

Therapeutic application of PPE2 protein of *Mycobacterium tuberculosis* in inhibiting tissue inflammation

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Table S1: List of oligonucleotides used in the study**Real-time PCR (qPCR)**

TNF-α	Forward primer- 5' GGTGCCTATGTCTCAGCCTCTT 3' Reverse Primer- 5' GCCATAGAAGTATGAGAGGGAG 3'
IL-6	Forward primer- 5' TACCACTTCACAAGTCGGAGGC 3' Reverse Primer- 5' CTGCAAGTGCATCATCGTTGTTC 3'
MCP-3	Forward primer- 5' CAGAAGGATCACCAGTAGTCGG 3' Reverse Primer- 5' ATAGCCTCCTCGACCCACTTCT 3'
Mcpt-4	Forward primer- 5' CGACTATAACCTCCAGGTCTGC 3' Reverse Primer- 5' GAGGAGATTTCGGGTGAAGACTG 3'
SCF	Forward primer- 5' TCCAAGAAAATCGCTCCGGG 3' Reverse Primer - 5' GCAAGTGATAATCCAAGTCGGG 3'
GAPDH	Forward primer- 5' CATCACTGCCACCCAGAAGACTG 3' Reverse Primer- 5' ATGCCAGTGAGCTTCCCGTTCAG 3'

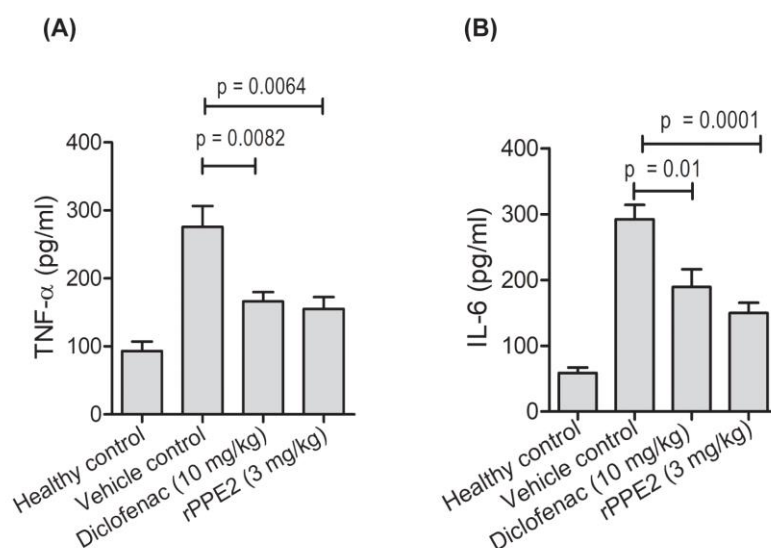
Molecular Cloning

SCF-600 bp promoter	Forward Primer- 5' CGCGTGTAGGGGAAAAGAACCAAGTGAAGTCTATCCGAC 3' Reverse Primer- 5' ATCTCGAGTCTTCTAAGGAAAGGCAGCGCTGCGATC 3'
SCF-400 bp promoter	Forward primer- 5' ATACGCGTAGACAGGGGACGCACCAGGCTCGAT 3' Reverse Primer- 5' ATCTCGAGTCTTCATAAGGAAAGGCAGCGCTGCGAT 3'
SCF-200 bp promoter	Forward Primer- 5' ATACGCGTGCCCTTCAGCGCGCTCCCGG 3' Reverse Primer- 5' ATCTCGAGCGGCTGCAGATAGTCCCAGCATTGGGT 3'

EMSA oligonucleotide

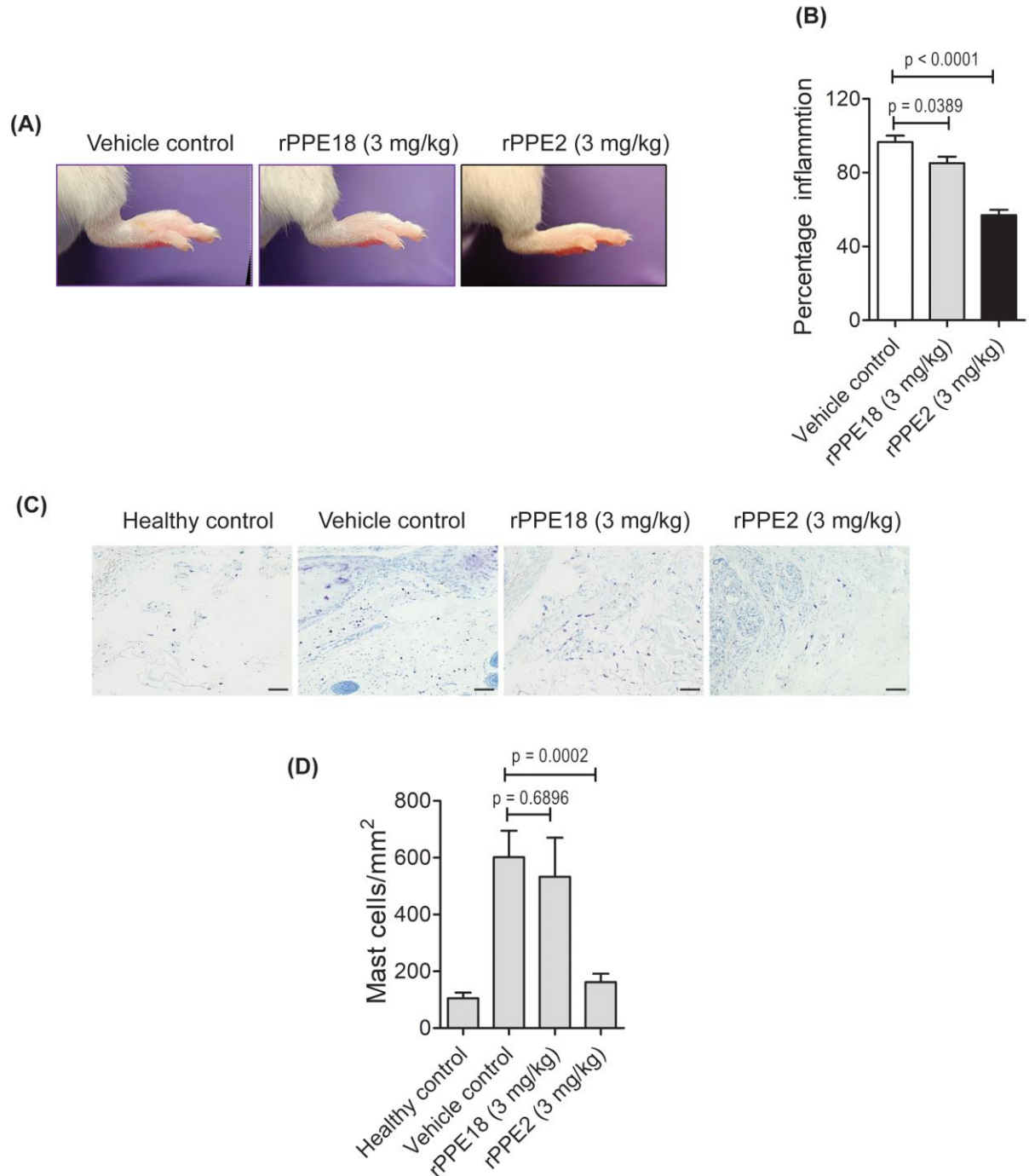
EPD predicted scf promoter	5'CGCGTGCGGGCGGGAGGGAGCTGTATAAAAAGCGCTGGCGGCGCAGCAGC CGGGGCTTCATTTGCG 3'
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Appendix Figure S1



Appendix Figure S1. Serum levels of TNF- α and IL-6 cytokines are reduced in mice treated with rPPE2. A subplantar injection of 5% of formalin (20 μ l) was administered in the right hind paw of Balb/c mice and an equal volume of PBS was injected in the left hind paw. After 1 hour of development of edema, mice were administered intraperitoneally with a single dose of either Diclofenac (10 mg/kg) or rPPE2 (3 mg/kg) or PBS (vehicle control). After 3 hours of treatment, mice were sacrificed and the whole blood sera were collected. The levels of TNF- α (**A**) and IL-6 (**B**) cytokines in blood sera were measured by EIA. Data shown are Mean $\pm$ SEM of 8 mice per group. Unpaired *t*-test was applied to calculate *p* values.

Appendix Figure S2



Appendix Figure S2. rPPE18 is not as effective as rPPE2 to reduce formalin-induced paw inflammation. Recombinant PPE18 (rPPE18) was purified using metal affinity chromatography. Briefly, 6X-histidine-tagged *ppe18* cloned in pRSETa, was overexpressed in *E. coli* BL21 (DE3) cells under IPTG inducible promoter. After induction, bacterial cells were pelleted down, suspended and lysed in PBS (pH 7.4) containing 0.3 mg/ml lysozyme (Sigma-Aldrich) and 1 mM PMSF (Sigma-Aldrich) and 1% sodium lauryl sarcosine (Sigma-

Aldrich). Bacterial lysates were incubated with TALON (Clontech Laboratories, Mountain View, CA) beads. rPPE18-bound TALON beads were washed with 20 mM imidazole (Sigma-Aldrich) and then eluted with 200 mM imidazole and the purified protein was collected. **(A-D)** A subplantar injection of 5% formalin (20 μ l) was administered in the right hind paw of Balb/c mice and an equal volume of PBS was injected in the left hind paw. After 1 hour of development of edema, mice were administered intraperitoneally with a single dose of either rPPE18 (3 mg/kg) or rPPE2 (3 mg/kg) or PBS (vehicle control). After 3 hours of treatment, the percentage of inflammation was calculated. **(A)** Representative photographs of inflamed paws are shown after 3 hours of treatment. **(B)** Graphical representation of percentage inflammation in paw (paw thickness) was shown. **(C-D)** After three hours of treatment, mice were sacrificed and the paw sections were prepared and stained with Toluidine blue to check mast cell population (scale bar = 100 μ m). Photographs of representative sections were visualized at 20X magnification **(C)**. Counting of mast cells was performed in Toluidine blue stained paw sections using ImageJ software and was normalized per unit area (mm^2) **(D)**. Data shown are Mean $\pm$ SEM of 5 mice. Unpaired *t*-test was applied to calculate p values.