

Expanded View Figures

Figure EV1. Expression of Elongator subunits and Ctu1/2 by M1 polarization signals.

- A, B M1 polarization signals downregulate the expression of tRNA-modifying enzymes at both mRNA (A) and protein (B) levels. Peritoneal macrophages were untreated or stimulated with IFN γ (50 ng/ml), LPS (100 ng/ml) or with both IFN γ and LPS *ex-vivo* for the indicated periods of time and mRNA levels of the indicated candidates were assessed in all experimental conditions by Real-Time PCR (A). mRNA levels in unstimulated cells were set to 1 and levels in other experimental conditions were relative to that after normalization with Gapdh mRNA levels ($n = 3$ mice; mean $\pm$ SD, Student *t*-test, * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$). Protein levels (B) were assessed by western blot analyses using the indicated antibodies.
- C BMDMs and intestinal epithelial cells (IECs) lacking *Elp3* show decreased thiolated tRNA levels. Northern blots were carried out with enriched small RNAs extracted from BMDMs of *Elp3*^{Control} and *Elp3* ^{Δ Mye} mice or with total RNAs from IECs of *Elp3*^{Control} and *Elp3* ^{Δ IEC} mice, using the indicated probes. Values are from three independent measurements (mean $\pm$ SD, **** $P < 0.0001$, Welch one-way Anova test).
- D *Elp3* expression in myeloid cells is dispensable in the architecture of intestinal crypts. Colons from both *Elp3*^{Control} and *Elp3* ^{Δ Mye} mice were subjected to IHC analyses.
- E Colon length is not altered upon *Elp3* deficiency in myeloid cells. Data at day 6 from 8 mice per group are illustrated (mean $\pm$ SD, Student *t*-test, nonsignificant).
- F mRNA levels of pro-inflammatory cytokines in peritoneal macrophages lacking *Elp3* do not change in unstimulated mice. mRNAs of the listed pro-inflammatory cytokines in peritoneal macrophage from *Elp3*^{Control} or *Elp3* ^{Δ Mye} mice were assessed by Real Time PCR. mRNA levels in *Elp3*^{Control} mice were set to 1 and levels in other experimental conditions were relative to that after normalization with Gapdh mRNA levels ($n = 5$ mice; mean $\pm$ SD, nonsignificant).

Source data are available online for this figure.

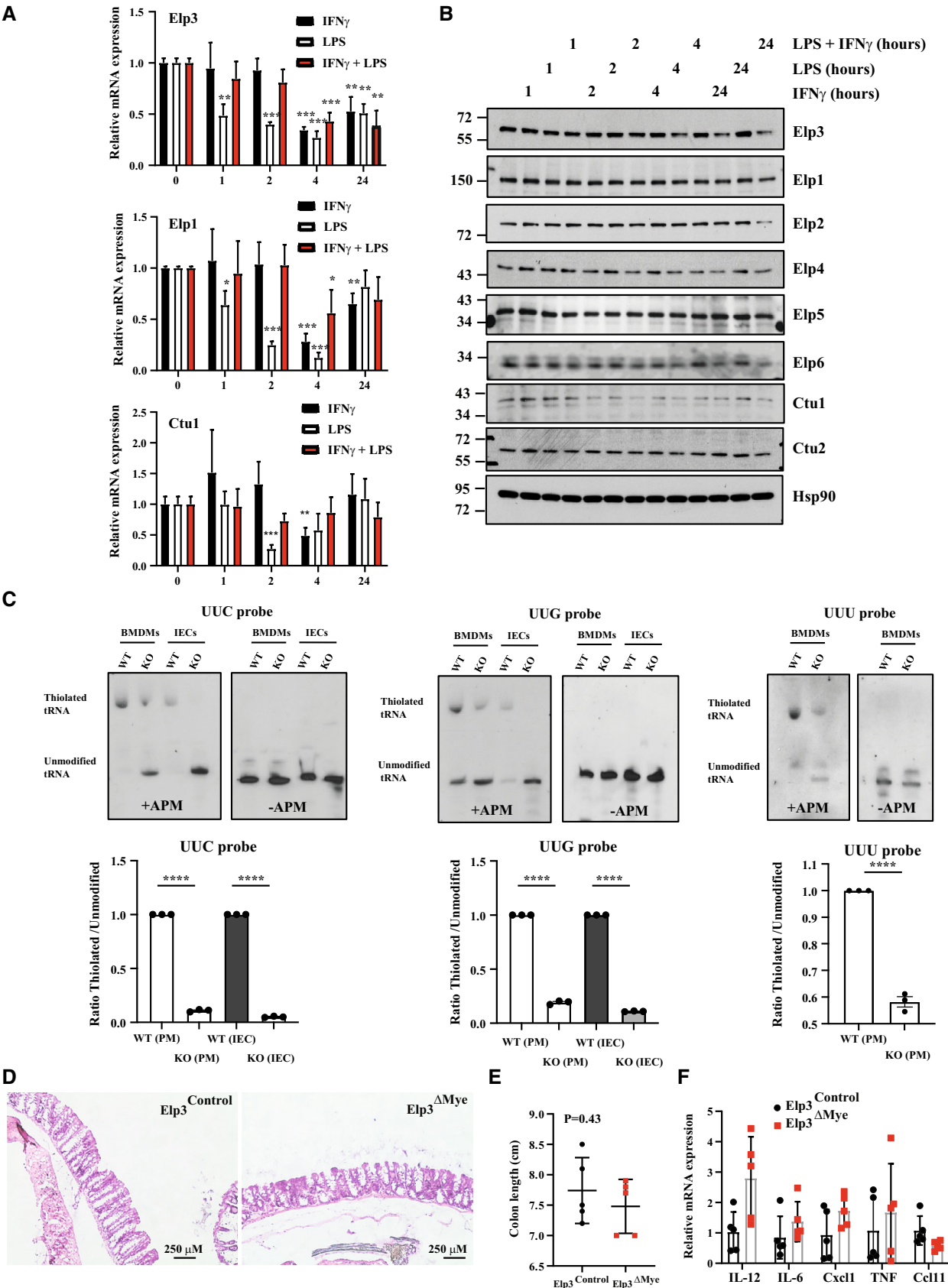


Figure EV1.

Figure EV2. Ctu2 promotes IL-4-dependent mTORC2 activation and M2 macrophage polarization.

- A Elp1/3 and Ctu1/2 expression are induced by M2 polarization signals. Peritoneal macrophages were untreated or stimulated with IL-4 (20 ng/ml), IL-13 (20 ng/ml) or with both IL-4 and IL-13 *ex-vivo* for the indicated periods of time and mRNA levels of the indicated candidates were assessed in all experimental conditions by Real-Time PCR. mRNA levels in unstimulated cells were set to 1 and levels in other experimental conditions were relative to that after normalization with Gapdh mRNA levels ($n = 3$ mice; mean $\pm$ SD, Student *t*-test, * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$).
- B IL-4-dependent mTORC2 activation relies on Ctu2. Control and Ctu2-depleted bone marrow-derived macrophages (BMDMs) were treated or not with IL-4 (10 ng/ml) and the resulting cell extracts were subjected to western blot (WB) analyses.
- C Ctu2 deficiency impairs M2 macrophage polarization. Control and Ctu2-depleted BMDMs were treated or not with IL-4 (10 ng/ml) for 24 h and the resulting mRNAs of the listed M2 markers were assessed by Real Time PCR. mRNA levels in control BMDMs were set to 1 and levels in other experimental conditions were relative to that after normalization with Gapdh mRNA levels ($n = 3$ mice; mean $\pm$ SD, Student *t*-test, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$).

Source data are available online for this figure.

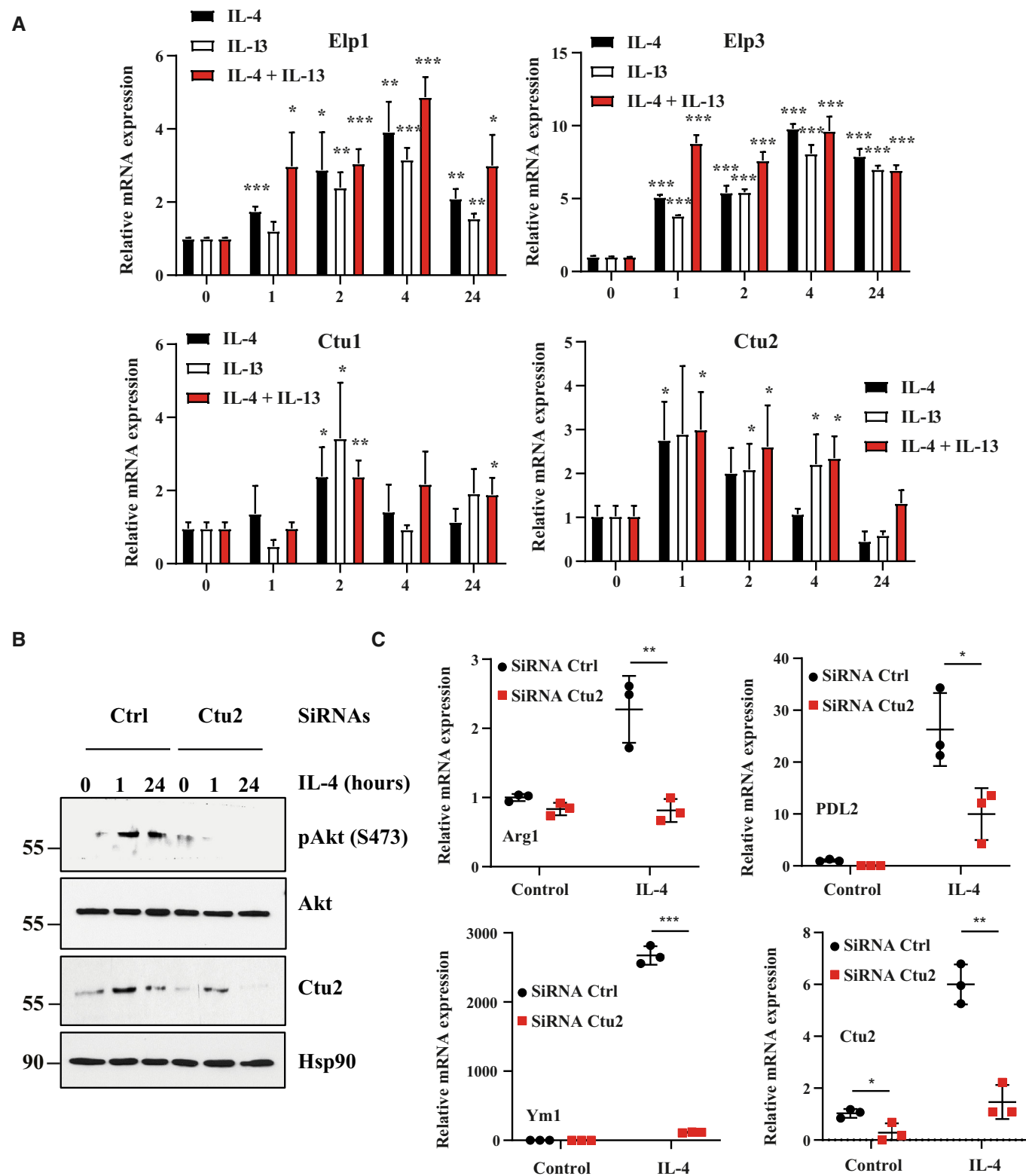


Figure EV2.

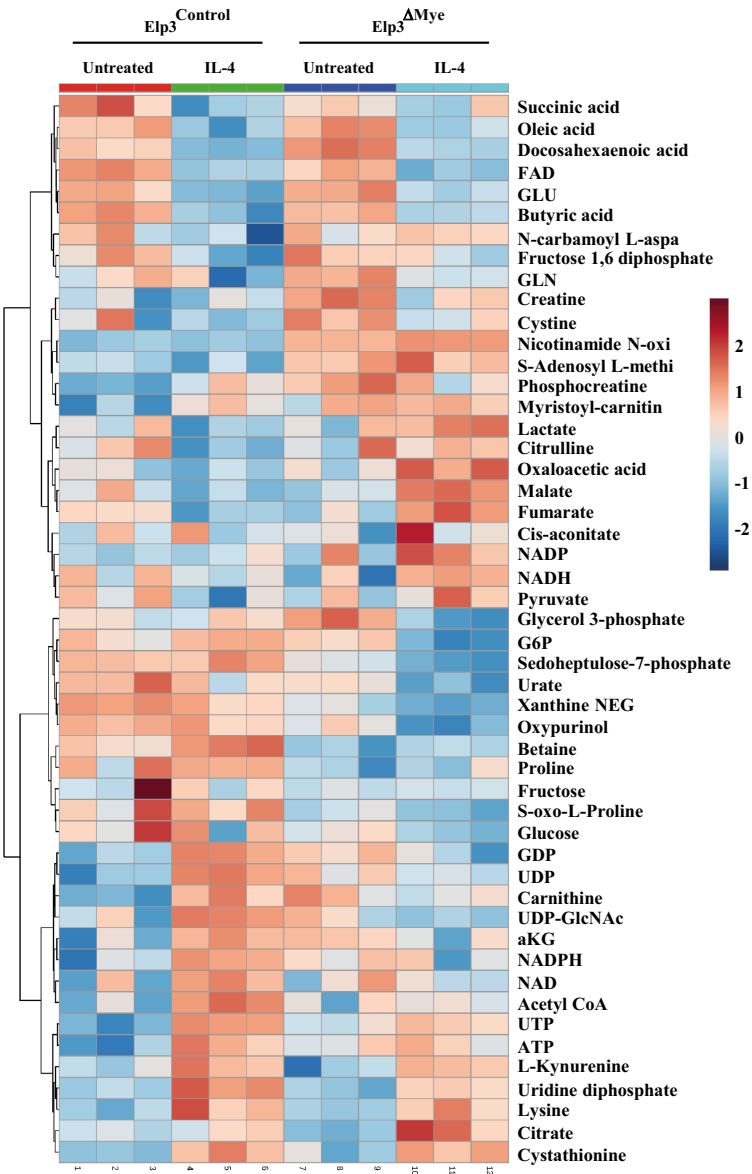


Figure EV3. Metabolic reprogramming by IL-4 relies on Elp3.
Peritoneal macrophages from *Elp3*^{Control} and *Elp3*^{ΔMye} mice (*n* = 3 per genotype) were treated with IL-4 for 24 h *ex-vivo* and the resulting extracts were subjected to Mass Spec analyses to extensively quantify metabolites. At the bottom, defective TCA cycle upon *Elp3* deficiency in IL-4-treated macrophages. Red arrows identify TCA metabolites whose levels change upon *Elp3* deficiency.

Figure EV4. Elp3 promotes Ric8b expression.

- A Elp3 does not regulate Ric8b mRNA levels. mRNAs of BMDMs from the indicated genotypes and stimulated *ex-vivo* with IL-4 or with PBS (control) were subjected to Real-Time PCRs. mRNA levels of Ric8b in BMDMs treated with PBS from *Elp3*^{Control} mice were set to 1 and levels in other experimental conditions were relative to that after normalization with Gapdh mRNA levels ($n = 3$ mice per genotype (mean $\pm$ SD)).
- B Ric8b promotes LPS-dependent mTORC2 activation. Control or Ric8b-depleted BMDMs were treated or not with IFN γ (50 ng/ml) and LPS (100 ng/ml) for the indicated periods of time and the resulting cell extracts were subjected to WB analyses.
- C Increased aggregate formation by IL-4 in macrophages. Flow cytometry of aggregates with extracts of BMDMs from *Elp3*^{Control} and *Elp3*^{AMye} mice treated with IL-4 (10 ng/ml) for 24 h are illustrated. Experiments were conducted in duplicates.
- D Visualization of protein aggregates. Coomassie blue staining of aggregate samples from *Elp3*^{Control} and *Elp3*^{AMye} BMDMs treated with IL-4 or PBS (control; $n = 8$ mice).
- E Identification of a proteomic signature of candidates whose expression is increased upon IL-4 stimulation and in BMDMs lacking Elp3 (Venn diagram).
- F Identification of proteins enriched in aggregates in macrophages lacking Elp3. Dot plot of protein aggregates from IL-4-treated BMDMs cells lacking or not *Elp3* are shown. Genes enriched in Lys^{AAA}, Gln^{CAA}, and Glu^{CAA} codons are shown in blue. Mrpls are shown in Red.
- G The expression of selected Mrpl proteins is controlled by Elp3 in macrophages. Peritoneal macrophages from *Elp3*^{Control} and *Elp3*^{AMye} mice were treated or not with IL-4/IL-13 (10 ng/ml) for the indicated periods of time and cell extracts from resulting cells were subjected to WBs using the indicated antibodies.
- H Elp3 does not regulate mRNA levels of Mrpl proteins. PMs from the indicated genotypes were stimulated or not with IL-4/IL-13 (10 ng/ml) for 24 h and mRNA levels of the indicated candidates were quantified by Real-Time PCR. mRNA levels of all candidates in untreated cells from *Elp3*^{Control} mice were set to 1 and levels in other experimental conditions were relative to that after normalization with Gapdh mRNA levels ($n = 3$ mice, mean $\pm$ SD, Student *t*-test, nonsignificant).
- I Elp3 controls Mt-co3 expression in macrophages. Protein extracts from PMs of the indicated genotypes treated or not with IL-4/IL-13 (10 ng/ml) for the indicated periods of time were subjected to WB analyses.

Source data are available online for this figure.

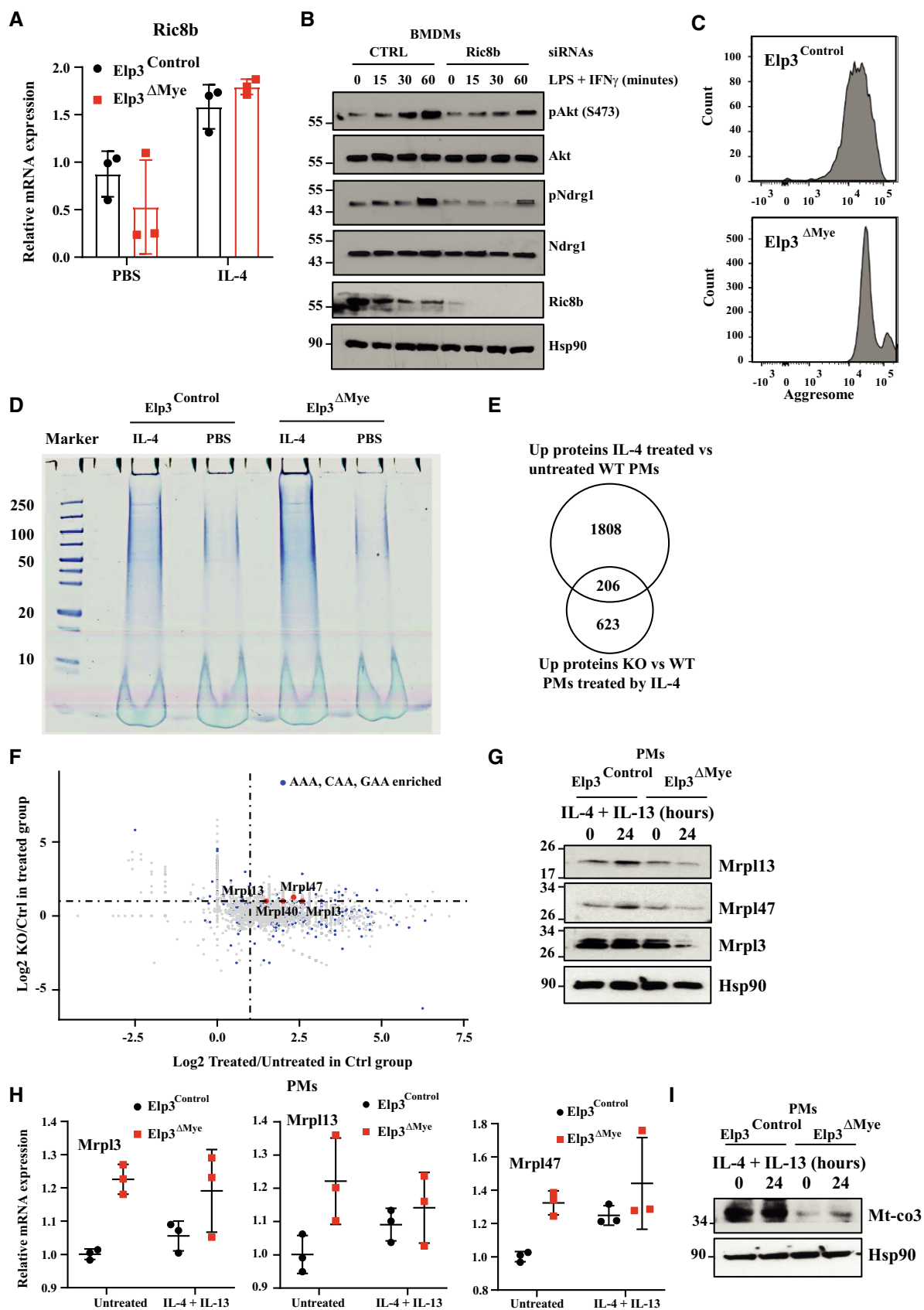


Figure EV4.

Figure EV5. Elp3 deficiency in peritoneal macrophages does not stabilize Atf4.

- A Peritoneal macrophages from the indicated genotypes were stimulated or not with IL-4/IL-13 (10 ng/ml) for the indicated periods of time and the resulting extracts were subjected to WB analyses.
- B *Elp3* deficiency does not potentiate Atf4-driven transcription in myeloid cells. Peritoneal macrophages from the indicated genotypes were stimulated or not with IL-4 (10 ng/ml) for 24 h and mRNA levels of the indicated Atf4 target genes were quantified by Real-Time PCR. mRNA levels of all candidates in untreated cells from *Elp3*^{Control} mice were set to 1 and levels in other experimental conditions were relative to that after normalization with Gapdh mRNA levels ($n = 4$ mice, mean $\pm$ SD, Student *t*-test, nonsignificant).

Source data are available online for this figure.

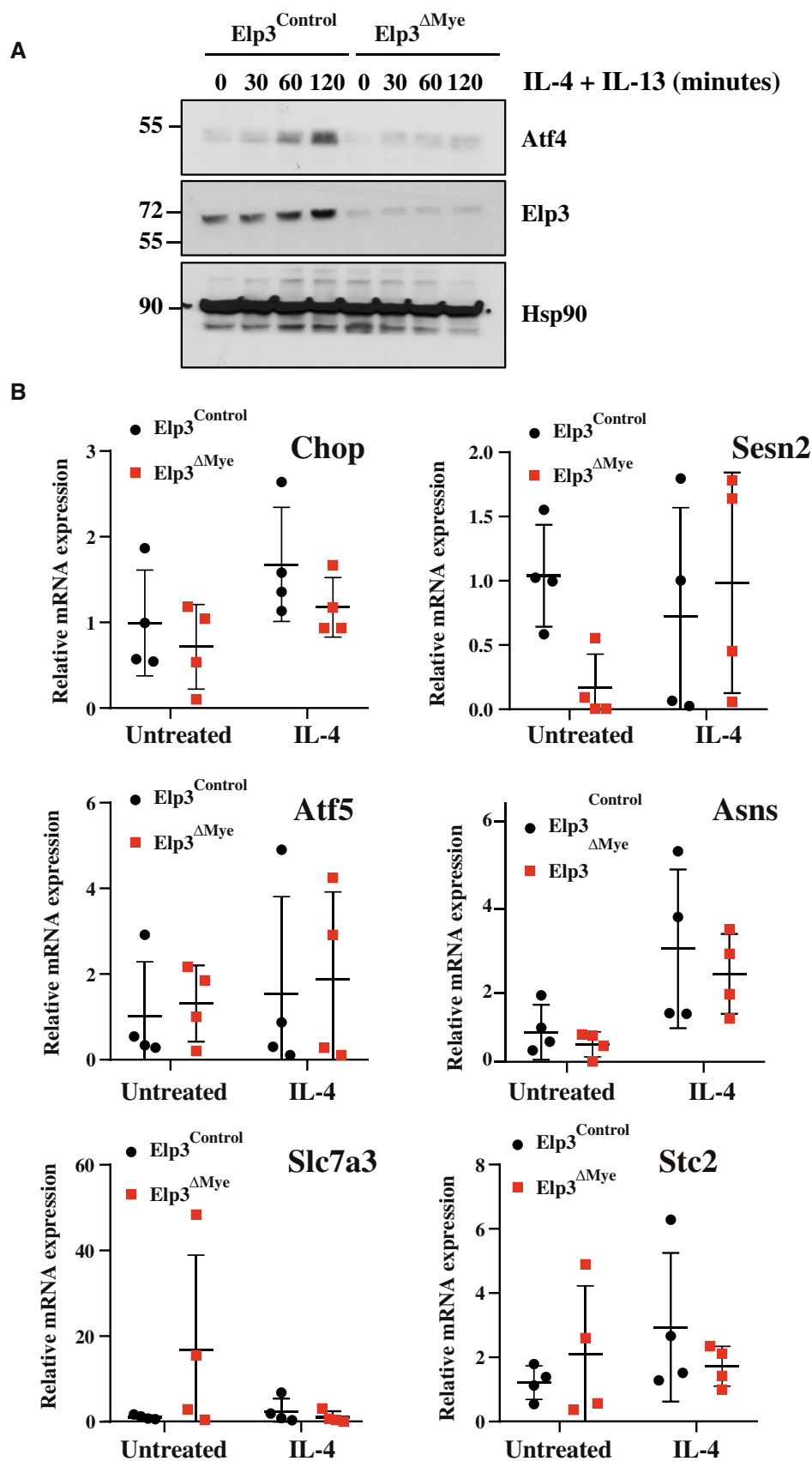


Figure EV5.