. Lines indicating median values in black and average values in red.

2 Supplementary tables

Table S1: Datasets included in experimentation and their characteristics.

Dataset	Abbreviation	Length	L	U	%L	Subst.
YAP1_HUMAN_Fields2021 [1]	yap1_human	34	313	248 330	0.1	single
UBE4B_MOUSE_Klevit2013 [15]	ube4b_mouse	102	518	97 159	0.5	single
GAL4_YEAST_Shendure2015 [6]	gal4_yeast	64	803	171 882	0.5	single
BLAT_ECOLX_Tenaillon2013 [5]	blat_ecolx_4	286	975	30 356	3.1	single
PABP_YEAST_Fields2013 [7]	pabp_yeast_1	75	1 142	948 246	0.1	single
RL401_YEAST_Bolon2013 [13]	rl401_yeast_1	75	1 154	96 590	1.2	single
BRCA1_HUMAN_Fields2015_y2h [16]	brca1_human_2	303	1 278	1 909	40.1	single
RL401_YEAST_Bolon2014 [12]	rl401_yeast_2	75	1 282	96 590	1.3	single
MTH3_HAESTABILIZED_Tawfik2015 [10]	mth3_haeaest.	329	1 611	75 260	2.1	single
POLG_HCVJF_Sun2014 [9]	polg_hcvif	86	1 613	17 184	8.6	single
BG_STRSQ_Abate2015 [11]	bg_strsq	478	2 598	124 487	2.0	single
BRCA1_HUMAN_Fields2015_e3 [16]	brca1_human_1	303	2 846	1 909	59.9	single
HSP82_YEAST_Bolon2016 [8]	hsp82_yeast	230	4 065	59 490	6.4	single
BLAT_ECOLX_Ostermeier2014 [4]	blat_ecolx_1	286	4 799	30 356	13.7	single
BLAT_ECOLX_Ranganathan2015 [17]	blat_ecolx_3	286	4 921	30 356	13.9	single
BLAT_ECOLX_Palzkill2012 [2]	blat_ecolx_2	286	4 922	30 356	14.0	single
HG_FLU_Bloom2016 [3]	hg_flu	564	10 337	71 029	12.7	single
PABP_YEAST_Fields2013 [7]	pabp_yeast_2	75	33 771	948 246	3.4	double
avGFP_Kondrashov2016 [14]	avgfp	235	32 610	697	97.9	multiple

Sorted by number of labelled sequences. *Length* stands for the number of amino acids in the protein sequence, *L* stands for the number of labelled sequences, *U* stands for the number of homologous sequences found (unlabelled) and *%L* represents the percentage of labelled sequences out of the total number of sequences.

Table S2: **Datasets included in experimentation and the protein function for which the fitness is given.**

Dataset	Protein function for which the fitness is given
YAP1_HUMAN_Fields2021 [1]	Ability of a WW domain to bind to a peptide ligand.
UBE4B_MOUSE_Klevit2013 [15]	Interaction strength between murine ubiquitination factor E4B (Ube4b) U-box domain and auto-ubiquitination activity in the presence of the E2 UBcH5c.
GAL4_YEAST_Shendure2015 [6]	Interaction strength between the DNA-binding domain (DBD) and the yeast transcription factor Gal4.
BLAT_ECOLX_Tenaillon2013 [5]	Resistance to ampicillin of TEM-1 β -lactamase enzyme.
PABP_YEAST_Fields2013 [7]	Enrichment scores of <i>Saccharomyces cerevisiae</i> poly-A binding protein (PABP).
RL401_YEAST_Bolon2013 [13]	Effects of mutations to ubiquitin on yeast growth.
BRCA1_HUMAN_Fields2015_y2h [16]	Interaction strength between breast cancer 1 (BRCA1) protein and the BRCA1-associated RING domain protein 1.
RL401_YEAST_Bolon2014 [12]	Effects of mutations to ubiquitin on activating enzyme (E1) activity.
MTH3_HAESTABILIZED_Tawfik2015 [10]	Methylation activity of HaeIII methyltransferase.
POLG_HCVJF_Sun2014 [9]	Effects of mutations to domain IA of NS5A inhibitor on Hepatitis C virus (HCV) replication rates.
BG_STRSQ_Abate2015 [11]	Hydrolysis activity of a β -glucosidase enzyme.
BRCA1_HUMAN_Fields2015_e3 [16]	Interaction strength between breast cancer 1 (BRCA1) protein and the BRCA1-associated RING domain protein 1.
HSP82_YEAST_Bolon2016 [8]	The spread of mutant lineages of the heat shock protein 90 (Hsp90).
BLAT_ECOLX_Ostermeier2014 [4]	Resistance to ampicillin of TEM-1 β -lactamase enzyme.
BLAT_ECOLX_Ranganathan2015 [17]	Resistance to ampicillin of TEM-1 β -lactamase enzyme.
BLAT_ECOLX_Palzkil2012 [2]	Resistance to ampicillin of TEM-1 β -lactamase enzyme.
HG_FLU_Bloom2016 [3]	Effects of mutations to H1 influenza hemagglutinin (HA) on viral replication rates
PABP_YEAST_Fields2013 [7]	Enrichment scores of <i>Saccharomyces cerevisiae</i> poly-A binding protein (PABP).
avGFP_Kondrashov2016 [14]	Brightness values of <i>Aequorea Victoria</i> green fluorescent protein (avGFP).

Table S3: **Time and peak memory performance of the training process of the best methods found for each strategy. Calculated using 100, 1000 and 10000 labeled training sequences.**

Method	Time (s)			Peak memory (MB)		
	N=100	N=1000	N=10000	N=100	N=1000	N=10000
Unirep + RF	4.14	113.55	2300.79	0.87	7.37	72.85
DCA + SVM	0.0024	0.093	12.05	0.23	1.67	16.09
DCA + MERGE[SVM]	0.71	0.81	29.69	0.29	1.67	16.16
PAM250 + TriTR[RF]	10.52	385.06	20368.90	89.56	151.75	1282.42
DCA + TriTR[SVM]	0.030	3.31	235.62	1.94	11.51	102.03

3 Supplementary algorithms

Algorithm S1 Original Tri-Training learning function.

```

1: function TRI-TRAINING( $L, U, \text{base\_classifier}$ )
2:   for  $i \in \{1...3\}$  do
3:      $S_i \leftarrow \text{BootstrapSample}(L)$ 
4:      $h_i \leftarrow \text{base\_classifier}(S_i)$ 
5:      $e'_i \leftarrow 0.5$ 
6:      $l'_i \leftarrow 0$ 
7:   repeat
8:     for  $i \in \{1...3\}$  do
9:        $L_i \leftarrow \emptyset$ 
10:       $\text{update}_i \leftarrow \text{FALSE}$ 
11:       $e_i \leftarrow \frac{|\{\mathbf{x} \in L : h_j(\mathbf{x}) = h_k(\mathbf{x}) \neq y\}|}{|\{\mathbf{x} \in L : h_j(\mathbf{x}) = h_k(\mathbf{x})\}|}$ 
12:      if  $e_i < e'_i$  then
13:        for each  $\mathbf{x} \in U$  do
14:          if  $h_j(\mathbf{x}) = h_k(\mathbf{x}) \quad (j, k \neq i)$  then
15:             $L_i \leftarrow L_i \cup \{(\mathbf{x}, h_j(\mathbf{x}))\}$ 
16:          if  $l'_i = 0$  then
17:             $l'_i \leftarrow \left\lfloor \frac{e_i}{e'_i - e_i} + 1 \right\rfloor$ 
18:          if  $l'_i < |L_i|$  then
19:            if  $e_i |L_i| < e'_i l'_i$  then
20:               $\text{update}_i \leftarrow \text{TRUE}$ 
21:            else if  $l'_i > \frac{e_i}{e'_i - e_i}$  then
22:               $L_i \leftarrow \text{Subsample}\left(L_i, \left\lceil \frac{e'_i l'_i}{e_i} - 1 \right\rceil\right)$ 
23:               $\text{update}_i \leftarrow \text{TRUE}$ 
24:      for  $i \in \{1...3\}$  do
25:        if  $\text{update}_i = \text{TRUE}$  then
26:           $h_i \leftarrow \text{base\_classifier}(L \cup L_i)$ 
27:           $e'_i \leftarrow e_i$ 
28:           $l'_i \leftarrow |L_i|$ 
29:  until none of  $h_i$  ( $i \in \{1...3\}$ ) changes
30:  return  $h(\mathbf{x}) \leftarrow \arg \max_{y \in \text{label}} \sum_{i: h_i(\mathbf{x}) = y}$ 

```

▷ Number of labelled examples on which both h_j and h_k make incorrect classification, by the number of labelled examples on which the classification made by h_j is the same as that made by h_k .

Algorithm S2 Tri-Training Regressor learning function.

Differences from Algorithm S1 are highlighted in red.

```
1: function TRI-TRAINING( $L, U, \text{base\_regressor}, y\_tol\_per$ )
2:   for  $i \in \{1...3\}$  do
3:      $S_i \leftarrow \text{BootstrapSample}(L)$ 
4:      $h_i \leftarrow \text{base\_regressor}(S_i)$ 
5:      $e'_i \leftarrow 0.5$ 
6:      $l'_i \leftarrow 0$ 
7:      $y\_tol \leftarrow \text{Tolerance}(L, y\_tol\_per)$   $\triangleright$  Calculates the tolerance as a percentage ( $y\_tol\_per$ ) of the
                                                difference between the largest output value and the smallest
                                                output value.

8:   repeat
9:     for  $i \in \{1...3\}$  do
10:       $L_i \leftarrow \emptyset$ 
11:       $update_i \leftarrow \text{FALSE}$ 
12:       $e_i \leftarrow \frac{|\{\mathbf{x} \in L : |h_j(\mathbf{x}) - h_k(\mathbf{x})| < y\_tol \wedge |\text{mean}(h_j(\mathbf{x}), h_k(\mathbf{x})) - y| \geq y\_tol\}|}{|\{\mathbf{x} \in L : |h_j(\mathbf{x}) - h_k(\mathbf{x})| < y\_tol\}|}$   $\triangleright$  Number of labelled
                                                                    examples for which
                                                                    both  $h_j$  and  $h_k$  pre-
                                                                    dict similar values
                                                                    but different to the
                                                                    actual  $y$ , divided by
                                                                    the number of la-
                                                                    belled examples for
                                                                    which both  $h_j$  and
                                                                     $h_k$  predict similar
                                                                    values.

13:      if  $e_i < e'_i$  then
14:        for each  $\mathbf{x} \in U$  do
15:          if  $|h_j(\mathbf{x}) - h_k(\mathbf{x})| < y\_tol \quad (j, k \neq i)$  then
16:             $L_i \leftarrow L_i \cup \{(\mathbf{x}, \text{mean}(h_j(\mathbf{x}), h_k(\mathbf{x})))\}$ 
17:          if  $l'_i = 0$  then
18:             $l'_i \leftarrow \left\lfloor \frac{e_i}{e'_i - e_i} + 1 \right\rfloor$ 
19:          if  $l'_i < |L_i|$  then
20:            if  $e_i |L_i| < e'_i l'_i$  then
21:               $update_i \leftarrow \text{TRUE}$ 
22:            else if  $l'_i > \frac{e_i}{e'_i - e_i}$  then
23:               $L_i \leftarrow \text{Subsample}\left(L_i, \left\lceil \frac{e'_i l'_i}{e_i} - 1 \right\rceil\right)$ 
24:               $update_i \leftarrow \text{TRUE}$ 
25:      for  $i \in \{1...3\}$  do
26:        if  $update_i = \text{TRUE}$  then
27:           $h_i \leftarrow \text{base\_regressor}(L \cup L_i)$ 
28:           $e'_i \leftarrow e_i$ 
29:           $l'_i \leftarrow |L_i|$ 
30:  until none of  $h_i$  ( $i \in \{1...3\}$ ) changes
31:  return  $h(\mathbf{x}) \leftarrow \text{mean}(h_1(\mathbf{x}), h_2(\mathbf{x}), h_3(\mathbf{x}))$ 
```

References

- [1] Carlos L Araya, Douglas M Fowler, Wentao Chen, Ike Muniez, Jeffery W Kelly, and Stanley Fields. A fundamental protein property, thermodynamic stability, revealed solely from large-scale measurements of protein function. *Proceedings of the National Academy of Sciences*, 109(42):16858–16863, 2012.
- [2] Zhifeng Deng, Wanzhi Huang, Erol Bakalbasi, Nicholas G Brown, Carolyn J Adamski, Kacie Rice, Donna Muzny, Richard A Gibbs, and Timothy Palzkill. Deep sequencing of systematic combinatorial libraries reveals β -lactamase sequence constraints at high resolution. *Journal of molecular biology*, 424(3-4):150–167, 2012.
- [3] Michael B Doud and Jesse D Bloom. Accurate measurement of the effects of all amino-acid mutations on influenza hemagglutinin. *Viruses*, 8(6):155, 2016.
- [4] Elad Firnberg, Jason W Labonte, Jeffrey J Gray, and Marc Ostermeier. A comprehensive, high-resolution map of a gene’s fitness landscape. *Molecular biology and evolution*, 31(6):1581–1592, 2014.
- [5] Hervé Jacquier, André Birgy, Hervé Le Nagard, Yves Mechulam, Emmanuelle Schmitt, Jérémy Glodt, Beatrice Bercot, Emmanuelle Petit, Julie Poulain, Guilène Barnaud, et al. Capturing the mutational landscape of the beta-lactamase tem-1. *Proceedings of the National Academy of Sciences*, 110(32):13067–13072, 2013.
- [6] Jacob O Kitzman, Lea M Starita, Russell S Lo, Stanley Fields, and Jay Shendure. Massively parallel single-amino-acid mutagenesis. *Nature methods*, 12(3):203–206, 2015.
- [7] Daniel Melamed, David L Young, Caitlin E Gamble, Christina R Miller, and Stanley Fields. Deep mutational scanning of an rrm domain of the *saccharomyces cerevisiae* poly (a)-binding protein. *Rna*, 19(11):1537–1551, 2013.
- [8] Parul Mishra, Julia M Flynn, Tyler N Starr, and Daniel NA Bolon. Systematic mutant analyses elucidate general and client-specific aspects of hsp90 function. *Cell reports*, 15(3):588–598, 2016.
- [9] Hangfei Qi, C Anders Olson, Nicholas C Wu, Ruian Ke, Claude Loverdo, Virginia Chu, Shawna Truong, Roland Remenyi, Zugen Chen, Yushen Du, Sheng-Yao Su, Laith Q Al-Mawsawi, Ting-Ting Wu, Shu-Hua Chen, Chung-Yen Lin, Weidong Zhong, James O Lloyd-Smith, and Ren Sun. A quantitative high-resolution genetic profile rapidly identifies sequence determinants of hepatitis c viral fitness and drug sensitivity. *PLoS pathogens*, 10(4):e1004064, 2014.
- [10] Liat Rockah-Shmuel, Ágnes Tóth-Petróczy, and Dan S Tawfik. Systematic mapping of protein mutational space by prolonged drift reveals the deleterious effects of seemingly neutral mutations. *PLoS computational biology*, 11(8):e1004421, 2015.
- [11] Philip A Romero, Tuan M Tran, and Adam R Abate. Dissecting enzyme function with microfluidic-based deep mutational scanning. *Proceedings of the National Academy of Sciences*, 112(23):7159–7164, 2015.
- [12] Benjamin P Roscoe and Daniel NA Bolon. Systematic exploration of ubiquitin sequence, e1 activation efficiency, and experimental fitness in yeast. *Journal of molecular biology*, 426(15):2854–2870, 2014.
- [13] Benjamin P Roscoe, Kelly M Thayer, Konstantin B Zeldovich, David Fushman, and Daniel NA Bolon. Analyses of the effects of all ubiquitin point mutants on yeast growth rate. *Journal of molecular biology*, 425(8):1363–1377, 2013.
- [14] Karen S Sarkisyan, Dmitry A Bolotin, Margarita V Meer, Dinara R Usmanova, Alexander S Mishin, George V Sharonov, Dmitry N Ivankov, Nina G Bozhanova, Mikhail S Baranov, Onuralp Soylemez, et al. Local fitness landscape of the green fluorescent protein. *Nature*, 533(7603):397–401, 2016.
- [15] Lea M Starita, Jonathan N Pruneda, Russell S Lo, Douglas M Fowler, Helen J Kim, Joseph B Hiatt, Jay Shendure, Peter S Brzovic, Stanley Fields, and Rachel E Klevit. Activity-enhancing mutations in an e3 ubiquitin ligase identified by high-throughput mutagenesis. *Proceedings of the National Academy of Sciences*, 110(14):E1263–E1272, 2013.
- [16] Lea M Starita, David L Young, Muhtadi Islam, Jacob O Kitzman, Justin Gullingsrud, Ronald J Hause, Douglas M Fowler, Jeffrey D Parvin, Jay Shendure, and Stanley Fields. Massively parallel functional analysis of brca1 ring domain variants. *Genetics*, 200(2):413–422, 2015.

- [17] Michael A Stiffler, Doeke R Hekstra, and Rama Ranganathan. Evolvability as a function of purifying selection in tem-1 β -lactamase. *Cell*, 160(5):882–892, 2015.
- [18] Jesper E Van Engelen and Holger H Hoos. A survey on semi-supervised learning. *Machine learning*, 109(2):373–440, 2020.