

SUPPLEMENTAL FIGURE LEGENDS

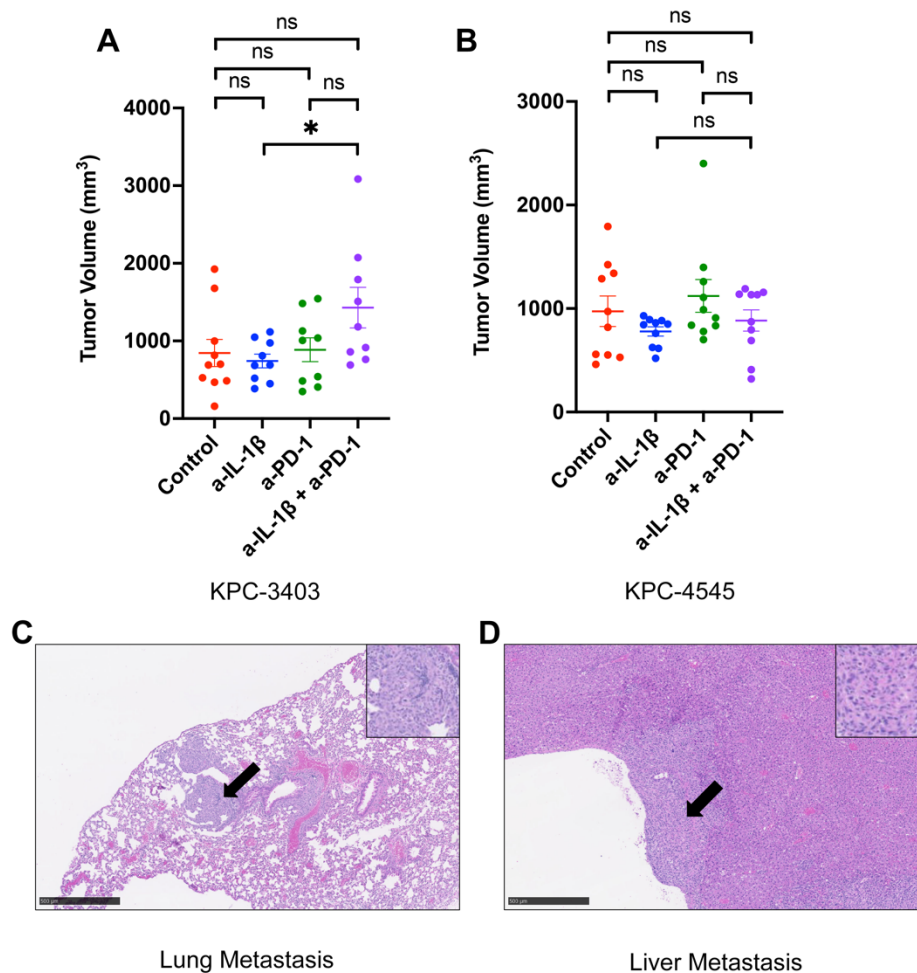


Figure S1. Tumor volume at necropsy and representative H&E images of liver and lung metastatic sites in PDAC orthotopic mouse model syngeneic wild-type C57Bl/6 mice.

(A-B) Tumor volume at necropsy across different treatments of anti-PD-1 and anti-IL-1 β as measured by a caliper in the KPC-3403 (A) and KPC-4545 model (B). **(C-D)** 5X magnification of a lung metastasis site (C) and a liver metastasis site (D) and metastatic regions (arrow) are magnified and displayed in top right corner, respectively.

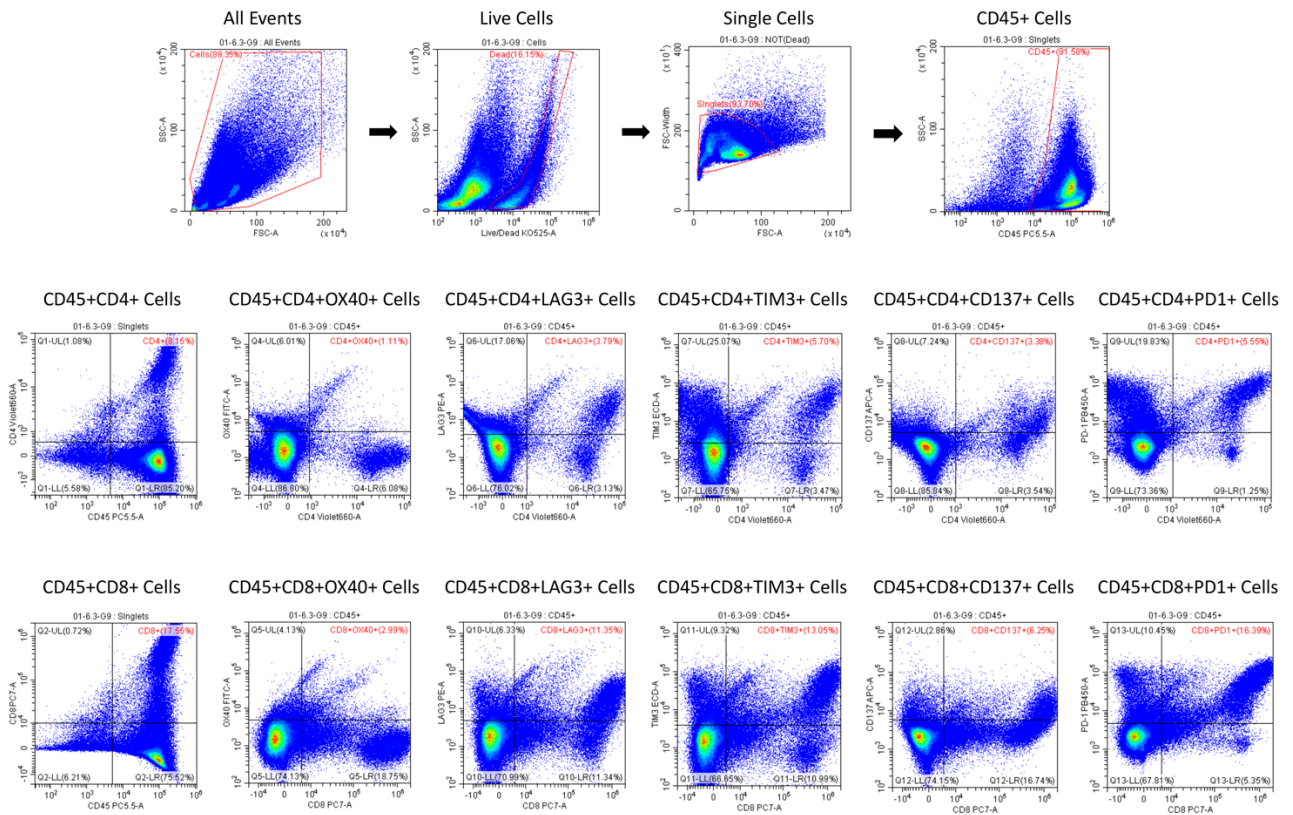


Figure S2. Flow cytometry gating strategy for identification of effector T cells.

Dead cells were excluded by gating on cells negative for viability marker Aqua Blue. Forward scatter width (FSC-W) and scatter area (FSC-A) were used to exclude doublets. CD45+ T-cells were identified. The flow cytometry gating strategy of different effector T-cell types are shown.

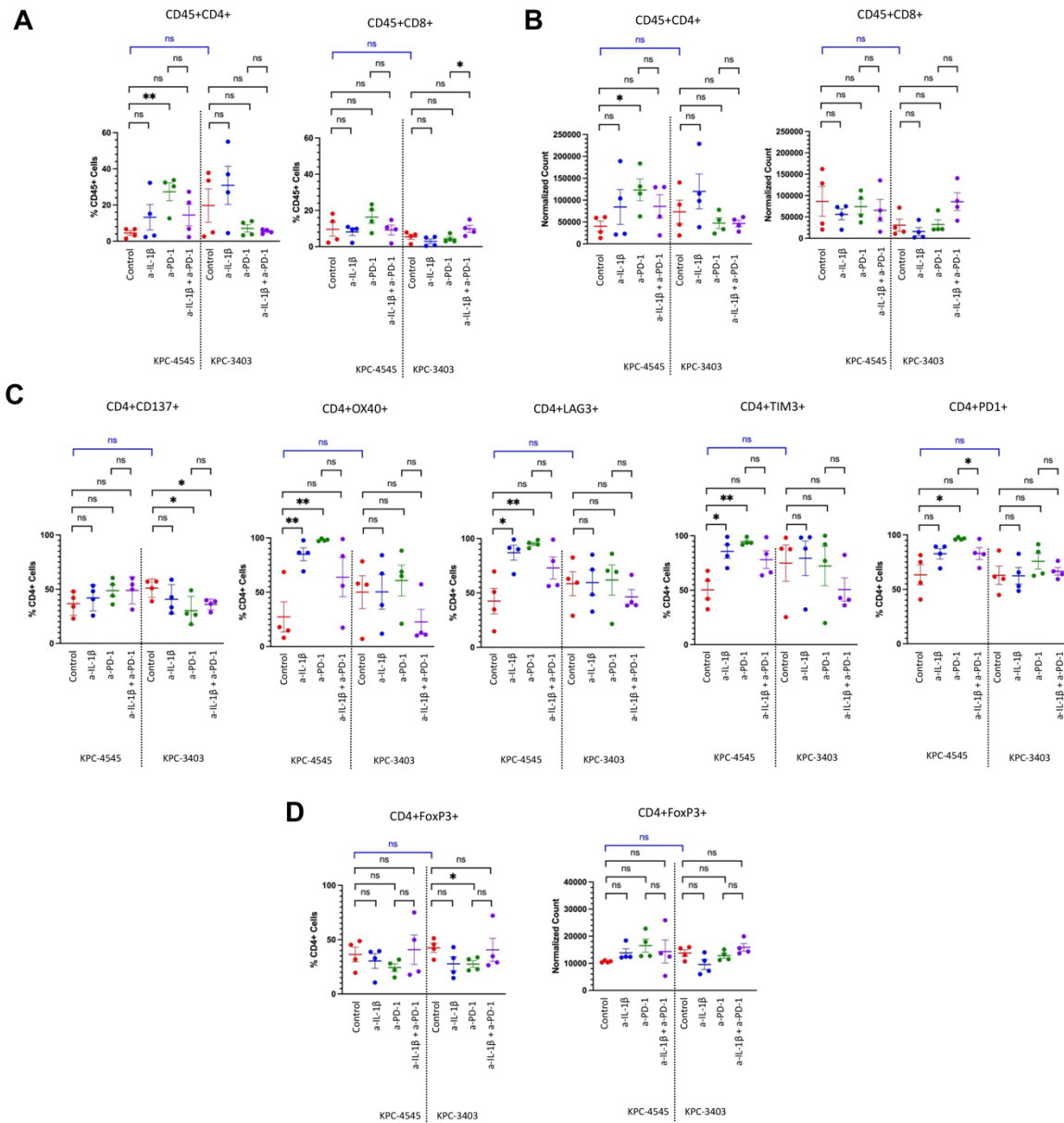


Figure S3. The combination of anti-PD-1 and anti-IL-1 β treatment had comparable levels of CD45+CD8+ T cells, CD45+CD4+ T cells, and regulatory T cells but modulates the activation and exhaustion of tumor infiltrating CD4+ T cells in the KPC-3403 model.

(A-D) Flow cytometry was performed on isolated tumor infiltrating leukocytes from resected orthotopic tumor on day 13. The following isolated tumor-infiltrating leukocytes were analyzed

23 (n=4 mice per group): **(A)** percentage of CD4⁺ and CD8⁺ cells among CD45⁺ cells, **(B)**
24 normalized cell counts of CD4⁺ and CD8⁺ cells per 1 x 10⁶ cells, **(C)** percentage of CD137⁺,
25 OX40⁺, LAG3⁺, TIM3⁺, and PD1⁺ cells among CD45⁺CD4⁺ cells, **(D)** percentage of FoxP3⁺
26 cells among CD4⁺ cells (n = 4 mice per group), normalized cell counts of CD4⁺FoxP3⁺ per 1 x
27 10⁶ cells (n = 4 mice per group). *, p<0.05; **, p<0.01; ***, p<0.001, by unpaired t-test. Data
28 represent mean ± SEM.

29

30

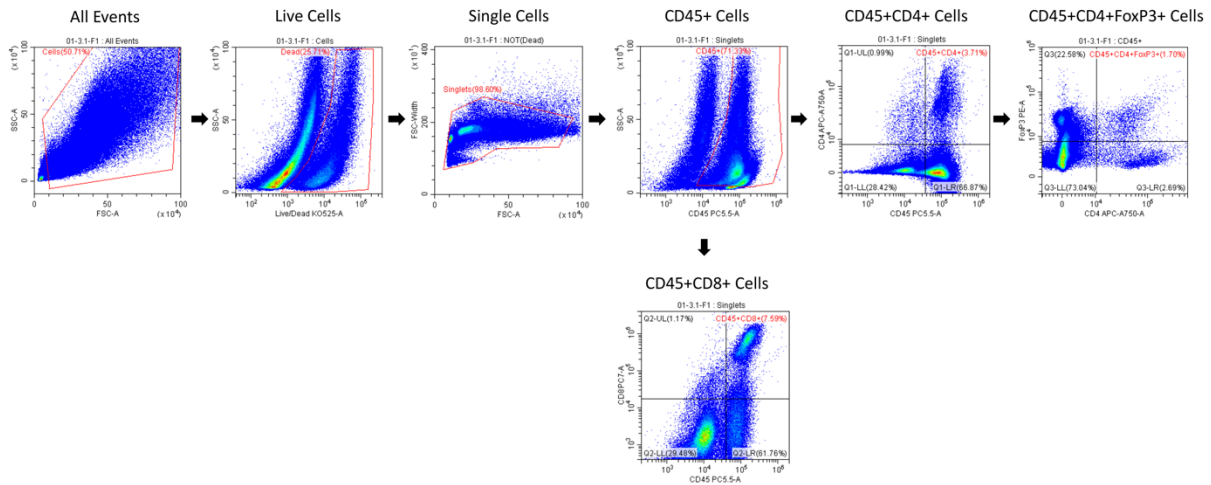


Figure S4. Flow cytometry gating strategy for identification of regulatory T-cells.

Dead cells were excluded by gating on cells negative for viability marker Aqua Blue. Forward scatter width (FSC-W) and scatter area (FSC-A) were used to exclude doublets. CD45+ cells were identified. Regulatory T-cells were defined as CD45+CD4+FoxP3+ cells.

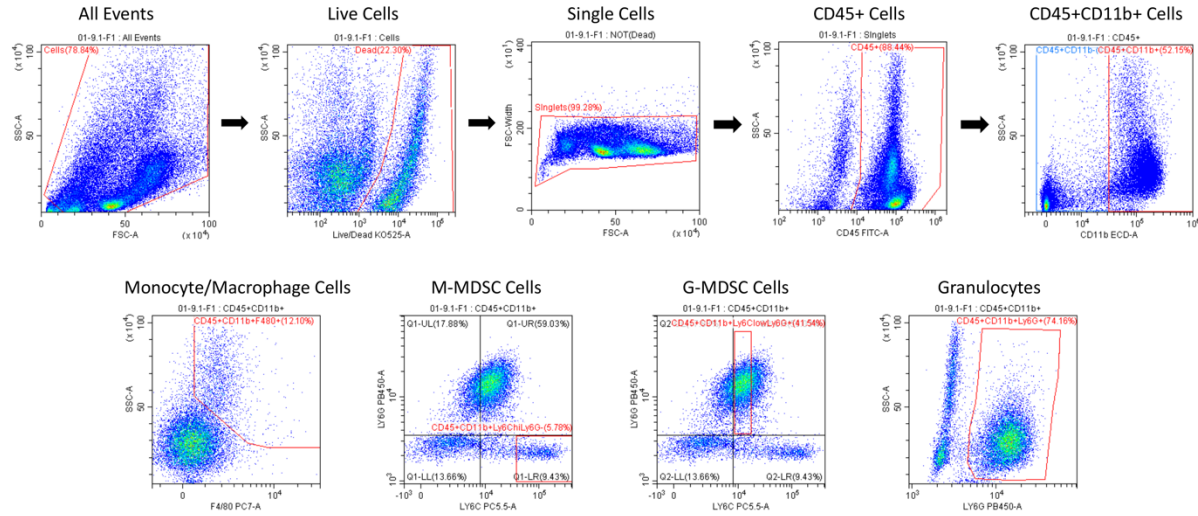


Figure S5. Flow cytometry gating strategy for identification of myeloid cell subtypes.

Dead cells were excluded by gating on cells negative for viability marker Aqua Blue. Forward scatter width (FSC-W) and scatter area (FSC-A) were used to exclude doublets. CD45+CD11b+ cells were identified. Monocyte/macrophage cells were defined as CD45+CD11b+F4/80+ cells. M-MDSC cells were defined as CD45+CD11b+Ly6C^{high}Ly6G- cells. G-MDSC cells were defined as CD45+CD11b+Ly6C^{low}Ly6G+ cells. Granulocytes were defined as CD45+CD11b+Ly6G+ cells.

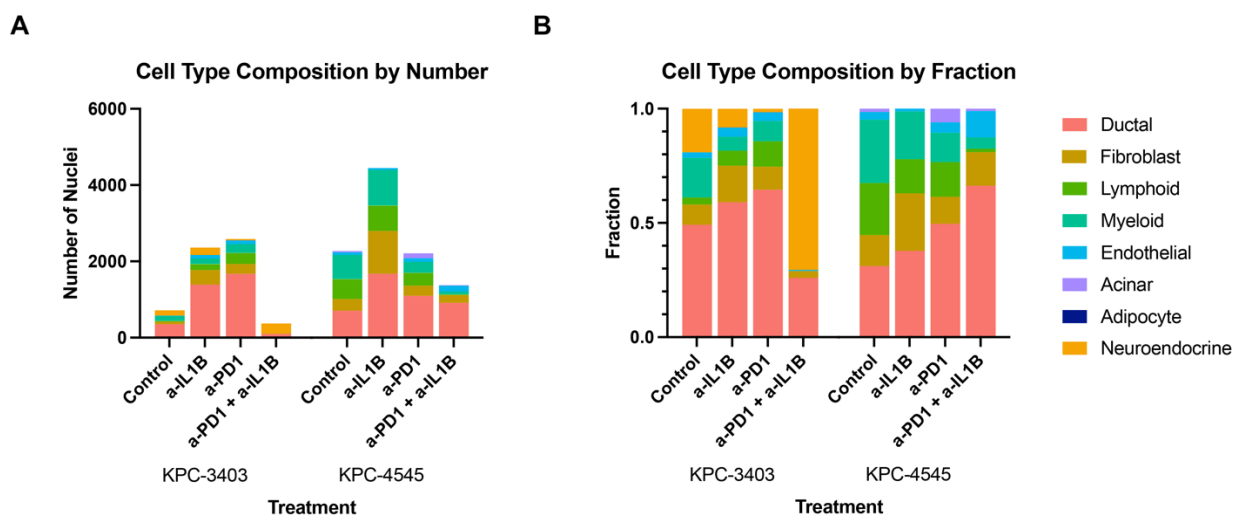
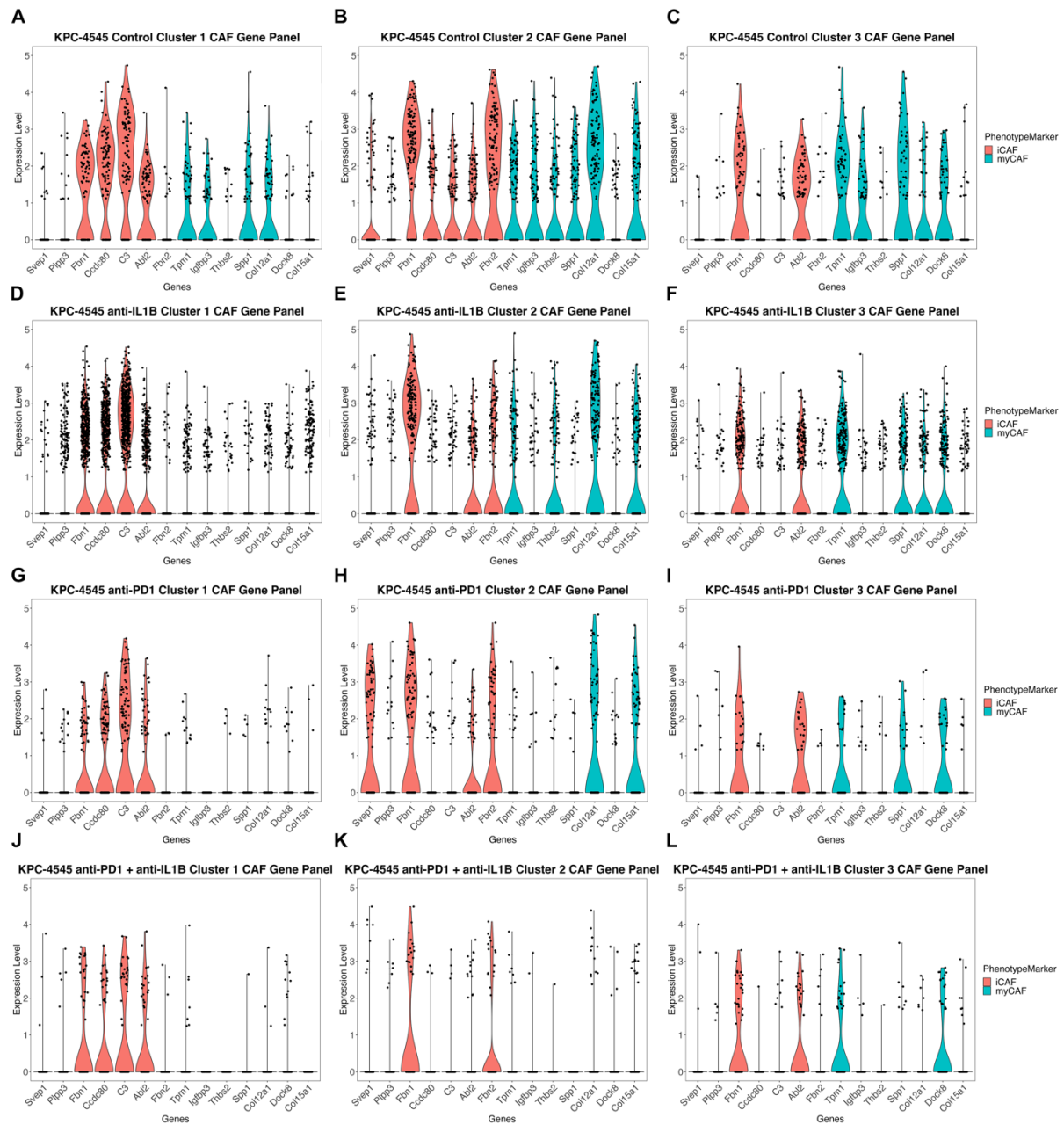


Figure S6. Cell Type Distribution in the KPC-4545 and KPC-3403 Model.

(A-B) Distribution of each cell type for each individual sample by cell count **(A)** and fraction of total nuclei **(B)** in each sample.



54

55 **Figure S7. Anti-IL-1 β antibody does not significantly modulate CAF phenotype across**
 56 **different CAF clusters in the KPC-4545 model.**

57 **(A-L)** CAF phenotype gene panel with iCAF (red) and myCAF (blue) signature genes across the
58 three major KPC-4545 CAF clusters of the control **(A-C)**, a-IL-1 β treated **(D-F)**, a-PD-1 treated
59 **(G-I)**, and combination treated samples **(J-L)**.

60

61

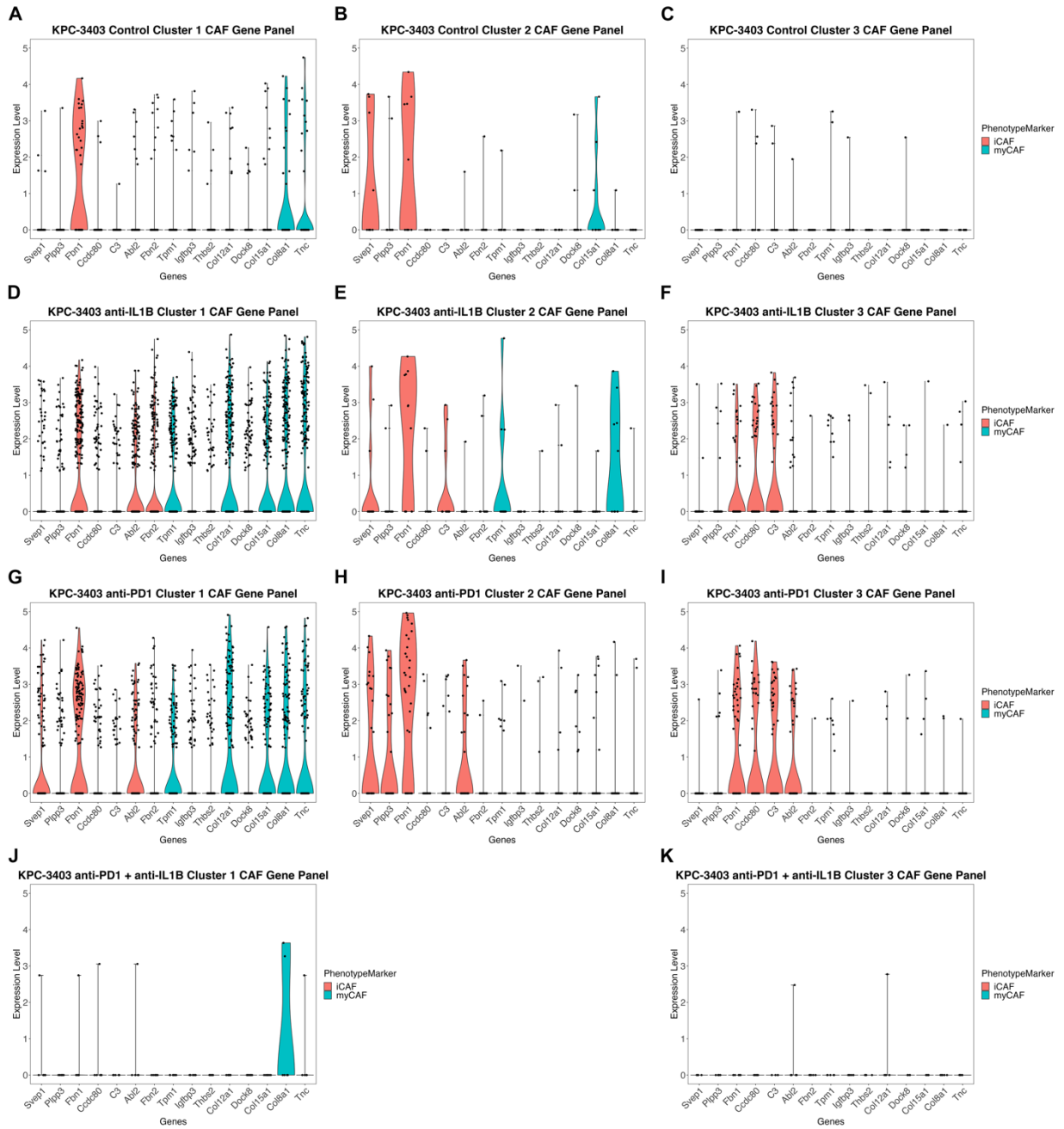


Figure S8. Anti-IL-1 β antibody does not modulate CAF phenotype across different CAF clusters in the KPC-3403 model.

(A-K) CAF phenotype gene panel with iCAF (red) and myCAF (blue) signature genes across the three major KPC-3403 CAF clusters in the control (A-C), a-IL-1 β -treated (D-F) a-PD-1-treated

67 (G-I), and combination-treated (J-K) samples. No cells were observed in cluster 2 for the combo
68 treated sample.

69

70

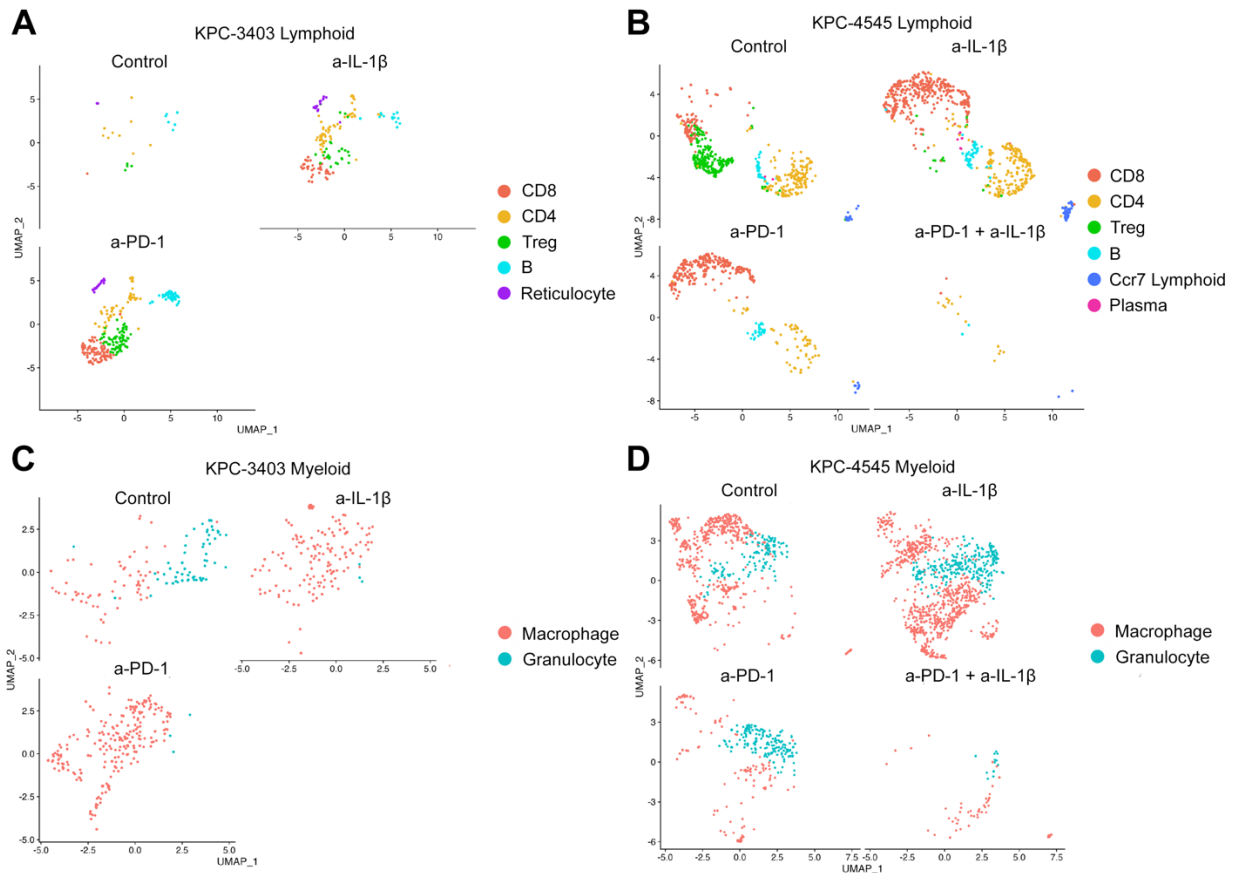


Figure S9. Lymphoid and myeloid landscape of the KPC-3403 and KPC-4545 tumor model following treatment with anti-PD-1 and anti-IL-1 β .

(A&B) Re-clustered uniform manifold approximation and projection (UMAP) embeddings of annotated lymphoid cells across the four a-IL-1 β samples in the KPC-3403 **(A)** and the KPC-4545 model **(B)** stratified by treatment. **(C&D)** Re-clustered UMAP embeddings of annotated myeloid cells across the four a-IL-1 β samples in the KPC-3403 **(C)** and the KPC-4545 model **(D)** stratified by treatment.

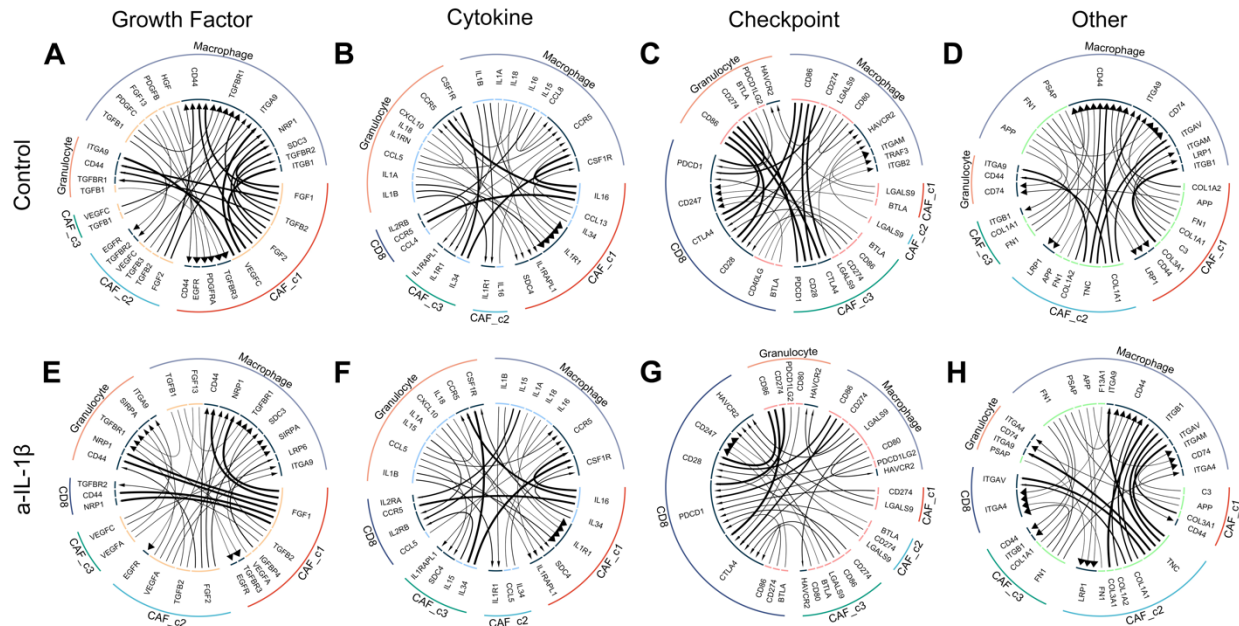


Figure S10. Ligand receptor analysis reveals complex ligand-receptor interactions between CAFs and immune cells in the KPC-4545 model.

Chord diagram of ligand-receptor interactions between all three major clusters of CAFs, CD8 T cells, macrophages, and granulocytes. Each row represents a treatment sample in the order of control (A-D) and a-IL-1 β (E-H) from top to bottom. Each column represents a category defined by iTALK, including growth factor, cytokines, checkpoint, and other going from left to right.

SUPPLEMENTAL TABLE DESCRIPTIONS

Supplementary Table 1 (Table.S1): Top 30 Ligand Receptor Interactions for Control

versus anti-IL-1 β Treated Samples in KPC-4545 Liver Metastasis Tropism for

Comparison. Separated into four different ligand receptor interactions: growth factor, cytokines, checkpoint, and other.

Supplementary Table 2 (Table.S2): Top 30 Ligand Receptor Interactions for Control

versus anti-IL-1 β Treated Samples in KPC-3403 Lung Metastasis Tropism for

Comparison. Separated into four different ligand receptor interactions: growth factor, cytokines, checkpoint, and other.

Supplementary Table 3 (Table.S3): Top 30 Ligand Receptor Interactions for Control

versus anti-PD-1 Treated Samples in KPC-3403 Lung Metastasis Tropism for Comparison.

Separated into four different ligand receptor interactions: growth factor, cytokines, checkpoint, and other.

Supplementary Table 4 (Table.S4): Top 30 Ligand Receptor Interactions for Control

versus anti-PD-1 Treated Samples in KPC-4545 Liver Metastasis Tropism for Comparison.

Separated into four different ligand receptor interactions: growth factor, cytokines, checkpoint, and other.

Supplementary Table 5 (Table.S5): Top 30 Ligand Receptor Interactions for anti-PD-1

versus Combination Treated Samples in KPC-4545 Liver Metastasis Tropism for

114 **Comparison.** Separated into four different ligand receptor interactions: growth factor, cytokines,
115 checkpoint, and other.
116